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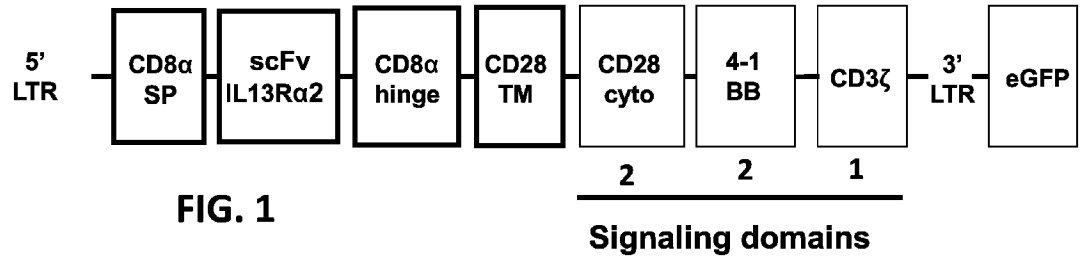


FIG. 1

(57) Abstract: Immunotherapies, particularly chimeric antigen receptors targeting IL-13Rα2 and their use for treating cancer are provided. A single chain fragment variable (scFv) that specifically binds IL-13Rα2, chimeric antigen receptors (CARs) including the IL-13Rα2 scFv, nucleic acids encoding the CARs, vectors including the nucleic acids encoding the CARs, and immune cells expressing the CARs are provided. Also provided are methods of treating a subject with cancer, including administering to the subject an immune cell expressing an IL-13Rα2 scFv-CAR alone or in combination with other cancer therapies.

WO 2022/212879 A1

IL-13RA2 CHIMERIC ANTIGEN RECEPTORS AND METHODS OF USE

CROSS REFERENCE TO RELATED APPLICATIONS

This claims the benefit of U.S. Provisional Application No. 63/169,575, filed April 1, 2021, which is incorporated by reference herein.

FIELD OF THE DISCLOSURE

This disclosure relates to immunotherapies, particularly chimeric antigen receptors targeting IL-13Rα2 and their use for treating cancer.

BACKGROUND

IL-13Rα2 is a high affinity receptor for cytokine interleukin-13 (IL-13) and a known tumor antigen. While the significance of IL-13Rα2 expression in cancer is not fully understood, it has been shown that IL-13Rα2 is overexpressed in a variety of human cancers, including malignant gliomas, head and neck cancer, Kaposi's sarcoma, renal cell carcinoma, ovarian carcinoma, breast cancer, and pancreatic cancer, making IL-13Rα2 a potential target for cancer immunotherapy.

Chimeric antigen receptor (CAR)-T cell therapy has become a promising immunotherapeutic strategy for the treatment of various blood cancers. While CAR-T cells are effective in hematological malignancies, such as leukemia and lymphoma, to date very limited activity has been seen in human solid cancers. Improving targeting of CAR-T cells in solid tumors, either alone or in combination with other agents targeting tumor stroma (the tumor microenvironment, including myeloid derived suppressor cells, tumor associated macrophages and fibroblasts), would signify a major advance in the field of CAR-T immunotherapy.

SUMMARY OF THE DISCLOSURE

IL-13Rα2 is significantly upregulated in several cancers, thus it is a target for immunotherapy for the treatment of a variety of cancers. Chimeric antigen receptors (CARs) specifically targeting cells expressing IL-13Rα2 are provided herein. These CARs can be used in immunotherapy to target cancers expressing or overexpressing IL-13Rα2.

Disclosed herein are single chain fragment variables (scFvs) that specifically bind IL-13Rα2. In some embodiments, the scFv has an amino acid sequence that includes the variable heavy chain (VH) domain complementarity determining region 1 (CDR1), CDR2 and CDR3 amino acid sequences of SEQ ID NO: 1 and the variable light chain (VL) domain CDR1, CDR2 and CDR3 amino acid sequences of SEQ ID NO: 1. In some examples, the scFv has at least 90%

identity to the amino acid sequence of SEQ ID NO: 1, or includes or consists of the amino acid sequence of SEQ ID NO: 1. In some examples, the scFv specifically binds to cells expressing IL-13Rα2.

Also provided are nucleic acids encoding the IL-13Rα2 scFv and vectors comprising the nucleic acids (such as a viral vector). In some embodiments, the nucleic acid encoding the IL-13Rα2 scFv is codon-optimized. In some examples, the IL-13Rα2 scFv is encoded by a nucleic acid sequence with at least 90% identity to the nucleic acid sequence of SEQ ID NO: 2. In other examples, the IL-13Rα2 scFv is encoded by a nucleic acid that includes or consists of the nucleic acid sequence of SEQ ID NO: 2.

Also provided are CARs that include the IL-13Rα2 scFv, a hinge domain, a transmembrane domain, and an intracellular domain with one or more signaling domains. In some embodiments, the CAR further comprises a signal peptide. In some examples, the signal peptide is a CD8α signal peptide. In some examples, the transmembrane domain is a CD28 transmembrane domain. In some examples, the intracellular domain includes one or more of CD28, 4-1BB, and CD3ζ signaling domains, or any combination of two or more thereof. In a non-limiting embodiment, the IL-13Rα2-CAR includes the IL-13Rα2 scFv, a CD8α hinge domain, a CD28 transmembrane domain, and an intracellular domain including a CD28 domain, a 4-1BB domain, and a CD3ζ domain. In some examples, the IL-13Rα2-CAR includes an amino acid sequence with at least 90% identity to the amino acid sequence of SEQ ID NO: 3, or includes or consists of the amino acid sequence of SEQ ID NO: 3.

Nucleic acids encoding the IL-13Rα2-CARs disclosed herein, and vectors including the nucleic acids (such as a viral vector) are also provided. In some embodiments, the nucleic acid encoding the IL-13Rα2-CAR is codon-optimized for expression in a particular cell type (*e.g.*, bacteria, yest, insect, mouse, human). In some examples, the IL-13Rα2-CAR is encoded by a nucleic acid sequence with at least 90% identity to the nucleic acid sequence of SEQ ID NO: 4. In other examples, the IL-13Rα2-CAR is encoded by a nucleic acid that includes or consists of the nucleic acid sequence of SEQ ID NO: 4. Vectors including a nucleic acid molecule encoding an IL-13Rα2-CAR are also disclosed herein. In some examples, the vector is a viral vector, such as a lentiviral vector.

Immune cells expressing the disclosed IL-13Rα2-CARs, such as immune cells including a nucleic acid encoding a disclosed IL-13Rα2-CAR, or a vector encoding a disclosed IL-13Rα2-CAR, are also provided. In some examples, the cells are T cells, natural killer (NK) cells, natural killer T (NKT) cells, or macrophages. In specific, non-limiting examples, the immune cells are T cells. In some examples, the immune cells are obtained from peripheral blood. Methods of

producing cells expressing the IL-13Rα2-CARs are provided. These methods include transducing or transfecting cells, such as T cells, NK cells, NKT cells, or macrophages, with a vector encoding a disclosed IL-13Rα2-CAR.

Also provided are methods for treating a subject with cancer, for example by administering a cell (such as a T cell, NK cell, NKT cell, or macrophage) expressing an IL-13Rα2-CAR, or including a vector encoding an IL-13Rα2-CAR. In some examples, the subject has a cancer that expresses or over-expresses IL-13Rα2. In particular non-limiting examples, the subject has pancreatic cancer, glioblastoma, head and neck squamous cell carcinoma, ovarian cancer, uterine cancer, prostate cancer, breast cancer, melanoma, non-small cell lung cancer (NSCLC), renal cell carcinoma, Kaposi sarcoma, or adrenal carcinoma. In some examples, the method further includes treating the subject with one or more of surgery, radiation, chemotherapy, or an additional immunotherapy. In further examples, a histone deacetylase (HDAC) inhibitor, a cell cycle and/or checkpoint inhibitor, adrenomedullin, an IL-13-PE immunotoxin, or any combination of two or more thereof, is administered to the subject in combination with IL-13Rα2-CAR immune cells.

The foregoing and other features of the disclosure will become more apparent from the following detailed description, which proceeds with reference to the accompanying figures.

BRIEF DESCRIPTION OF THE DRAWINGS

FIG. 1 shows a schematic diagram of an exemplary scFv-IL-13Rα2 CAR construct. The construct contains a 5'-CD8α signal peptide, scFv IL-13Rα2, CD8α hinge domain, CD28 transmembrane domain, and endodomains from CD28 cytoplasmic domain, 4-1BB and CD3ζ.

FIG. 2 shows amino acid numbering of heavy chain residues of the IL-13Rα2 antibody using the Kabat numbering scheme. The sequence shown is residues 22-139 of SEQ ID NO: 3.

FIGS. 3A-3B show analyses of the heavy chain of the IL-13Rα2 antibody. FIG. 3A is a table providing the region, sequence, residue number and length of heavy chain amino acids of scFv-IL-13Rα2. FIG. 3B is a loop structure of heavy chain CDRs with amino acid residue position number. The sequence shown is residues 22-139 of SEQ ID NO: 3.

FIG. 4 shows amino acid numbering of light chain residues of the IL-13Rα2 antibody using the Kabat numbering scheme. The sequence shown is residues 155-263 of SEQ ID NO: 3.

FIGS. 5A-5B show analyses of the light chain of the IL-13Rα2 antibody. FIG. 5A is a table providing the region, sequence, residue number and length of light chain amino acids of scFv-IL-13Rα2. FIG. 5B is a loop structure of light chain CDRs with amino acid residue position number. The sequence shown is residues 155-260 of SEQ ID NO: 3.

FIG. 6 shows the distribution of heavy chain amino acids of the IL-13Rα2 antibody and

their respective frequency.

FIG. 7 shows the distribution of light chain amino acids of the IL-13Rα2 antibody and their respective frequency.

FIG. 8 is a comparative sequence alignment of the heavy chain residues of the IL-13Rα2 antibody according to IMGT, Kabat, Chothia and Martin numbering schemes (SEQ ID NO: 3, residues 22-139).

FIG. 9 is a comparative sequence alignment of the light chain residues of the IL-13Rα2 antibody according to IMGT, Kabat, Chothia and Martin numbering schemes (SEQ ID NO: 3, residues 155-260).

FIG. 10 shows the amino acid sequence of an exemplary scFv IL-13Rα2 CAR and location of each element (SEQ ID NO: 18).

FIG. 11 shows a restriction digest of a vector containing a scFv IL-13Rα2 CAR. Approximately 1.0 μg plasmid DNA was digested with BamH1 and Not1 at 37°C for 1 hour and electrophoresed in 1% agarose gel then stained. Lane 1 is undigested scFv-IL-13Rα2 CAR construct plasmid, lane 2 is the plasmid DNA digested with BamH1 and Not1, and lane 3 is a KB reference ladder.

FIGS. 12A-12B show the expression of CD28 (cytoplasmic) (FIG. 12A) and CD3ζ (FIG. 12B) signaling domains in transduced CAR-Jurkat cells. FACS analysis of the CAR-Jurkat cells was performed after permeabilizing transduced Jurkat cells and immunostaining with anti-CD28 (CD28.2) mouse mAb PE conjugate (Cat#27826, Cell Signaling Technology, Danvers, MA) and CD3ζ monoclonal antibody PE conjugate (Cat# 12-2479-82, ThermoFisher eBioscience, Carlsbad, CA). The data are expressed as normalized to mode values.

FIGS. 13A-13B show the identification of the scFv IL-13Rα2 CAR transgene in transduced Jurkat cells and T cells by indirect immunofluorescence assay (IFA) analysis. Transduced Jurkat cells and T cells were incubated with 500 ng/ml biotinylated recombinant human IL-13Rα2Fc chimeric protein followed by streptavidin-Alexa 594 to develop fluorescence in scFv-IL-13Rα2 expressing cells. The cells expressing ≥ 2+ fluorescence intensity were counted at 200X magnification by viewing in NIKON epifluorescence microscope. Each value is mean ± SD of quadruple experiments determined in a blinded manner and expressed as % positive cells (FIG. 13A). FIG. 13B shows a representative IFA image of transduced Jurkat cells.

