

# 182. Method to inhibit neutrophil recruitment to damaged tissue

(Wisconsin Alumni Research Foundation)

5<sup>TH</sup> KDDF GLOBAL  
C&D TECH FAIR

## ► Asset Overview

<b>Product Type</b>	Protein
<b>Disease Area</b>	Inflammatory Disease
<b>Indication</b>	Autoimmune Disorders
<b>Current Stage</b>	Lead Optimization
<b>Target</b>	The HIF-1a pathway
<b>MoA</b>	Myeloid-derived growth factor (MYDGF) is a damage signal that regulates neutrophil interstitial motility and inflammation through a HIF-1a pathway in response to tissue damage.
<b>Brief Description</b>	<ul style="list-style-type: none"><li>• Myeloid-derived growth factor (MYDGF) is known to mediate cardiac repair following myocardial infarction by inhibiting cardiac myocyte apoptosis. MYDGF (C19orf10) mediates cardiac repair following myocardial infarction. MYDGF-deficient mice developed larger infarct scars and more severe contractile dysfunction compared to wild-type mice. Treating the mice with recombinant MYDGF reduced scar size and contractile dysfunction after myocardial infarction.</li><li>• A method to inhibit neutrophil recruitment to damaged tissue in a subject, the method comprising administering to a subject, the subject having a wound and/or a burn at a site on the subject, an amount of a MYDGF, a biologically active fragment thereof, an analogous sequence thereof, or a pharmaceutically suitable salt of any of the foregoing, wherein the amount is effective to inhibit neutrophil recruitment to the wound and/or burn site.</li><li>• A method to inhibit inflammation in a subject, the method comprising administering to the subject an anti-inflammatory-effective amount of a MYDGF, a biologically active fragment thereof, an analogous sequence thereof, or a pharmaceutically suitable salt of any of the foregoing.</li></ul>
<b>Intellectual Property</b>	US20220168390A1
<b>Publication</b>	Myeloid-derived growth factor regulates neutrophil motility in interstitial tissue damage. J Cell Biol, (2021)
<b>Inventors</b>	Anna Huttenlocher, Deane Mosher, Valeriu Bortnov, David Bennis

## ► Highlights

- MYDGF regulates neutrophil responses to tissue damage, but not infection.
- MYDGF depletion leads to a neutrophil-dependent defect in wound healing.
- MYDGF depletion alters neutrophil motility in the wound microenvironment.
- MYDGF depletion impairs neutrophil reverse migration and resolution following injury.
- Neutrophil accumulation in the mydgm mutant is dependent on the HIF-1 $\alpha$  pathway.

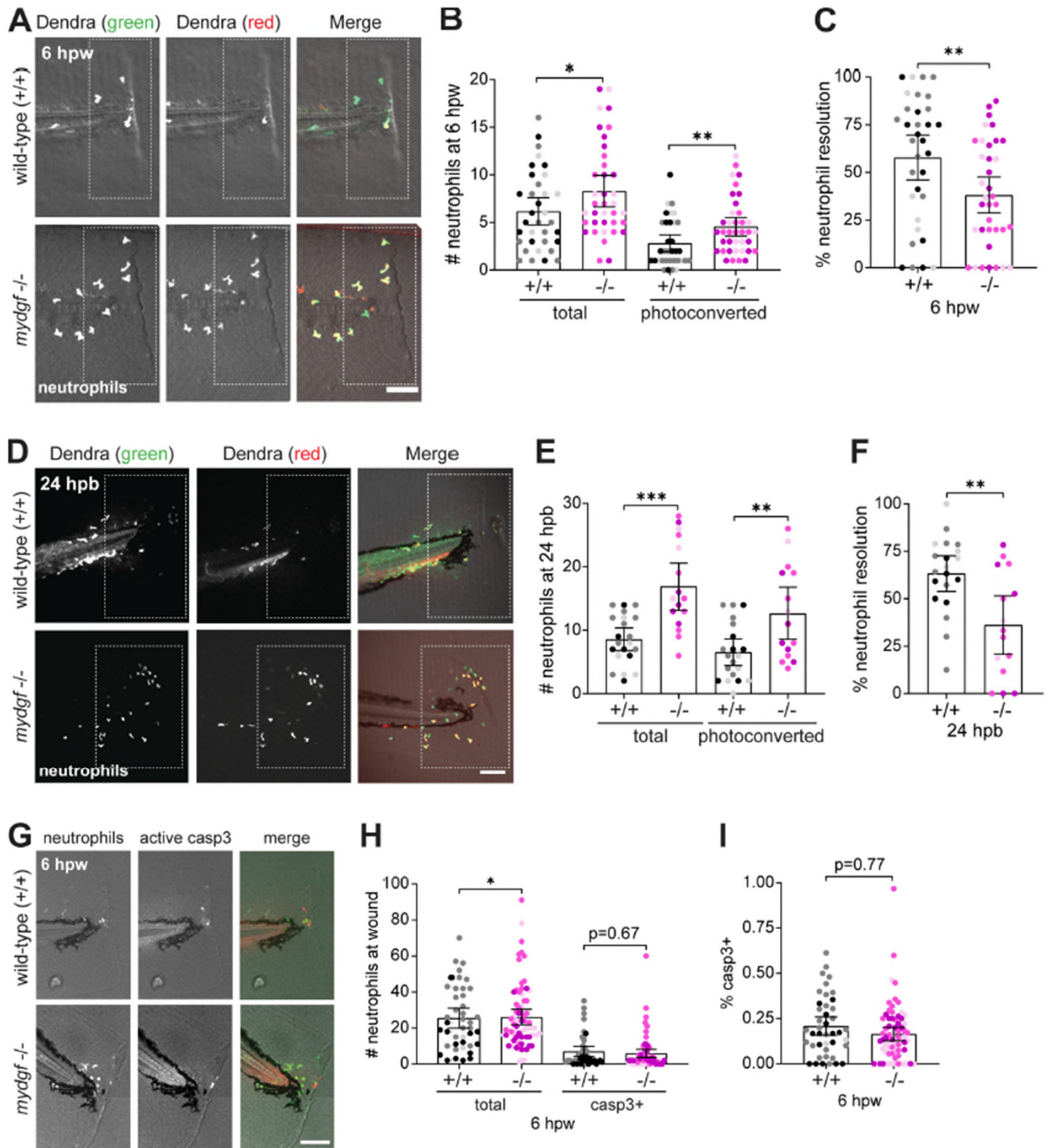
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## ► Key Data