FIG. 14 shows cell viability of scFv IL-13Rα2 CAR-Jurkat and scFv IL-13Rα2 CAR-T cells in cell culture over the course of seven days. Viability was determined by trypan blue exclusion and is expressed as number of viable cells.

FIG. 15 shows cell proliferation of scFv IL-13Rα2 CAR-T cells *in vitro* by MTS assay. Transduced Jurkat cells and T cells were monitored for cell proliferation up to 7 days in appropriate culture medium. The proliferative activity of transduced cells was assessed by measuring the optical density of reduced MTS tetrazolium by proliferating cells at 490nm at specified time points. Each value is a mean ± SD of four independent experiments

FIG. 16 shows expression of T cell activation markers in CAR-T cells. Expression of T cell activation markers (CD25, CD44, CD69) and intracellular IFN-γ expression was measured on day 7.

FIG. 17 shows scFv IL-13Rα2 CAR-T cell specificity and cytotoxicity to IL-13Rα2+ glioma tumor cells in co-culture. After 6 hour co-culture with a fixed number of calcein-violet loaded tumor cells (5000) as target cells at various effector to target (E:T) ratios, release of fluorescent dye was measured to determine cell lytic activity of the CAR-T cells. IL-13Rα2 knock-out (KO) tumor cells were used as negative controls. Each value is a mean ± SD of three independent experiments performed in quadruplicate.

FIG. 18 shows siRNA silencing of IL-13Rα2 in U251 and U87MG glioma cell lines. Relative fluorescence units of IL-13Rα2/β-actin were measured by RT-qPCR assay. Each value denotes mean ± SD of quadruple runs performed independently in triplicate.

FIG. 19 shows IFN-γ release by scFv IL-13Rα2 CAR-T cells when co-cultured with tumor cells. When the scFv IL-13Rα2 CAR-T were exposed to IL-13Rα2 positive tumors, the CAR-T cells made a large quantity of IFN-γ. However, when exposed to IL-13Rα2 negative T98G or IL-13Rα2 KO tumor cells, they secreted only a basal amount of IFN-γ. Representative data of 3 independent experiments performed in quadruplicate is shown.

FIG. 20 shows a Boyden chamber assay. The *in vitro* migration assay demonstrates that scFv-IL-13Rα2 CAR-T cells can migrate from the upper chamber to the lower chamber in a concentration dependent manner in medium containing different concentrations of HuIL-13Rα2Fc or conditioned medium from IL-13Rα2 positive U251 tumor cell culture. No migration was observed in the control or conditioned medium from IL-13Rα2 negative T98G tumor cell cultures. Each value is a mean ± SD of four independent experiments performed in triplicate.

SEQUENCE LISTING

Any nucleic acid and amino acid sequences listed herein or in the accompanying Sequence Listing are shown using standard letter abbreviations for nucleotide bases and amino acids, as defined in 37 C.F.R. § 1.822. In at least some cases, only one strand of each nucleic acid sequence is shown, but the complementary strand is understood as included by any reference to the displayed

strand. The Sequence Listing is submitted as an ASCII text file, "Sequence.txt," created on March 30, 2022, 24,576 bytes, which is incorporated by reference herein. In the accompanying sequence listing:

SEQ ID NO: 1 is the amino acid sequence of an exemplary IL-13Rα2 scFv.

QVQLVQSGAEVKKPGASVKVSCKASGYTFTSYAMHWVRQAPGQRLEWMGWINAGNGN
TKYSQKFQGRVTITRDTSASTAYMELSSLRSEDTAVYYCARMNHMIPLKAWGQGTLVTVS
SGGGGSGGGGSGGSALAIQMTQSPSSLSASVGDRVTITCRASQGIRNDLGWYQQKPGKAP
KLLIYAASSLQSGVPSRFSGSGSGTDFTLTISSLQPEDFATYYCLQMYNYRTFGQGTKLEIK
RA

SEQ ID NO: 2 is an exemplary nucleic acid sequence encoding the IL-13Rα2 scFv of SEQ ID NO:1.

AGGTGCAGCTGGTGCAGAGCGGAGCAGAGGTGAAGAAGCCAGGAGCCTCTGTGAAGG
TGAGCTGCAAGGCCTCCGGCTACACATTCACCTCCTATGCCATGCACTGGGTGAGACA
GGCACCTGGACAGAGGCTGGAGTGGATGGGCTGGATCAACGCCGGCAACGGCAATAC
AAAGTACTCTCAGAAGTTTCAGGGCCGCGTGACAATCACCCGGGACACATCCGCCTCT
ACCGCCTATATGGAGCTGAGCTCCCTGCGGTCCGAGGATACCGCCGTGTACTATTGCG
CCAGAATGAATCACATGATCCCACTGAAGGCATGGGGACAGGGCACACTGGTGACCG
TGTCTAGCGGAGGAGGAGGCAGCGGAGGAGGAGGCTCCGGCGGCTCTGCCCTGGCCA
TCCAGATGACCCAGTCCCCATCCTCTCTGAGCGCCTCCGTGGGCGACCGCGTGACAAT
CACCTGTCGGGCCAGCCAGGGCATCAGAAACGATCTGGGCTGGTACCAGCAGAAGCC
CGGCAAGGCCCCTAAGCTGCTGATCTATGCAGCAAGCTCCCTGCAGTCTGGAGTGCCT
AGCCGGTTCTCTGGCAGCGGCTCCGGAACAGACTTTACACTGACCATCTCTAGCCTGC
AGCCAGAGGATTTCGCCACCTACTATTGCCTGCAGATGTACAATTATAGAACATTTGG
CCAGGGCACCAAGCTGGAGATCAAGAGGGCC

SEQ ID NO: 3 is the amino acid sequence of an exemplary IL-13Rα2-CAR.

MALPVTALLLPLALLLHAARPQVQLVQSGAEVKKPGASVKVSCKASGYTFTSYAMHWVR
QAPGQRLEWMGWINAGNGNTKYSQKFQGRVTITRDTSASTAYMELSSLRSEDTAVYYCA
RMNHMIPLKAWGQGTLVTVSSGGGGSGGGGSGGSALAIQMTQSPSSLSASVGDRVTITCR
ASQGIRNDLGWYQQKPGKAPKLLIYAASSLQSGVPSRFSGSGSGTDFTLTISSLQPEDFATY
YCLQMYNYRTFGQGTKLEIKRAAKPTTTPAPRPPTPAPTIASQPLSLRPEACRPAAGGAVH
TRGLDFACDFWVLVVVGGVLACYSLLVTVAFIIFWVRSKRSRGGHSDYMNMTPRRPGPTR
KHYQPYAPPRDFAAYRSVDKRGRKKLLYIFKQPFMRPVQTTQEEDGCSCRFPEEEEGGCEL
GGGRVKFSRSADAPAYQQGQNQLYNELNLGRREEYDVLDKRRGRDPEMGGKPQRRKNP
QEGLYNELQKDKMAEAYSEIGMKGERRRGKGHDGLYQGLSTATKDTYDALHMQALPPR

SEQ ID NO: 4 is an exemplary nucleic acid sequence encoding the IL-13Rα2-CAR construct of SEQ ID NO: 3.

ATGGCACTGCCTGTGACCGCCCTGCTGCTGCCACTGGCCCTGCTGCTGCACGCAGCCCG
GCCACAGGTGCAGCTGGTGCAGAGCGGAGCAGAGGTGAAGAAGCCAGGAGCCTCTGT
GAAGGTGAGCTGCAAGGCCTCCGGCTACACATTCACCTCCTATGCCATGCACTGGGTG
AGACAGGCACCTGGACAGAGGCTGGAGTGGATGGGCTGGATCAACGCCGGCAACGGC
AATACAAAGTACTCTCAGAAGTTTCAGGGCCGCGTGACAATCACCCGGGACACATCCG
CCTCTACCGCCTATATGGAGCTGAGCTCCCTGCGGTCCGAGGATACCGCCGTGTACTAT

TGCGCCAGAATGAATCACATGATCCCACTGAAGGCATGGGGACAGGGCACACTGGTG
ACCGTGTCTAGCGGAGGAGGAGGCAGCGGAGGAGGAGGCTCCGGCGGCTCTGCCCTG
GCCATCCAGATGACCCAGTCCCCATCCTCTCTGAGCGCCTCCGTGGGCGACCGCGTGA
CAATCACCTGTCGGGCCAGCCAGGGCATCAGAAACGATCTGGGCTGGTACCAGCAGA
AGCCCGGCAAGGCCCCTAAGCTGCTGATCTATGCAGCAAGCTCCCTGCAGTCTGGAGT
GCCTAGCCGGTTCTCTGGCAGCGGCTCCGGAACAGACTTTACACTGACCATCTCTAGCC
TGCAGCCAGAGGATTTCGCCACCTACTATTGCCTGCAGATGTACAATTATAGAACATTT
GGCCAGGGCACCAAGCTGGAGATCAAGAGGGCCGCCAAGCCAACCACAACCCCAGCA
CCTCGCCCCCCTACACCAGCACCAACCATCGCATCCCAGCCTCTGTCTCTGAGACCAGA
GGCATGTAGGCCAGCAGCAGGAGGAGCAGTGCACACAAGGGGCCTGGACTTCGCCTG
CGATTTTTGGGTGCTGGTGGTGGTGGGAGGCGTGCTGGCCTGTTACTCTCTGCTGGTGA
CCGTGGCCTTCATCATCTTTTGGGTGAGGAGCAAGCGGAGCAGGGGAGGACACAGCG
ACTACATGAACATGACACCACGGAGACCTGGACCAACCAGGAAGCACTACCAGCCTT
ATGCACCACCAAGGGACTTCGCAGCATACCGCAGCGTGGATAAGAGAGGCAGGAAGA
AGCTGCTGTATATCTTCAAGCAGCCCTTCATGCGGCCCGTGCAGACAACCCAGGAGGA
GGACGGCTGCTCCTGTAGATTCCCCGAGGAGGAGGAGGGAGGATGTGAGCTGGGAGG
CGGCAGAGTGAAGTTTTCTCGGAGCGCCGATGCACCTGCATACCAGCAGGGACAGAAT
CAGCTGTATAACGAGCTGAATCTGGGCAGGCGCGAGGAGTACGACGTGCTGGATAAG
AGGCGGGGCCGGGACCCCGAGATGGGAGGCAAGCCACAGAGGCGCAAGAACCCCCA
GGAGGGCCTGTACAATGAGCTGCAGAAGGACAAGATGGCCGAGGCCTATTCCGAGAT
CGGCATGAAGGGAGAGCGGAGAAGGGGCAAGGGACACGATGGCCTGTACCAGGGCCT
GTCTACAGCCACCAAGGACACCTATGATGCCCTGCACATGCAGGCCCTGCCTCCACGC
TAAGCGGCCGC

SEQ ID NO: 5 is the amino acid sequence of an exemplary CD8α signal peptide. MALPVTALLLPLALLLHAARP

SEQ ID NO: 6 is the amino acid sequence of an exemplary CD8α hinge domain. AKPTTTPAPRPPTPAPTIASQPLSLRPEACRPAAGGAVHTRGLDFAC

SEQ ID NO: 7 is the amino acid sequence of an exemplary CD28 transmembrane domain (CD28TM). DFWVLVVVGGVLACYSLLVTVAFIIFWVR

SEQ ID NO: 8 is the amino acid sequence of an exemplary CD28 cytoplasmic signaling domain. SKRSRGGHSDYMNMTPRRPGPTRKHYQPYAPPRDFAAYRSVD

SEQ ID NO: 9 is the amino acid sequence of an exemplary 4-1BB cytoplasmic signaling domain. KRGRKKLLYIFKQPFMRPVQTTQEEDGCSCRFPEEEEGGCEL

SEQ ID NO: 10 is the amino acid sequence of an exemplary CD3ζ cytoplasmic signaling domain.

RVKFSRSADAPAYQQGQNQLYNELNLGRREEYDVLDKRRGRDPEMGGKPQRRKNPQEGL
YNELQKDKMAEAYSEIGMKGERRRGKGHDGLYQGLSTATKDTYDALHMQALPPR

SEQ ID NO: 11 is an exemplary nucleic acid sequence encoding the CD8α signal peptide of SEQ ID NO: 5.