MYDGF depletion impairs neutrophil reverse migration and resolution following injury.



To be continued

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## ► Key Data

### MYDGF depletion impairs neutrophil reverse migration and resolution following injury.

(A and B) Representative images (A) and quantification (B) of green (total) and red(photoconverted) dendra2-labeled neutrophils in the wound microenvironment at 6 hpw following tail transection of 3 dpf WT and *mydgm*<sup>-/-</sup> larvae; three independent replicates with n = 33 +/+ and 36 -/-; scale bar = 100  $\mu$ m.

(C) Quantification of the percentage of photoconverted neutrophils present at the wound at 2 hpw that are no longer present at 6 hpw in the same larva.

(D and E) Representative images (D) and quantification (E) of green (total) and red (photoconverted) dendra2-labeled neutrophils in the burn at 24 hpb following thermal injury of the caudal fin of 3 dpf WT and *mydgm*<sup>-/-</sup> larvae; three independent replicates with n = 20 +/+ and 15 -/-; scale bar = 100  $\mu$ m.

(F) Quantification of the percentage of photoconverted neutrophils present in the burn microenvironment at 3 hpb that are no longer present at 24 hpb in the same larva.

(G) Representative images of immunostaining for active caspase-3 (*caspm3*) following tail transection of 3 dpf WT and *mydgm*<sup>-/-</sup> larvae with mCherry-labeled neutrophils at 6 hpw; scale bar = 100  $\mu$ m.

(H) Quantification of total and active caspase-3-expressing neutrophils in the wound at 6 hpw; three independent replicates with n = 42 +/+ and 71 -/-.

(I) Proportion of neutrophils in the wound expressing active caspase-3 at 6 hpw. In B, C, E, F, H, and I, data are expressed as mean with 95% CI; each symbol represents one larva, and different colors represent the results of three independent replicates. \*, P < 0.05; \*\*, P < 0.01; \*\*\*, P < 0.001. P values were calculated by ANOVA with Tukey's multiple comparisons.