ATGGCACTGCCTGTGACCGCCCTGCTGCTGCCACTGGCCCTGCTGCTGCACGCAGCCCGGCC
AC

SEQ ID NO: 12 is an exemplary nucleic acid sequence encoding the CD8α hinge domain of SEQ ID NO: 6.

GCCAAGCCAACCACAACCCCAGCACCTCGCCCCCCTACACCAGCACCAACCATCGCATCCC
AGCCTCTGTCTCTGAGACCAGAGGCATGTAGGCCAGCAGCAGGAGGAGCAGTGCACACAA
GGGGCCTGGACTTCGCCTGC

SEQ ID NO: 13 is an exemplary nucleic acid sequence encoding the CD28 transmembrane domain (CD28TM) of SEQ ID NO: 7.

GATTTTTGGGTGCTGGTGGTGGTGGGAGGCGTGCTGGCCTGTTACTCTCTGCTGGTGAC
CGTGGCCTTCATCATCTTTTGGGTGAGG

SEQ ID NO: 14 is an exemplary nucleic acid sequence encoding the CD28 cytoplasmic signaling domain of SEQ ID NO: 8.

AGCAAGCGGAGCAGGGGAGGACACAGCGACTACATGAACATGACACCACGGAGACCTGG
ACCAACCAGGAAGCACTACCAGCCTTATGCACCACCAAGGGACTTCGCAGCATACCGCAG
C

SEQ ID NO: 15 is an exemplary nucleic acid sequence encoding the 4-1BB cytoplasmic signaling domain of SEQ ID NO: 9.

AAGAGAGGCAGGAAGAAGCTGCTGTATATCTTCAAGCAGCCCTTCATGCGGCCCGTGCAGA
CAACCCAGGAGGAGGACGGCTGCTCCTGTAGATTCCCCGAGGAGGAGGAGGGAGGATGTG
AGCTG

SEQ ID NO: 16 is an exemplary nucleic acid sequence encoding the CD3ζ cytoplasmic signaling domain of SEQ ID NO: 10.

AGAGTGAAGTTTTCTCGGAGCGCCGATGCACCTGCATACCAGCAGGGACAGAATCAGCTG
TATAACGAGCTGAATCTGGGCAGGCGCGAGGAGTACGACGTGCTGGATAAGAGGCGGGG
CCGGGACCCCCGAGATGGGAGGCAAGCCACAGAGGCGCAAGAACCCCCAGGAGGGC
CTGTACAATGAGCTGCAGAAGGACAAGATGGCCGAGGCCTATTCCGAGATCGGCATG
AAGGGAGAGCGGAGAAGGGGCAAGGGACACGATGGCCTGTACCAGGGCCTGTCTACAGC
CACCAAGGACACCTATGATGCCCTGCACATGCAGGCCCTGCCTCCACGC

SEQ ID NO: 17 is the nucleic acid sequence of an exemplary siRNA for silencing IL-13Rα2.

GCTACCATTTGGTTTCATCTT

SEQ ID NO: 18 is the amino acid sequence of an exemplary scFv IL-13Rα2 CAR

MALPVTALLLPLALLLHAARPQVQLVQSGAEVKKPGASVKVSCKASGYTFTSYAMHWVRQAP
GQRLEWMGWINAGNGNTKYSQKFQGRVTITRDTSASTAYMELSSLRSEDTAVYYCARMNHMI
PLKAWGQGTLVTVSSGGGGSGGGGSGGSALAIQMTQSPSSLSASVGDRVTITCRASQGIRNDLG
WYQQKPGKAPKLLIYAASSLQSGVPSRFSGSGSGTDFTLTISSLQPEDFATYYCLQMYNYRTFG
QGTKLEIKRAAKPTTTPAPRPPTPAPTIASQPLSLRPEACRPAAGGAVHTRGLDFACDFWVLVVV
GGVLACYSLLVTVAFIIFWVRSKRSRGGHSDYMNMTPRRPGPTRKHYQPYAPPRDFAAYRSVD
KRGRKKLLYIFKQPFMRPVQTTQEEDGCSCRFPEEEEGGCELGGGRVKFSRSADAPAYQQGQN
QLYNELNLGRREEYDVLDKRRGRDPEMGGKPQRRKNPQEGLYNELQKDKMAEAYSEIGMKGE
RRRGKGHDGLYQGLSTATKDTYDALHMQALPPR

DETAILED DESCRIPTION

I. Terms

Unless otherwise noted, technical terms are used according to conventional usage. Definitions of common terms in molecular biology may be found in *Lewin's Genes X*, ed. Krebs *et al.*, Jones and Bartlett Publishers, 2009 (ISBN 0763766321); Kendrew *et al.* (eds.), *The Encyclopedia of Molecular Biology*, published by Blackwell Publishers, 1994 (ISBN 0632021829); Robert A. Meyers (ed.), *Molecular Biology and Biotechnology: a Comprehensive Desk Reference*, published by Wiley, John & Sons, Inc., 1995 (ISBN 0471186341); and George P. Rédei, E*ncyclopedic Dictionary of Genetics, Genomics, Proteomics and Informatics,* 3rd Edition, Springer, 2008 (ISBN: 1402067534), and other similar references.

Unless otherwise explained, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which this disclosure belongs. The singular terms "a," "an," and "the" include plural referents unless the context clearly indicates otherwise. "Comprising A or B" means including A, or B, or A and B. It is further to be understood that all base sizes or amino acid sizes, and all molecular weight or molecular mass values, given for nucleic acids or polypeptides are approximate, and are provided for description.

Although methods and materials similar or equivalent to those described herein can be used in the practice or testing of the present disclosure, suitable methods and materials are described below. All publications, patent applications, patents, and other references mentioned herein are incorporated by reference in their entirety. In case of conflict, the present specification, including explanations of terms, will control. In addition, the materials, methods, and examples are illustrative only and not intended to be limiting.

In order to facilitate review of the various embodiments of the disclosure, the following explanations of specific terms are provided:

Antibody: A polypeptide ligand comprising at least one variable region that recognizes and binds (such as specifically recognizes and specifically binds) an epitope of an antigen. Mammalian immunoglobulin molecules are composed of a heavy (H) chain and a light (L) chain, each of which has a variable region, termed the variable heavy (V_H) region and the variable light (V_L) region, respectively. Together, the V_H region and the V_L region are responsible for binding the antigen recognized by the antibody. There are five main heavy chain classes (or isotypes) of mammalian immunoglobulin, which determine the functional activity of an antibody molecule: IgM, IgD, IgG, IgA and IgE.

Antibody variable regions contain "framework" regions and hypervariable regions, known as "complementarity determining regions" or "CDRs." The CDRs are primarily responsible for binding to an epitope of an antigen. The framework regions of an antibody serve to position and align the CDRs in three-dimensional space. The amino acid sequence boundaries of a given CDR can be readily determined using any of a number of well-known numbering schemes, including those described by Kabat *et al.* (*Sequences of Proteins of Immunological Interest*, U.S. Department of Health and Human Services, 1991; the "Kabat" numbering scheme), Chothia *et al.* (see Chothia and Lesk, *J Mol Biol* 196:901-917, 1987; Chothia *et al.*, *Nature* 342:877, 1989; and Al-Lazikani *et al.*, (JMB 273,927-948, 1997; the "Chothia" numbering scheme), and the ImMunoGeneTics (IMGT) database (see, Lefranc, *Nucleic Acids Res* 29:207-9, 2001; the "IMGT" numbering scheme). The Kabat and IMGT databases are maintained online.

A single-chain fragment variable (scFv) antibody is a genetically engineered molecule containing the V_H and V_L domains of one or more antibody(ies) linked by a suitable polypeptide linker as a genetically fused single chain molecule (see, for example, Bird *et al.*, *Science*, 242:423-426, 1988; Huston *et al.*, *Proc. Natl. Acad. Sci.,* 85:5879-5883, 1988; Ahmad *et al.*, *Clin. Dev. Immunol.*, 2012, doi:10.1155/2012/980250; Marbry, *IDrugs*, 13:543-549, 2010). The intramolecular orientation of the V_H-domain and the V_L-domain in a scFv, is typically not decisive for scFvs. Thus, scFvs with both possible arrangements (V_H-domain-linker domain-V_L-domain; V_L-domain-linker domain-V_H-domain) may be used. In a dsFv the V_H and V_L have been mutated to introduce a disulfide bond to stabilize the association of the chains. Diabodies also are included, which are bivalent, bispecific antibodies in which V_H and V_L domains are expressed on a single polypeptide chain, but using a linker that is too short to allow for pairing between the two domains on the same chain, thereby forcing the domains to pair with complementary domains of another chain and creating two antigen binding sites (see, for example, Holliger *et al.*, *Proc. Natl. Acad. Sci.,* 90:6444-6448, 1993; Poljak *et al.*, *Structure,* 2:1121-1123, 1994).

Antibodies also include genetically engineered forms such as chimeric antibodies (such as humanized murine antibodies) and heteroconjugate antibodies (such as bispecific antibodies). *See also*, *Pierce Catalog and Handbook*, 1994-1995 (Pierce Chemical Co., Rockford, IL); Kuby, J., *Immunology*, 3rd Ed., W.H. Freeman & Co., New York, 1997.

Cancer: A disease characterized by abnormal or uncontrolled cell growth. Other features often associated with cancer include metastasis, interference with the normal functioning of neighboring cells, release of cytokines or other secretory products at abnormal levels, suppression or aggravation of inflammatory or immunological response, invasion of surrounding or distant tissues or organs, such as lymph nodes, etc. "Metastatic disease" refers to cancer cells that have left the original tumor site and migrated to other parts of the body, for example via the bloodstream or lymph system.

Checkpoint Inhibitor: Includes inhibitors of cell cycle checkpoints as well as immune checkpoints. Cell cycle checkpoints refer to safeguard mechanisms that ensure a cell correctly completes each cell cycle phase during mitotic division. Checkpoint inhibitors can sensitize cancer cells to DNA damaging drugs by causing cells with DNA damage to bypass the S and G2/M arrest and enter mitosis, leading to cell death by mitotic catastrophe. Cell cycle checkpoint inhibitors are described in more detail by Visconti *et al.*, *J Exp Clin Cancer Res.* 35(1): 153, 2016. Immune checkpoints refer to safeguard mechanisms that prevent autoreactive immune cells. Immune checkpoints activate when an inhibitory receptor of an immune cell (such as a T cell) recognizes antigens on a cell as "self" antigens. While immune checkpoints are important mechanisms to prevent autoimmune disorders, cancerous cells can take advantage of immune checkpoints to suppress or evade immune recognition. Thus, checkpoint inhibitors can help immune cells recognize and eliminate cancerous cells.

Many checkpoint inhibitors are known in the art. Some non-limiting examples include ipilimumab (Yervoy®), nivolumab (Opdivo®), pembrolizumab (Keytruda®), atezolizumab (Tencentriq®), avelumab (Bavencio®), durvalumab (Imfinzi®), cemiplimab (Libtayo®), palbociclib (Ibrance®), ribociclib (Kisquali®), and abemaciclib (Verzenio®). Further examples are provided in Qiu *et al.*, *Journal of the European Society for Therapeutic Radiology and Oncology*, 126(3):450-464, 2018; Visconti *et al.*, *J Exp Clin Cancer Res.* 35(1): 153, 2016; and Mills *et al. Cancer Res.* 77(23): 6489-6498, 2017.

A checkpoint inhibitor may also include a spindle assembly checkpoint inhibitor. For example, spindle assembly checkpoint inhibitors include MK-1775 (AZD1775), taxanes, or vinca alkaloids (*see* Zhou and Giannakakou. *Curr Med Chem Anticancer Agents.* 5:65–71, 2005; and Visconti *et al.*, *J Exp Clin Cancer Res.* 35(1): 153, 2016).

Chimeric antigen receptor (CAR): A chimeric molecule that includes an antigen-binding portion (such as a single domain antibody or scFv) and a signaling domain, such as a signaling domain from a T cell receptor (*e.g.,* CD3ζ). Typically, CARs include an antigen-binding portion, a transmembrane domain, and an intracellular domain. The intracellular domain typically includes a signaling chain having an immunoreceptor tyrosine-based activation motif (ITAM), such as CD3ζ or FcεRIγ. In some instances, the intracellular domain also includes the intracellular portion of at least one additional co-stimulatory domain, such as CD28, 4-1BB (CD137), ICOS, OX40 (CD134), CD27 and/or DAP10.

Complementarity determining region (CDR): A region of hypervariable amino acid sequence that defines the binding affinity and specificity of an antibody. The light and heavy chains of a mammalian immunoglobulin each have three CDRs, designated VL-CDR1, VL-CDR2, VL-CDR3 and VH-CDR1, VH-CDR2, VH-CDR3, respectively.

Effective Amount: The amount of an agent, such as the disclosed IL-13Rα2-CAR-T cells or IL-13Rα2-CAR-NK cells, that is sufficient to treat, reduce, and/or ameliorate the symptoms and/or underlying cause of a disease or pathological condition, such as cancer in a subject. In a non-limiting example, an effective amount is an amount sufficient to inhibit or reduce tumor growth in the subject.

Histone Deacetylase (HDAC) Inhibitors: Several HDAC inhibitors have been shown to induce cell growth arrest and apoptosis, and thus are potential anti-cancer therapeutics. Some non-limiting examples of HDAC inhibitors include trichostatin A (TSA), suberoylanilide hydroxamic acid (SAHA), sodium butyrate (NaB), SP600125 (Sigma-Aldrich), SR11302 (Tocris Bioscience), and romidepsin (*see* Marks and Jiang, *Cell Cycle* 2005, 4:549-551; Duvic *et al. Blood* 109:31-39, 2007; and Fujisawa *et al. Journal of Translational Medicine* 9:37, 2011).

IL-13-PE Immunotoxin: IL-13-PE is a recombinant immunotoxin consisting of IL-13 and a truncated Pseudomonas exotoxin. IL-13-PE has been shown to be highly cytotoxic to tumor cells expressing high levels of IL-13Rα2 both *in vitro* and *in vivo*. (*see* Debinski *et al.*, *Clin Cancer Res.* 1:1253-1258, 1995; Debinski *et al.*, *Biol Chem.* 270:16775-16780, 1995; and Joshi *et al. Clin Cancer Res.* 8:1948-1956, 2002).

Interleukin-13 Receptor α2 (IL-13Rα2): IL-13Rα2 is a high affinity receptor for the pleiotropic immune regulatory cytokine interleukin-13 (IL-13) and a known tumor antigen. The significance of IL-13Rα2 expression in cancer is not known and the mechanism of upregulation is still unclear, however, it has been shown that IL-13Rα2 is overexpressed in a variety of human cancers, including malignant glioma, head and neck cancer, Kaposi's sarcoma, renal cell carcinoma, and ovarian carcinoma. Further, in a recent study, it has been shown that 71% of

pancreatic tumors overexpress IL-13Rα2 (Fujisawa *et al. Journal of Translational Medicine* 9:37, 2011).

Isolated: An "isolated" biological component, such as a nucleic acid, protein (including antibodies) or cell, that has been substantially separated or purified away from other biological components in the environment (such as a cell) in which the component occurs, *e.g.*, other chromosomal and extra-chromosomal DNA and RNA, proteins and cells. Nucleic acids and proteins that have been "isolated" include nucleic acids and proteins purified by standard purification methods. The term also embraces nucleic acids and proteins prepared by recombinant expression in a host cell as well as chemically synthesized nucleic acids.

Macrophage: Large, migratory mononuclear phagocytic cells derived from bone marrow precursors that are found in most tissues of the human body. Macrophages express pattern recognition receptors (PRRs), including Toll-like receptors (TLRs), C-type lectin receptors, scavenger receptors, retinoic acid-inducible gene 1 (RIG1)-like helicase receptors (RLRs) and NOD-like receptors, to recognize signals associated with pathogens, foreign substances, and dead or dying cells.

Macrophages are highly heterogenous and include several sub-classes. Classically activated macrophages (M1 macrophages) mediate defense of the host from various pathogens (*e.g.*, bacteria, protozoa, and viruses) and also mediate anti-tumor immune responses. Alternatively activated macrophages (M2 macrophages) have an anti-inflammatory role and regulate wound healing. Regulatory macrophages secrete large amounts of interleukin-10 (IL-10) in response to Fc receptor-γ ligation. Less-well-defined macrophage subsets include tumor-associated macrophages, which suppress anti-tumor immunity, and myeloid-derived suppressor cells (Murray *et al.* (2011) *Nat Rev Immunol* 11:723–737).

Natural Killer (NK) cells: Cells of the immune system that kill target cells in the absence of a specific antigenic stimulus and without restriction according to MHC class. Target cells can be tumor cells or cells harboring viruses. NK cells are characterized by the presence of CD56 and the absence of CD3 surface markers. NK cells typically comprise approximately 10 to 15% of the mononuclear cell fraction in normal peripheral blood. Historically, NK cells were first identified by their ability to lyse certain tumor cells without prior immunization or activation. NK cells are thought to provide a "back up" protective mechanism against viruses and tumors that might escape the CTL response by down-regulating MHC class I presentation. In addition to being involved in direct cytotoxic killing, NK cells also serve a role in cytokine production, which can be important to control cancer and infection.

In some examples, a **"modified NK cell"** is a NK cell transduced or transfected with a

heterologous nucleic acid (such as one or more of the nucleic acids or vectors disclosed herein) or expressing one or more heterologous proteins. The terms "modified NK cell" and "transduced NK cell" are used interchangeably in some examples herein.

Pharmaceutically Acceptable Carrier: Includes any and all solvents, dispersion media, coatings, antibacterial and antifungal agents, isotonic and absorption delaying agents, and the like, compatible with pharmaceutical administration (see, *e.g., Remington: The Science and Practice of Pharmacy*, Adejare (Ed.), Academic Press, London, United Kingdom, 23rd Edition (2021). Examples of such carriers or diluents include, but are not limited to, water, saline, Ringer's solutions, dextrose solution, balanced salt solutions, and 5% human serum albumin. Liposomes and non-aqueous vehicles such as fixed oils may also be used. Supplementary active compounds can also be incorporated into the compositions. Actual methods for preparing administrable compositions include those provided in *Remington: The Science and Practice of Pharmacy*, Adejare (Ed.), Academic Press, London, United Kingdom, 23rd Edition (2021).

Purified: The term purified does not require absolute purity; rather, it is intended as a relative term. Thus, for example, a purified protein, nucleic acid, or cell preparation is one in which the protein, nucleic acid, or cell is more enriched than the protein, nucleic acid, or cell is in its initial environment. In one embodiment, a preparation is purified such that the protein, nucleic acid, or cell represents at least 50% of the total content of the preparation. A substantially purified protein or nucleic acid is at least 60%, 70%, 80%, 90%, 95% or 98% pure. Thus, in one specific, non-limiting example, a substantially purified protein or nucleic acid is 90% free of other components.

Recombinant: A nucleic acid or protein that has a sequence that is not naturally occurring or has a sequence that is made by an artificial combination of two otherwise separated segments of sequence (*e.g.*, a "chimeric" sequence). This artificial combination can be accomplished by chemical synthesis or by the artificial manipulation of isolated segments of nucleic acids, for example, by genetic engineering techniques.

Subject: Living multi-cellular vertebrate organisms, a category that includes human and non-human mammals, including but not limited to non-human primates, rodents, and the like. In specific examples disclosed herein, the subject is human.

T cell: A white blood cell (lymphocyte) that is an important mediator of the immune response. T cells include, but are not limited to, $CD4^+$ T cells and $CD8^+$ T cells. A $CD4^+$ T cell is an immune cell that carries a marker on its surface known as "cluster of differentiation 4" (CD4). These cells, also known as helper T cells, help orchestrate the immune response, including antibody responses as well as killer T cell responses. $CD8^+$ T cells carry the "cluster of differentiation 8"

(CD8) marker. In one embodiment, a $CD8^+$ T cell is a cytotoxic T lymphocyte (CTL). In another embodiment, a $CD8^+$ cell is a suppressor T cell.

Activated T cells can be detected by an increase in cell proliferation and/or expression or secretion of one or more cytokines (such as IL-2, IL-4, IL-6, IFN-γ, or TNFα). Activation of $CD8^+$ T cells can also be detected by an increase in cytolytic activity in response to an antigen.

A **natural killer T (NKT) cell** is a class of T cell that expresses both T-cell receptors (TCR) characteristic of adaptive immunity, and surface receptors for NK cells, which are part of the innate immune response (*e.g.*, the cell-surface marker NK1.1 normally associated with NK cells along with an α/β T-cell receptor) (Tupin *et al.* (2007) *Nat Rev Microbiol* 5:405–417).

In some examples, a **"modified T cell"** is a T cell transduced or transfected with a heterologous nucleic acid (such as one or more of the nucleic acids or vectors disclosed herein) or expressing one or more heterologous proteins. The terms "modified T cell" and "transduced T cell" are used interchangeably in some examples herein. Similarly, a "**modified NKT cell"** is an NKT cell transduced or transfected with a heterologous nucleic acid (such as one or more of the nucleic acids or vectors disclosed herein) or expressing one or more heterologous proteins.

Transduced or Transformed: A transformed cell is a cell into which a nucleic acid molecule has been introduced by molecular biology techniques. As used herein, the terms transduction and transformation encompass all techniques by which a nucleic acid molecule might be introduced into such a cell, including transduction or transfection with viral vectors, the use of plasmid vectors, and introduction of DNA by electroporation, lipofection, and particle gun acceleration.

Treating or ameliorating a disease: "Treating" refers to a therapeutic intervention that decreases or inhibits a sign or symptom of a disease or pathological condition after it has begun to develop, such as a reduction in tumor size or tumor burden. "Ameliorating" refers to the reduction in the number or severity of signs or symptoms of a disease, such as cancer.

Vector: A nucleic acid molecule that can be introduced into a host cell (for example, by transfection or transduction), thereby producing a transformed host cell. Recombinant DNA vectors are vectors having recombinant DNA. A vector can include nucleic acid sequences that permit it to replicate in a host cell, such as an origin of replication. A vector can also include one or more selectable marker genes and other genetic elements known in the art. Viral vectors are recombinant nucleic acid vectors having at least some nucleic acid sequences derived from one or more viruses. A replication deficient viral vector is a vector that requires complementation of one or more regions of the viral genome required for replication due to a deficiency in at least one replication-essential gene function.

II. Overview of Several Embodiments

Disclosed herein are interleukin-13 receptor α2 (IL-13Rα2) binding agents, including an scFv. Also disclosed are chimeric antigen receptors (CARs) that include an IL-13Rα2-specific binding agent (such as an IL-13Rα2 scFv disclosed herein) fused to a hinge region, a transmembrane domain and an intracellular domain comprising one or more signaling domains (*e.g.*, one or more co-stimulatory domains). In some examples, the one or more signaling domains are from CD28, 4-1BB and/or CD3ζ. In some examples, the hinge region is from CD8α and the transmembrane region is from CD28. The CAR may also encode a signal peptide, such as a CD8α signal peptide. Also provided are nucleic acids encoding IL-13Rα2-specific binding agents and IL-13Rα2-CARs and vectors including the nucleic acids.

Also provided herein are immune cells (*e.g.*, T cells, NK cells, NKT cells, or macrophages) transformed with a nucleic acid or vector encoding an IL-13Rα2-specific binding agent or an IL-13Rα2-CAR. In some examples the immune cell expresses an IL-13Rα2-CAR. The cells can be obtained from a blood sample, for example, from peripheral blood of a subject, prior to transforming with a nucleic acid or vector.

Also provided are methods of treating a cancer that expresses IL-13Rα2 in a subject. In some embodiments, the method includes administering to the subject an effective amount of immune cells expressing an IL-13Rα2-CAR. In some examples, the immune cells are autologous to the subject. In other examples, the immune cells are allogeneic. In some examples, the subject receives an additional treatment, such as surgery, radiation, chemotherapy, an additional immunotherapy, administration of a histone deacetylase (HDAC) inhibitor, administration of a cell cycle and/or checkpoint inhibitor, administration of adrenomedullin, administration of an IL-13-PE immunotoxin, or any combination of two or more thereof. In some examples, the subject has cancer, for example, pancreatic cancer, glioblastoma, head and neck squamous cell carcinoma, ovarian cancer, uterine cancer, prostate cancer, breast cancer, melanoma, non-small cell lung cancer (NSCLC), renal cell carcinoma, Kaposi sarcoma, or adrenal carcinoma.