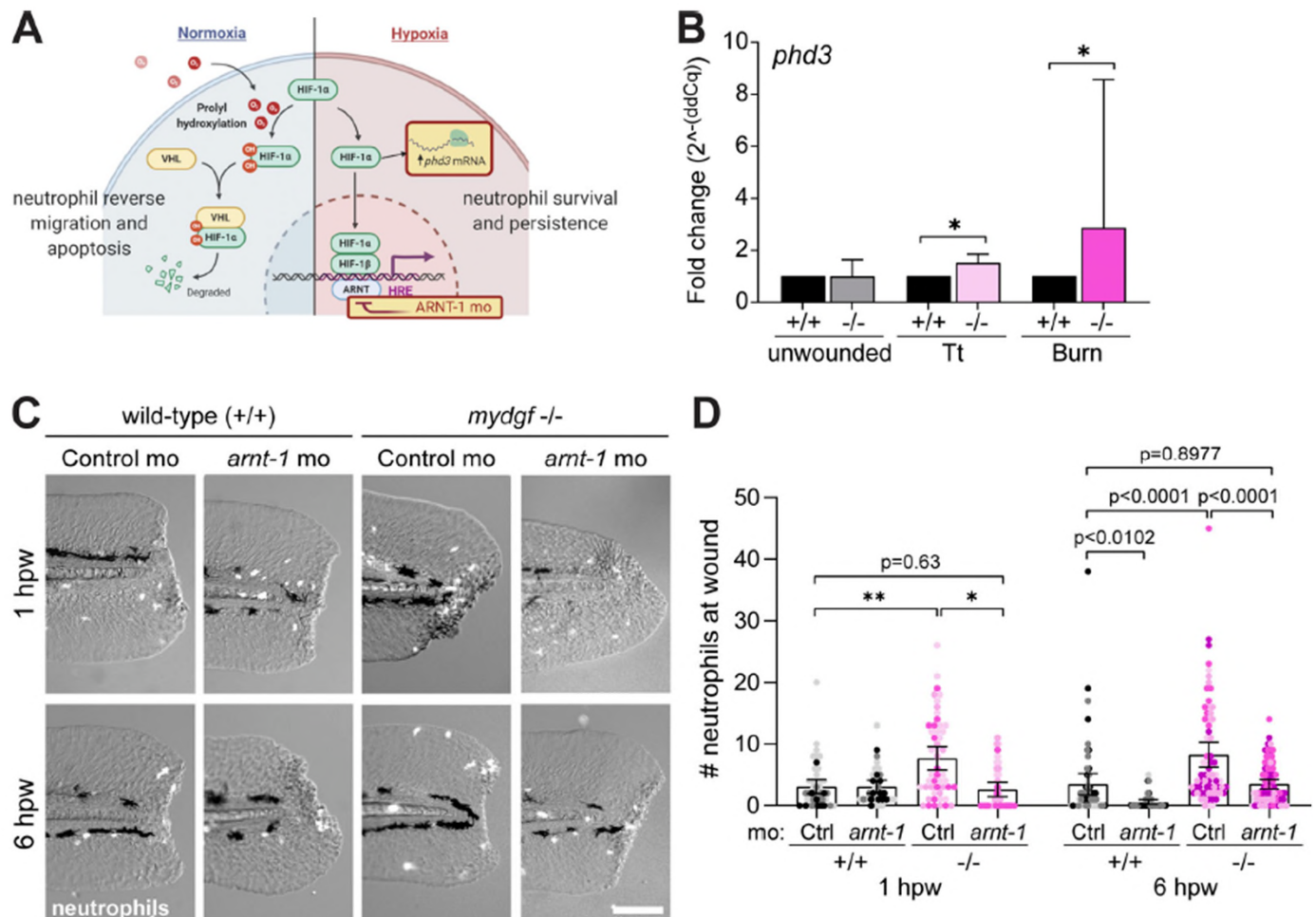
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## ► Key Data

### Neutrophil accumulation in the *mydgf* mutant is dependent on the HIF-1 $\alpha$ pathway.



(A) Schematic representation of HIF-1 $\alpha$  activation in neutrophils. Pathway activation results in neutrophil persistence and survival and is characterized by increased expression of *phd3*. HIF-1 $\alpha$  pathway activation can be blocked at the level of transcription factor nuclear binding using *arnt-1* morpholino. Illustration was created at [www.biorender.com](http://www.biorender.com). (B) *phd3* expression in pooled tail fin tissue collect from WT larvae, either unwounded or 3 h following tail transection (Tt) or thermal injury (burn), measured by RT-qPCR. Data comprise three (burn) to five (Tt) independent replicates performed in technical triplicates and normalized to *mydgf* expression in unwounded tails and to *ef1 $\alpha$* .  $n = 50$  tails per condition per independent replicate. \*,  $P < 0.05$ . Fold changes in gene expression were compared with the normalized value of 1 using one-sample t tests. (C and D) Representative images (C) and quantification (D) of the number of mCherry-labeled neutrophils at the wound in WT and *mydgf*<sup>-/-</sup> larvae, with control or *arnt-1* morpholino, at 1 and 6 hpw after Tt; three or four independent replicates with  $n = 46$  +/+ control mo, 33 +/+ *arnt-1* mo, 50 -/- control mo, and 38 -/- *arnt-1* mo at 1 hpw and 54 +/+ control mo, 38 +/+ *arnt-1* mo, 68 -/- control mo, and 73 -/- *arnt-1* mo at 6 hpw; scale bar = 100  $\mu$ m. Data are expressed as mean with 95% CI; each symbol represents one larva, and different colors represent independent replicates. \*,  $P < 0.05$ ; \*\*,  $P < 0.01$ . P values were calculated by ANOVA with Tukey's multiple comparisons.