III. IL-13Rα2 Specific Binding Agents

Disclosed herein is an interleukin-13 receptor α2 (IL-13Rα2) binding agent, that in some examples is used as the targeting portion of a chimeric antigen receptor (CAR). In some embodiments, the IL-13Rα2 binding agent is a single-chain fragment variable (scFv) that specifically binds IL-13Rα2.

In some embodiments, the IL13-Rα2 binding agent is an antibody or scFv that includes the CDR sequences provided in Table 1. In some examples, the antibody or scFv specifically binds IL-13Rα2 and includes the variable heavy chain (VH) domain complementarity determining region 1 (CDR1), CDR2, and CDR3 amino acid sequences of SEQ ID NO: 1, and variable light chain (VL) domain complementarity determining region 1 (CDR1), CDR2, and CDR3 of SEQ ID NO: 1, respectively. In some examples, the antibody or scFv includes an amino acid sequence having at least 90% sequence identity (for example, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98% or at least 99% sequence identity) to the V_H and V_L sequences disclosed herein. In some examples, the V_H domain includes amino acids 1-118 of SEQ ID NO: 1 and the V_L domain includes amino acids 134-239 of SEQ ID NO: 1.

In some embodiments, the binding agent is an scFv that includes the CDR amino acid sequences provided in Table 1, and has at least 90% sequence identity (for example, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98% or at least 99% sequence identity) to the amino acid sequence of SEQ ID NO: 1. In other embodiments, the binding agent is an scFv and includes or consists of the amino acid sequence of SEQ ID NO: 1.

Table 1. Location of the CDRs in the IL-13Rα2 scFv sequence

CDR	**Amino Acid Sequence (Position in SEQ ID NO: 1)**	**Nucleic Acid Sequence (Position in SEQ ID NO: 2)**
VH CDR1	SYAMH (31-35)	TCCTATGCCATGCAC (90-104)
VH CDR2	WINAGNGNTKYSQKFQG (50-66)	TGGATCAACGCCGGCAACGGCAA TACAAAGTACTCTCAGAAGTTTCA GGGC (147-197)
VH CDR3	MNHMIPLKA (99-107)	ATGAATCACATGATCCCACTGAAG GCA (294-320)
VL CDR1	RASQGIRNDLG (157-167)	CGGGCCAGCCAGGGCATCAGAAA CGATCTGGGC (468-500)

CDR	Amino Acid Sequence (Position in SEQ ID NO: 1)	Nucleic Acid Sequence (Position in SEQ ID NO: 2)
VL CDR2	AASSLQS (183-189)	GCAGCAAGCTCCCTGCAGTCT (546-566)
VL CDR3	LQMYNYRT (222-229)	CTGCAGATGTACAATTATAGAACA (663-686)

In some embodiments, the scFv binds a target cell, such as a cell expressing IL-13Rα2. In some examples, the cell is a cancer cell (including a tumor cell), such as a pancreatic cancer, glioblastoma, head and neck squamous cell carcinoma, ovarian cancer, uterine cancer, prostate cancer, breast cancer, melanoma, non-small cell lung cancer (NSCLC), renal cell carcinoma, Kaposi sarcoma, or adrenal carcinoma cell. scFv binding may be increased or improved when IL-13Rα2 expression is upregulated in the target cell.

IV. IL-13Rα2 Chimeric Antigen Receptors

Provided herein are IL-13Rα2 chimeric antigen receptors (IL-13Rα2-CARs) that include the IL13-Rα2 specific binding agents described in Section III. In some embodiments, the IL-13Rα2 CAR includes (a) an antigen binding domain including an IL-13Rα2-specifc scFv (such as SEQ ID NO: 1); (b) a hinge domain; (c) a transmembrane domain; and (d) an intracellular domain including one or more signaling domains. The CAR may also include a signal peptide, linker(s), and/or sequences for recombinant construction (such as a restriction enzyme site).

In some embodiments, the antigen binding domain is an IL-13Rα2-specifc scFv, such as those described in Section II. In some examples, the CAR includes an IL-13Rα2 scFv including an amino acid sequence with at least 90% sequence identity (for example, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98% at least 99% identity) to SEQ ID NO: 1 or including or consisting of the amino acid sequence of SEQ ID NO: 1.

In some embodiments, the hinge domain is a CD8α hinge domain for example including or consisting of the amino acid sequence of SEQ ID NO: 6. Other hinge domains can be used, such as hinge regions from other immunoglobulins (for example, IgG1, IgG4, or IgD) or a hinge region from CD28, or CD40. In some embodiments, the transmembrane domain is a CD28 transmembrane domain, for example including or consisting of the amino acid sequence of SEQ ID NO: 7. The transmembrane domain can also be from other T cell proteins, such as CD8, CD4,

CD3ζ, CD40, OX40-L, 4-1BB, ICOS, ICOS-L, CD80, CD86, ICAM-1, LFA-1, ICAM-1, CD56, CTLA-4, PD-1, TIM-3, NKP30, NKP44, NKP40, NKP46, B7-H3, PD-L1, PD-2, and CD70.

In some embodiments, the one or more signaling domains include one or more of CD28, 4-1BB (CD137), CD8, CD40, OX40 (CD134), ICOS, CD27, DAP10, DAP12, OX40-L, 4-1BBL, ICOS-L, CD80, CD86, ICAM-1, LFA-1, CD56, CTLA-4, PD-1, PDK, TIM-3, NKP30, NKP44, NKP40, NKP46, B7-H3, PD-L1, PD-2, CD70, CD3ζ, and FcεRIγ domains, or any combination of two or more thereof. In a specific non-limiting example, the one or more signaling domains are CD28, 4-1BB, and CD3ζ signaling domains, for example including or consisting of the amino acid sequences of SEQ ID NOs: 8, 9, and 10, respectively. In some embodiments, the chimeric antigen receptor further includes a signal peptide located N-terminal to the scFv. In some examples, the signal peptide is a CD8α signal peptide, for example including or consisting of the amino acid sequence of SEQ ID NO: 5. Other signal peptides include an IgG signal sequence or a GM-CSF signal sequence.

In a particular example, the IL-13Rα2-CAR includes an amino acid sequence with at least 90% identity (for example, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98% at least 99% identity) to the amino acid sequence of SEQ ID NO: 3. In some examples, the IL-13Rα2-CAR includes or consists of the amino acid sequence of SEQ ID NO: 3. In other examples, the IL-13Rα2 CAR does not include the signal peptide, for example, does not include amino acids 1-21 of SEQ ID NO: 3.

In other embodiments, the IL-13Rα2-CAR further includes a domain that increases survival or persistence of an immune cell expressing the CAR. In some examples, the domain is an intracellular domain from a cytokine receptor, for example, an intracellular domain from interleukin (IL)-15 receptor, IL-12 receptor, or IL-18 receptor. In other examples, the domain is an intracellular domain from a growth factor receptor, such as an intracellular domain from CD40, NKG2D, NKP40, or NKP46. In some examples, the domain is located C-terminal to the CD3ζ domain of the CAR. In additional embodiments, the IL-13Rα2-CAR further includes an inducible gene (such as Caspase 9) that can be used to eliminate IL-13Rα2-CAR expressing cells (*e.g.*, a "suicide" gene). The inducible gene can be activated in the event of off target side effects (or on target/off tumor effects), such as cytokine release syndrome ("cytokine storm").

Also provided are functional variants of the IL-13Rα2-CARs or the domains thereof described herein, which retain the biological activity of the CAR of which it is a variant or retains the biological activity of the particular domain. The functional variant can be at least about 80%, about 85%, about 90%, about 91%, about 92%, about 93%, about 94%, about 95%, about 96%,

about 97%, about 98%, about 99% or more identical in amino acid sequence to the parent CAR or domain. Substitutions can be made, for example, in one or more of the extracellular targeting domain, hinge domain, transmembrane domain, and intracellular domains.

In some examples, the functional variant includes the amino acid sequence of the parent CAR or domain with at least one conservative amino acid substitution (such as up to 10 conservative amino acid substitutions, for example, 1, 2, 3, 4, 5, 6, 7, 8, 9, or 10 conservative substitutions). In other examples, the functional variant includes the amino acid sequence of the parent CAR or domain with at least one non-conservative amino acid substitution (such as up to 10 non-conservative amino acid substitutions, for example, 1, 2, 3, 4, 5, 6, 7, 8, 9, or 10 non-conservative substitutions). In this case, the non-conservative amino acid substitution does not interfere with or inhibit the biological activity of the functional variant. The non-conservative amino acid substitution may enhance the biological activity of the functional variant, such that the biological activity of the functional variant is increased as compared to the parent CAR or domain.

The CARs or domains thereof can in some examples, include one or more synthetic amino acids in place of one or more naturally occurring amino acids. Such synthetic amino acids include, for example, aminocyclohexane carboxylic acid, norleucine, a-amino n-decanoic acid, homoserine, S-acetylaminomethyl-cysteine, trans-3- and trans-4-hydroxyproline, 4- aminophenylalanine, 4-nitrophenylalanine, 4-chlorophenylalanine, 4-carboxyphenylalanine, β-phenylserine β-hydroxyphenylalanine, phenylglycine, α -naphthylalanine, cyclohexylalanine, cyclohexylglycine, indoline-2-carboxylic acid, 1 ,2,3,4- tetrahydroisoquinoline-3-carboxylic acid, aminomalonic acid, aminomalonic acid monoamide, N'-benzyl-N'-methyl-lysine, N',N'-dibenzyl-lysine, 6-hydroxylysine, ornithine, α-aminocyclopentane carboxylic acid, α-aminocyclohexane carboxylic acid, oc-aminocycloheptane carboxylic acid, -(2-amino-2-norbornane)-carboxylic acid, γ-diaminobutyric acid, α,β-diaminopropionic acid, homophenylalanine, and α-tert-butylglycine. The CARs may be glycosylated, amidated, carboxylated, phosphorylated, esterified, N-acylated, cyclized via, *e.g.*, a disulfide bridge, or converted into an acid addition salt and/or optionally dimerized or polymerized, or conjugated.

V. Nucleic Acids and Vectors

Also provided are nucleic acids encoding the IL13-Rα2 binding agents or IL-13Rα2 chimeric antigen receptors (IL-13Rα2-CAR) disclosed herein.

In some embodiments, the nucleic acid encodes a IL13-Rα2 binding agent. In some examples, the nucleic acid encodes each of the CDR sequences provided in Table 1, for example, the nucleic acid encodes the VH domain CDR1, CDR2, and CDR3 of SEQ ID NO: 1, and encodes

the VL domain CDR1, CDR2, and CDR3 of SEQ ID NO: 1, respectively. In some examples, the nucleic acid encodes an amino acid sequence including each of the CDR sequences provided in Table 1, and has at least 90% identity (for example, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98% or at least 99% sequence identity) to SEQ ID NO: 1. In some examples, the nucleic acid encodes an amino acid sequence including or consisting of SEQ ID NO: 1. In further examples, the nucleic acid encodes each of the CDR sequences provided in Table 1, and has at least 90% identity (for example, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98% or at least 99% sequence identity) to SEQ ID NO: 2. In some examples, the nucleic acid encoding the IL13-Rα2 binding agent includes or consists of SEQ ID NO: 2.

In some embodiments, the nucleic acid encodes the IL-13Rα2-CAR disclosed herein. In some examples, the nucleic acid molecule encodes an amino acid sequence having at least 90% sequence identity (for example, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98% or at least 99% sequence identity) to SEQ ID NO: 3. In some examples, the nucleic acid molecule encodes an amino acid sequence that includes each of the CDR sequences provided in Table 1 (the nucleic acid encodes the VH domain CDR1, CDR2, and CDR3 and the VL domain CDR1, CDR2, and CDR3 of SEQ ID NO: 1), and has at least 90% sequence identity (for example, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98% or at least 99% sequence identity) to SEQ ID NO: 3. In some examples, the nucleic acid molecule encodes an amino acid sequence including or consisting of SEQ ID NO: 3. In further examples, the nucleic acid molecule encoding the IL-13Rα2-CAR includes a nucleic acid sequence having at least 90% sequence identity (for example, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98% or at least 99% sequence identity) to SEQ ID NO: 4. In some examples, the nucleic acid molecule encoding the IL-13Rα2-CAR includes a nucleic acid sequence encoding each of the CDR sequences provided in Table 1, and the nucleic acid has at least 90% sequence identity (for example, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98% or at least 99% sequence identity) to SEQ ID NO: 4. In other examples, the nucleic acid molecule encoding the IL-13Rα2-CAR includes or consists of SEQ ID NO: 4.

Also provided are vectors (*e.g.*, plasmids, viral vectors, cosmids, artificial chromosomes) encoding a nucleic acid disclosed herein (*e.g.*, a nucleic acid encoding an IL-13Rα2-specific binding agent or an IL-13Rα2-CAR). Certain vectors are capable of directing the expression of genes to which they are operatively-linked. Such vectors are sometimes referred to as "expression vectors." In some embodiments, a nucleic acid molecule encoding a disclosed IL-13Rα2 scFv or

IL-13Rα2-CAR is included in a vector (such as a viral vector) for expression in a host cell, such as an immune cell (*e.g.*, T cell, NK cell, NKT cell, or macrophage).

One type of vector is a "plasmid," which refers to a circular double stranded DNA loop into which additional DNA segments can be inserted, such as by standard molecular cloning techniques. Another type of vector is a viral vector, wherein virally-derived DNA or RNA sequences are present in the vector for packaging into a virus (*e.g.*, retroviruses, replication defective retroviruses, adenoviruses, replication defective adenoviruses, and adeno-associated viruses). A replication deficient viral vector is a vector that requires complementation of one or more regions of the viral genome required for replication due to a deficiency in at least one replication-essential gene function.

In some embodiments, the vector is a viral vector, such as a retrovirus (*e.g.*, MoMLV or lentivirus) or adeno-associated viral (AAV) vector. Other suitable viral vectors include polyoma, SV40, vaccinia virus, herpes viruses including HSV and EBV, Sindbis viruses, alphaviruses and retroviruses of avian, murine, and human origin, baculovirus (Autographa californica multinuclear polyhedrosis virus; AcMNPV) vectors, retrovirus vectors, polio vectors, orthopox vectors, avipox vectors, fowlpox vectors, capripox vectors, suipox vectors, adenoviral vectors, herpes virus vectors, alpha virus vectors, baculovirus vectors, Sindbis virus vectors, vaccinia virus vectors and poliovirus vectors. Specific exemplary pox viruses of use include orthopox, suipox, avipox, and capripox virus. Orthopox includes vaccinia, ectromelia, and raccoon pox. One example of an orthopox of use is vaccinia. Avipox includes fowlpox, canary pox and pigeon pox. Capripox includes goatpox and sheeppox. In one example, the suipox is swinepox. In a specific, non-limiting example, the vector is pCDH-MSCV-MCS-EF1a-GFP-T2A-Puro lentiviral vector (*e.g.*, Cat# CD713B-1, System Biosciences, Palo Alto, CA 94303).

Vectors can include one or more regulatory elements (*e.g.*, promoter (such as CMV, SV40, EF1α, β-actin, hPGK, or RPBSA); transcription start site (TSS); enhancers; insulators; A/T-rich regions; transcription factor (TF) binding sites; transcription and/or translation terminators; initiation sequences, or other regulatory element), which may be selected on the basis of the host cell to be used for expression, that is operatively-linked to the nucleic acid sequence to be expressed. "Operably linked" means that the nucleotide sequence of interest is linked to the regulatory element(s) in a manner that allows for expression of the nucleotide sequence (*e.g.*, in an *in vitro* transcription/translation system or in a host cell when the vector is introduced into the host cell). Vector expression can be constitutive (*e.g.*, SV40, CMV, UBC, EF1α, PGK, and CAGG), inducible (*e.g.*, IPTG), or cell/tissue specific (such as T cell specific, *e.g.*, dLck or CD3δ). In some examples, the promoter is a synthetic promoter that is inducible upon CAR T cell or TCR activation

(*e.g.*, "iSynPro" promoters, *see*, WO 2018/213332). A vector can be introduced into a host cell to express a product encoded by a nucleic acid described herein (*e.g.*, an IL13-Rα2 binding agent or IL-13Rα2-CAR). A vector can also include one or more selectable marker genes, such as an antibiotic (e.g., puromycin, hygromycin), or a detectable marker (*e.g.*, GFP, YFP, RFP, luciferase, X-gal). In some examples, a selectable marker or reporter is not included in the vector.

Vectors can include a safety switch system, such as an inducible proapoptotic molecule (*e.g.,* Fas-associated death domain–containing protein (FADD), Bcl-2-associated death promoter (BAD), or inducible caspase 9 (iCasp9)), or an inducible activation system (*e.g.*, My88/CD40 (iMC)). Further information on safety switch systems can be found, for example, in Gargett and Brown (2014) *Front. Pharmacol.* 5: 235 and Gerken *et al.* (2017) *Cancer Discov.* 7(11):1306-1319. In some examples, the vector encoding the safety switch is a different vector than the vector encoding an IL-13Rα2-CAR or IL13-Rα2 binding agent disclosed herein. In some examples, the vector encoding the safety switch also encodes the IL-13Rα2-CAR or IL13-Rα2 binding agent.

In some examples, the safety switch is a proapoptotic molecule fused with a FKBP (FK506-binding protein) variant that binds a chemical inducer of dimerization (CID) (*e.g.,* AP1903, AP20187, AP21967). In some examples, a proapoptotic molecule is activated to control growth or eliminate cells transduced or transformed with a vector, such as a vector encoding an IL-13Rα2-CAR or IL13-Rα2 binding agent disclosed herein. In some examples, activation of the inducible proapoptotic molecule eliminates the transduced or transformed cells, for example, by eliminating at least 25%, at least 50%, at least 60%, at least 70%, at least 80%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98% at least 99%, or 100% of the transduced or transformed cells. In some examples, the proapoptotic molecule is activated by contacting a transduced or transformed cell with a CID.

In some examples, the safety switch is an inducible activation system (*e.g.*, My88/CD40 (iMC)). In some examples, the inducible activation system is activated to increase activity of a cell transduced or transformed with a vector, such as a vector encoding an IL-13Rα2-CAR or IL13-Rα2 binding agent disclosed herein. In some examples, activation of the inducible activation system increases activity of transformed cells, for example, by increasing proliferation, cytokine production, or tumor targeting by at least 25%, at least 50%, at least 60%, at least 70%, at least 80%, at least 90%, at least 100%, at least 200%, at least 400%, or more. In some examples, the inducible activation system is activated by contacting a transformed cell with a CID (*e.g.*, AP1903, AP20187, AP21967).

VI. Cells Expressing IL-13Rα2 CARs

Also provided herein are cells (for example, immune cells) that express a disclosed IL-13Rα2-CAR, such as cells that include a nucleic acid disclosed herein (*e.g.*, a nucleic acid encoding an IL13-Rα2 binding agent or IL-13Rα2-CAR disclosed herein), or a vector encoding the nucleic acid. Exemplary immune cells include T cells, NK cells, NKT cells, and/or macrophages. In some examples, the cells are T cells. In some examples, the cells express an IL-13Rα2-CAR having at least 90% identity to SEQ ID NO: 3, or amino acids 22-538 of SEQ ID NO: 3. In some examples, the cells express an IL-13Rα2-CAR including or consisting of SEQ ID NO: 3, or amino acids 22-538 of SEQ ID NO: 3. In some examples, the cells are transduced or transfected with a nucleic acid sequence encoding an amino acid sequence including or consisting of SEQ ID NO: 3, or amino acids 22-538 of SEQ ID NO: 3. In further examples, the cells are transduced or transfected with a nucleic acid sequence having at least 90% identity to SEQ ID NO: 4. In some examples, the nucleic acid sequence includes or consists of SEQ ID NO: 4.

In some embodiments, the cells are isolated immune cells, such as isolated T cells (such as a primary T cell or T cells obtained from a subject), isolated NK cells (such as a primary NK cell or NK cells obtained from a subject), isolated NKT cells, or isolated macrophages. Cell types can be identified based on the presence or absence of expression markers, for example, NK cells are CD56 positive and CD3 negative, T cells can be CD4 or CD8 positive, NKT cells are NK1.1 positive and α/β T-cell receptor positive, and macrophages express a number or receptors, including PRRs, TLRs, C-type lectin receptors, scavenger receptors, RLRs, and NOD-like receptors. In some examples, the cells are isolated T cells. In further examples, the cells are isolated CD8+ T cells. In some examples, the cells are isolated naïve T cells. Naïve T cells express CD45RA, CCR7, CD62L, CD127, and CD132 surface markers, and lack CD25, CD44, CD69, CD45RO, or HLA-DR markers. In some examples, the immune cells are obtained from peripheral blood, lymph node, thymus, bone marrow, tumor tissue, adipose tissue, human embryonic stem cells (hESC), induced pluripotent stem cells (iPSC), or umbilical cord blood, of a subject. In a non-limiting example, the cells are T cells isolated from peripheral blood of a subject. In further examples, the immune cells obtained from the subject are enriched, purified, and/or expanded, for example before and/or after transformation or transduction with a vector or nucleic acid disclosed herein. In some examples, the immune cells (*e.g.*, T cell, NK cell, NKT cell, or macrophage), are harvested and expanded from a subject with cancer. In some examples, the immune cells (*e.g.*, T cell, NK cell, NKT cell, or macrophage), are harvested and expanded from a subject without cancer.

In some embodiments, the cells are a cell type suitable for protein expression and/or production of viruses (*e.g.*, producing retroviruses). Exemplary cell types include microbial (*e.g.*, bacterial or yeast), archea, insect, fungi, plant, or mammalian cells. Non-limiting examples of

specific cell lines include *Escherichia coli, Bacillus subtilis, Saccharomyces cerevisiae*, *Salmonella typhimurium,* SF9 cells, C129 cells, 293 cells, *Neurospora*, and immortalized mammalian myeloid and lymphoid cell lines. Non-limiting examples of commonly used mammalian host cell lines include VERO and HeLa, HEK 293T, CHO, WI38, BHK, and COS cell lines, however, other cell lines may be used, such as cells designed to provide higher expression, or desirable post-translational modifications (*e.g.*, glycosylation patterns), or other features. In some examples, a nucleic acid or vector disclosed herein (*e.g.*, nucleic acids or vectors encoding the IL13-Rα2 binding agent or IL-13Rα2-CAR disclosed herein) is codon optimized for expression in a particular organism, such as a bacterium (*e.g.*, *E. coli*), fungi (*e.g.*, *S. cerevisiae)*, plant (*e.g.*, *N. tabacum* NT-1), or animal (*e.g.*, human HEK 293T cells or T cells). Suitable examples of vectors (*e.g.*, expression vectors) are provided above.

In some embodiments, the cells are transduced or transformed (including transfected) with a nucleic acid encoding the IL13-Rα2 binding agent or IL-13Rα2-CAR disclosed herein, or a vector encoding the nucleic acid. Exemplary methods of transforming cells (*e.g.*, immune cells) include chemical transformation (calcium phosphate), electroporation, microinjection, heat shock, lipofection, and particle bombardment. In some examples, the nucleic acid or vector is introduced by contacting the cells with a nanoparticle including the nucleic acid or vector. Transformation can be used for stable or transient expression. Following transformation or transduction, cells expressing the IL-13Rα2-CAR can be detected and/or enriched, for example, by flow cytometry using a labeled antibody that binds to IL-13Rα2. In some examples, transduced or transformed cells (such as T cells, NK cells, NKT cells, or macrophages) are expanded, for example, by cell culture for a period of time following transformation or transduction. In some examples, some or all of the transduced or transformed immune cells are cryopreserved for later use.

VII. Methods of Immunotherapy

Also disclosed herein are methods of treating a subject with cancer, comprising administering an effective amount of cells expressing a disclosed IL-13Rα2-CAR (*e.g.*, immune cells expressing IL-13Rα2-CAR, such as IL-13Rα2-CAR-T cells, IL-13Rα2-CAR-NK cells, IL-13Rα2-CAR-NKT cells, or IL-13Rα2-CAR-macrophage cells). In a specific, non-limiting example, an effective amount of an IL-13Rα2-CAR-T cell is administered to a subject. The cells expressing a disclosed IL-13Rα2-CAR can be generated from immune cells (*e.g.*, T cells, NK cells, NKT cells, or macrophages) isolated from a sample, for example, a peripheral blood sample of a subject, and subsequently transformed or transduced with the IL-13Rα2-CAR. In some examples, the cells are autologous to the subject (from the subject). In other examples, the cells are allogeneic

(from a donor). In some examples, the sample is from a subject that has cancer. In some examples, the cells are enriched, purified, and/or expanded after isolation from the sample and/or after transformation or transfection with the IL-13Rα2-CAR.

In some embodiments, the methods include administering to a subject a composition including cells expressing the IL-13Rα2-CAR and a pharmaceutically acceptable carrier. In some examples, the cells include a vector encoding a safety switch system. In some embodiments, the methods include administering to the subject a pharmaceutical composition including a vector encoding the IL-13Rα2-CAR and a pharmaceutically acceptable carrier. In some examples, the vector encodes a safety switch system.

A "pharmaceutically acceptable carrier" includes any and all solvents, dispersion media, coatings, antibacterial and antifungal agents, isotonic and absorption delaying agents, and the like, compatible with pharmaceutical administration (see, *e.g., Remington: The Science and Practice of Pharmacy*, Adejare (Ed.), Academic Press, London, United Kingdom, 23rd Edition (2021). Examples of such carriers or diluents include, but are not limited to, water, saline, Ringer's solutions, dextrose solution, balanced salt solutions, and 5% human serum albumin. Liposomes and non-aqueous vehicles such as fixed oils may also be used. Supplementary active compounds can also be incorporated into the compositions. Actual methods for preparing administrable compositions include those provided in *Remington: The Science and Practice of Pharmacy*, Adejare (Ed.), Academic Press, London, United Kingdom, 23rd Edition (2021).

In some examples, the composition includes about 10^4 to 10^{12} of the IL-13Rα2-CAR cells (for example, about 10^4-10^8 cells, about 10^6-10^8 cells, or about 10^6-10^{12} cells). In some examples, the composition includes at least about 10^4, 10^5, 10^6, or 10^7 IL-13Rα2-CAR cells. In other examples, the composition includes no more than about 10^4, 10^5, 10^6, 10^7 10^8, 10^9, or 10^{10} IL-13Rα2-CAR cells. In some examples, the composition may be prepared such that about 10^4 to 10^{10} IL-13Rα2-CAR cells/kg (such as about 10^4, 10^5, 10^6, 10^7, 10^8, 10^9, or 10^{10} cells/kg) are administered to a subject. Appropriate doses can be determined through clinical trials.

The population of IL-13Rα2-CAR cells is typically administered parenterally, for example intravenously; however, injection or infusion to a tumor or close to a tumor (*e.g.*, local administration) or administration to the peritoneal cavity can also be used. Appropriate routes of administration can be determined by a skilled clinician based on factors such as the subject, the condition being treated, and other factors.

Multiple doses of the population of IL-13Rα2-CAR cells can be administered to a subject. For example, IL-13Rα2-CAR cells can be administered daily, every other day, twice per week, weekly, every other week, every three weeks, monthly, or less frequently. A skilled clinician can

select an administration schedule based on the subject, the condition being treated, the previous treatment history, and other factors.

In some examples, the effective amount of cells expressing a disclosed IL-13Rα2-CAR is an amount sufficient to prevent, treat, reduce, and/or ameliorate one or more signs or symptoms of cancer in the subject. In other examples, the effective amount is an amount sufficient to target and eliminate tumor cells expressing IL-13Rα2. In some examples, the effective amount is an amount sufficient to inhibit or slow cancer growth or metastasis in the subject. In another example, the effective amount is an amount sufficient to reduce tumor load or tumor density in the subject.

In some embodiments, the subject is further administered one or more cytokine(s) (such as one or more of IL-2, IL-7, IL-15, IL-21, and/or IL-12), for example, to support survival and/or growth of the IL-13Rα2-CAR cells. The cytokine(s) can be administered before, after, or substantially simultaneously with the IL-13Rα2-CAR cells. In some examples, about 1 to about 5 cytokines are administered to the subject, for example, about 1 to about 4, about 1 to about 3, about 1 to about 2, about 2 to about 5, about 2 to about 4, about 2 to about 3, about 3 to about 5, about 3 to about 4, or about 4 to about 5 cytokines are administered, before, after, or substantially simultaneously with the IL-13Rα2-CAR cells. In some examples, at least one of IL-2, IL-7, or IL-15 is administered to the subject before, after, or substantially simultaneously with the IL-13Rα2-CAR cells. In some examples, at least one cytokine (*e.g.*, IL-2, IL-7, and/or IL-15) is administered simultaneously, for example, with IL-13Rα2-CAR cells.

In some examples, the cells expressing a disclosed IL-13Rα2-CAR (*e.g.*, IL-13Rα2-CAR-T cells, IL-13Rα2-CAR-NK cells, IL-13Rα2-CAR-NKT cells, or IL-13Rα2-CAR-macrophage cells) are transformed or transfected with a vector encoding a safety switch system. In some examples a therapeutically effective amount of a CID (*e.g.*, AP1903, AP20187, AP21967) is administered to the subject to activate the safety switch system. The safety switch system can be activated at any desired time, for example, at the conclusion of treatment. In some examples, the CID is administered to control growth or to increase activity of the transformed or transfected cells in the subject. In some examples, the safety switch is an inducible proapoptotic molecule, and the inducible proapoptotic molecule is activated by administering a therapeutically effective amount of a CID to the subject. In some examples, administering the CID eliminates the transformed or transfected cells in the subject, for example by eliminating at least 50%, at least 60%, at least 70%, at least 80%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98% at least 99%, or 100% of the transformed or transfected cells in the subject. In other examples, the safety switch is an inducible activation system, and the inducible activation system is activated by administering an effective amount of a CID to the

subject. In some examples, administering the CID increases activity of the transformed or transfected cells in a subject as compared to a suitable control. For example, by increasing proliferation, cytokine production, or tumor targeting by at least 50%, at least 60%, at least 70%, at least 80%, at least 90%, at least 100%, at least 200%, at least 400%, or more of the transformed or transfected cells in the subject. Further information on safety switch systems can be found, for example, in Gargett and Brown (2014) *Front. Pharmacol.* 5: 235 and Gerken *et al.* (2017) *Cancer Discov.* 7(11):1306-1319.

In some examples, the subject has a cancer that expresses IL-13Rα2. Exemplary cancers include sarcomas, carcinomas, fibrosarcoma, myxosarcoma, liposarcoma, chondrosarcoma, osteosarcoma, and other sarcomas, synovioma, mesothelioma, Ewing's tumor, Kaposi sarcoma, leiomyosarcoma, rhabdomyosarcoma, colon carcinoma, lymphoid malignancy, pancreatic cancer, breast cancer, lung cancers, ovarian cancer, uterine cancer, prostate cancer, hepatocellular carcinoma, squamous cell carcinoma, basal cell carcinoma, adenocarcinoma, adrenal carcinoma, sweat gland carcinoma, medullary thyroid carcinoma, papillary thyroid carcinoma, pheochromocytomas sebaceous gland carcinoma, papillary carcinoma, papillary adenocarcinomas, medullary carcinoma, bronchogenic carcinoma, renal cell carcinoma, hepatoma, bile duct carcinoma, choriocarcinoma, Wilms' tumor, cervical cancer, testicular tumor, seminoma, bladder carcinoma, melanoma, and CNS tumors (such as a glioma (such as brainstem glioma and mixed gliomas), glioblastoma (also known as glioblastoma multiforme) astrocytoma, CNS lymphoma, germinoma, medulloblastoma, Schwannoma craniopharyogioma, ependymoma, pinealoma, hemangioblastoma, acoustic neuroma, oligodendroglioma, menangioma, head and neck cancers, neuroblastoma, retinoblastoma and brain metastasis. Cancer also includes hematological (or hematogenous) cancers, such as leukemia, such as lymphoma, Hodgkin's disease, non-Hodgkin's lymphoma (indolent or high grade forms), multiple myeloma, Waldenstrom's macroglobulinemia, heavy chain disease, myelodysplastic syndrome, hairy cell leukemia or myelodysplasia.

In some examples, the subject has a cancer that over-expresses IL-13Rα2, for example, relative to a non-cancerous cell of the same type. In some examples, the subject has a solid tumor. In further examples, the subject has pancreatic cancer, glioblastoma, head and neck squamous cell carcinoma, ovarian cancer, uterine cancer, prostate cancer, breast cancer, melanoma, non-small cell lung cancer (NSCLC), renal cell carcinoma, Kaposi sarcoma, or adrenal carcinoma. In some embodiments, a subject having a cancer that expresses or over-expresses IL-13Rα2 is selected for treatment.

In some embodiments, the subject receives an additional treatment, such as one or more of surgery, radiation, chemotherapy, additional immunotherapy, or other therapeutic. Exemplary

chemotherapeutic agents include (but are not limited to) alkylating agents, such as nitrogen mustards (such as mechlorethamine, cyclophosphamide, melphalan, uracil mustard or chlorambucil), alkyl sulfonates (such as busulfan), nitrosoureas (such as carmustine, lomustine, semustine, streptozocin, or dacarbazine); antimetabolites such as folic acid analogs (such as methotrexate), pyrimidine analogs (such as 5-FU or cytarabine), and purine analogs, such as mercaptopurine or thioguanine; or natural products, for example vinca alkaloids (such as vinblastine, vincristine, or vindesine), epipodophyllotoxins (such as etoposide or teniposide), antibiotics (such as dactinomycin, daunorubicin, doxorubicin, bleomycin, plicamycin, or mitocycin C), and enzymes (such as L-asparaginase). Additional agents include platinum coordination complexes (such as cis-diamine-dichloroplatinum II, also known as cisplatin), substituted ureas (such as hydroxyurea), methyl hydrazine derivatives (such as procarbazine), and adrenocrotical suppressants (such as mitotane and aminoglutethimide); hormones and antagonists, such as adrenocorticosteroids (such as prednisone), progestins (such as hydroxyprogesterone caproate, medroxyprogesterone acetate, and magestrol acetate), estrogens (such as diethylstilbestrol and ethinyl estradiol), antiestrogens (such as tamoxifen), and androgens (such as testosterone proprionate and fluoxymesterone). Examples of the most commonly used chemotherapy drugs include adriamycin, melphalan (Alkeran®) Ara-C (cytarabine), carmustine, busulfan, lomustine, carboplatinum, cisplatinum, cyclophosphamide (Cytoxan®), daunorubicin, dacarbazine, 5-fluorouracil, fludarabine, hydroxyurea, idarubicin, ifosfamide, methotrexate, mithramycin, mitomycin, mitoxantrone, nitrogen mustard, paclitaxel (or other taxanes, such as docetaxel), vinblastine, vincristine, VP-16, while newer drugs include gemcitabine (Gemzar®), trastuzumab (Herceptin®), irinotecan (CPT-11), leustatin, navelbine, rituximab (Rituxan®) imatinib (STI-571), Topotecan (Hycamtin®), capecitabine, ibritumomab (Zevalin®), and calcitriol. A skilled clinician can select appropriate additional therapies (from those listed here or other current therapies) for the subject, depending on factors such as the subject, the cancer being treated, treatment history, and other factors.

In some examples, the subject is administered an additional therapeutic, such as a checkpoint inhibitor (*e.g.*, anti-CTLA-4, anti-PD1, or anti-PDL1), a histone deacetylase (HDAC) inhibitor, a cell cycle checkpoint inhibitor, adrenomedullin, an IL-13-PE immunotoxin, or any combination of two or more thereof. The administration of an additional therapeutic may be before, after, or substantially simultaneously with the administration of the cell expressing the IL-13Rα2-CAR.

In some examples, the additional therapeutic increases expression of IL-13Rα2 on tumor cells. For example, it has been reported that treatment with HDAC inhibitors dramatically

upregulates IL-13Rα2 in pancreatic cancer cell lines expressing little to no IL-13Rα2. These inhibitors also modestly upregulated IL-13Rα2 in cells expressing higher levels of IL-13Rα2. Upregulation of IL-13Rα2 was found to sensitize pancreatic tumor cells to IL-13-PE, an immunotoxin that targets cells expressing IL-13Rα2, resulting in a synergistic anti-tumor effect (*see* Fujisawa *et al. Journal of Translational Medicine* 9:37, 2011). Thus, in some examples, the subject is administered an HDAC inhibitor, such as one or more of trichostatin A (TSA), suberoylanilide hydroxamic acid (SAHA), sodium butyrate (NaB), SP600125 (Sigma-Aldrich), SR11302 (Tocris Bioscience), or romidepsin. In further examples, the subject is administered adrenomedullin, which has also been shown to increase IL-13Rα2 expression (Joshi *et al., Cancer Res.* 68:9311-9317, 2008). In some examples, the HDAC inhibitor or adrenomedullin is administered substantially simultaneously with the cells expressing the IL-13Rα2-CAR. In other examples, the HDAC inhibitor or adrenomedullin is administered prior to administering the cells expressing the IL-13Rα2-CAR, for example, at least 1 day, at least 2 days, at least 3 days, at least 4 days, at least 5 days, at least 6 days, at least 7 days, at least 8 days, at least 9 days, at least 12 days, at least 14 days, at least three weeks, at least four weeks, at least one month, or more prior. Multiple doses of the HDAC inhibitor or adrenomedullin can be administered to a subject, for example, administered twice daily, once daily, every other day, twice per week, weekly, every other week, every three weeks, monthly, or less frequently. A skilled clinician can select an administration schedule based on the subject, the condition being treated, the previous treatment history, tumor load and type, clinical stage and grade of the disease, overall health of the subject, and other factors.

In some examples, the subject is administered a checkpoint inhibitor. In some examples, the checkpoint inhibitor targets PD-1, PD-L1, CTLA-4, CDK4, and/or CDK6. Exemplary inhibitors include ipilimumab, nivolumab, pembrolizumab, atezolizumab, avelumab, durvalumab, cemiplimab, palbociclib, ribociclib, and abemaciclib. The checkpoint inhibitor may be administered substantially simultaneously with the cells expressing the IL-13Rα2-CAR. In some examples, the checkpoint inhibitor is administered prior to administering the cells expressing the IL-13Rα2-CAR, for example, at least 1 day, at least 2 days, at least 3 days, at least 4 days, at least 5 days, at least 6 days, at least 7 days, at least 8 days, at least 9 days, at least 12 days, at least 14 days, at least three weeks, at least four weeks, at least one month, or more prior. Multiple doses of the checkpoint inhibitor can be administered to a subject, for example, administered twice daily, once daily, every other day, twice per week, weekly, every other week, every three weeks, monthly, or less frequently. A skilled clinician can select an administration schedule based on the subject, the

condition being treated, the previous treatment history, tumor load and type, clinical stage and grade of the disease and overall health of the subject, and other factors.

In some embodiments, the subject is administered an IL-13 PE immunotoxin, such as a recombinant protein including IL-13 conjugated to a truncated *Pseudomonas* exotoxin (*see*, *e.g.*, Kioi *et al., Mol. Cancer Ther.* 7(6):1579-1587, 2008). The IL-13 PE immunotoxin may be administered substantially simultaneously with the cells expressing the IL-13Rα2-CAR. In some examples, the IL-13 PE immunotoxin is administered prior to administering the cells expressing the IL-13Rα2-CAR, for example, at least 1 day, at least 2 days, at least 3 days, at least 4 days, at least 5 days, at least 6 days, at least 7 days, at least 8 days, at least 9 days, at least 12 days, at least 14 days, at least three weeks, at least four weeks, at least one month, or more prior. Multiple doses of the IL-13 PE immunotoxin can be administered to a subject, for example, administered twice daily, once daily, every other day, twice per week, weekly, every other week, every three weeks, monthly, or less frequently. A skilled clinician can select an administration schedule based on the subject, the condition being treated, the previous treatment history, tumor load and type, clinical stage and grade of the disease and overall health of the subject, and other factors. Without being bound by any particular theory, it is believed that, administering IL-13-PE prior to the administration of the IL-13Rα2-CAR cells reduces density of tumor stroma cells, thereby creating space in the tumor microenvironment for IL-13Rα2-CAR cells to infiltrate the tumor.

EXAMPLES

The following examples are provided to illustrate certain particular features and/or embodiments. These examples should not be construed to limit the disclosure to the particular features or embodiments described.

Example 1

Materials and Methods

Cell lines:

Human Jurkat-T cells, A172, U87MG and T98G glioma cells lines were obtained from ATCC and grown according to the supplier's instructions. The U251 glioma cell line was obtained from the National Cancer Institute and maintained in RPMI complete medium with 10% FBS. The T98G and U87MG glioma cell lines were maintained in EMEM complete medium supplemented with 10% FBS. These cell lines were previously characterized for IL-13Rα2 expression by RT-PCR of mRNA and immunocytochemistry (ICC) analyses of protein expression (Joshi *et al.* (2000) *Cancer Res.* 60:1168-1172). Two IL-13Rα2 positive glioma cells lines, U251 and U87MG were

used for IL-13Rα2 gene silencing by siRNA technique using SureSilencing® shRNA plasmids (Qiagen (Gaithersburg, MD)) following the manufacturer's instructions. These cell lines served as negative controls in some biological assays.

Design of lentiviral vector encoding scFv-IL-13Rα2-CAR:

A third-generation CAR construct consisting of a single chain Fv (scFv) antibody sequence against IL-13Rα2 (as ectodomain), a CD28 transmembrane domain along with CD3ζ and CD28 and 4-1BB endodomain sequences was designed, codon-optimized, and synthesized by GenScript® (GenScript, NJ) in pUC57 simple subcloning vector. The novel scFv was previously cloned using the Griffin.1 library, a scFv library made from synthetic v-gene segments, and derived by recloning VH and VL human synthetic Fab Lox library vectors into the phagemid vector pHEN2 as previously described (Kioi *et al.* (2008) *Mol Cancer Ther.* 7:1579-1587). A novel approach to re-panning the segment to improve binding affinity to huIL-13Rα2Fc chimeric protein was used. The CAR-T construct was codon optimized and flanked by Bam HI and NotI sites and placed into pCDH-MSCV-MCS-EF1-copGFP-T2A-Puro lentiviral vector (System Biosciences, MA). This transfer plasmid was packaged into 293T cells by co-transfecting with three helper plasmids (pCD/NL-BHΔ1, pCEF-VSV-G and pCMV-Rev) to produce self-inactivating (SIN) lentiviral vector expressing CAR-IL13Rα2-CAR pseudo-lentivector, which was further purified by ultracentrifugation.

Analysis of antigen binding residues and complementary determining regions (CDR) in scFv IL-13Rα2:

Standardized numbering methods were used to define CDRs, frameworks and residues from the light and heavy chains that have an impact on the interaction and/or affinity of the antibody for its target antigen. Amino acid sequences were analyzed using common numbering schemes of antibody variable domains to study the statistical variability in amino acid composition using the Kabat numbering scheme (ncbi.nlm.nih.gov/igblast). An analysis of residue distribution for both the heavy and light chains was performed, as well as an analysis of CDR sequences, including their lengths.

These data were further analyzed and compared with a structure-based numbering scheme for antibody variable regions, which formed the CDRs and corrected the position numbers of the points within CDRL1 and CDRH1 per Chothia numbering scheme, Martin numbering scheme, which was used for correction of the insertion point within the framework region of heavy and light chains, and ImMunoGeneTics® (IMGT®, The International ImMunoGeneTics® information

system, Montpellier, France) numbering scheme, which highlights protein sequences of the immunoglobin superfamily including variable domains from antibody light and heavy chains as well as T cell receptor chains (imgt.org; github.com/oxpig/ANARCI, academic.oup.com/bioinformatics/article/32/2/298/1743894; abysis.org/abysis/sequence_input/key_annotation/key_annotation.cgi; imgt.org/3Dstructure-DB/cgi/Collier-de-Perles.cgi; and chemogenomix.com).

Manufacturing of CAR-T cells:

Human PBMCs were isolated from buffy coats of normal healthy blood donors who donated blood at the Division of Transfusion Medicine, NIH. CAR-T cells were generated from CD4 and CD8 positive T cells isolated from the normal human blood donor buffy coat using a StraightFrom® Buffy Coat kit (Miltenyi Biotec, Waltham, MA). For CD4/CD8 T cell activation, the cells were activated with anti-CD3/CD28 antibody coated magnetic Dynabeads™ (Invitrogen, Carlsbad, CA) at a ratio of 3:1 (T cell to bead) and transduced with lentiviral vector at different multiplicity of infection (m.o.i). T cells were maintained in culture at 0.6-1 X 10^6 cells/mL in T cell culture medium (TCM) supplemented with 50 ng/ml IL-2 and 10 ng/ml each IL-7 and IL-15 (Miltenyi Biotec, Waltham, MA). These PBMC derived CAR-T cells are termed as CAR-T throughout the manuscript.

A parallel set of experiments was used for transduction and expansion to confirm the identity of transgene and signaling domains in human Jurkat T cells (termed CAR-Jurkat).

Indirect Immunofluorescence Assay for transgene in transduced CAR-Jurkat and CAR-T cells:

For detection of scFv-IL-13Rα2 expression on transduced Jurkat or T cells, a novel indirect immunofluorescence assay by plating 75,000 either Jurkat-CAR or CAR-T cells in a poly-L-Lysine coated 4-well chamber slide was used. Briefly, biotinylated recombinant human IL-13Rα2Fc chimera protein (R&D Biosystems, Minneapolis, MN) was produced using the EZ-Link™ micro sulfo-NHS-Biotinylation kit (Thermo Scientific, Waltham, MA). Transduced Jurkat and T cells were then incubated with 500 ng/ml purified biotinylated recombinant human IL-13Rα2Fc chimera protein followed by streptavidin-Alexa 594® (0.5 μg/ml) to develop red fluorescence in scFv-IL-13Rα2 expressing cells. The cells expressing ≥ 2+ fluorescence intensity were counted at 200X magnification using a Nikon® epifluorescence microscope. Each value is mean ± SD of quadruple experiments determined in a blinded manner and expressed as % positive cells.

Flow-cytometry analysis:

BD® FACSCanto™ or FACSCalibur™ (BD Bioscience, San Jose, CA) instruments were used to acquire immunofluorescence data which were analyzed with CellQuest™ (BD Bioscience) or FlowJo® v.7 (FlowJo, LLC Ashland, OR) software for final data analysis and graphic representation. Isotype control was immunoglobulin IgG1-PE (IgG1-PE, Cell Signaling Technology, Denver, MA). CD28 cytoplasmic and CD3ζ endodomains were immunostained with anti-CD28.2 mouse mAb PE conjugate or CD3ζ monoclonal antibody PE conjugate (Thermofisher eBioscience, Carlsbad, CA). Permeabilized CAR-Jurkat or CAR-T cells were immunostained with anti-CD28 and CD3ζ antibodies. The data were expressed as normalized to mode values.

Assessment of T cell activation marker expression in CAR-T cells:

T cell activation of CAR-T cells was evaluated by intracellular expression of CD44, CD25, CD69 and interferon-γ as T cell activation markers after treating the cells with brefeldin A (BioLegend, San Diego, CA), by indirect immunofluorescence assay (IFA, abCAM, Cambridge, MA). Each value is expressed as mean ± SD of four independent readings scored in a blinded fashion for % positive cells expressing ≥ 2+ immunofluorescence intensity.

Cell Viability and proliferation analysis of CAR-T cells:

Cell viability of CAR-T cell cultures was examined during cell expansion by trypan blue exclusion technique. To further assess the health of CAR-T cells, cellular proliferation was assessed by the CellTiter 96R® AQueous one solution kit (Promega, Madison, WI). Fisher, Waltham, MA, USA). A known number of CAR-T cells (2,500 cells/well) were plated in quadruple wells of a 96-well culture plate and maintained at 37°C in a CO_2 incubator. Twenty microliters of MTS reagent was added in each well on day 3, 5, and 7. The number of proliferating cells in each well was determined by measuring the optical density of reduced MTS tetrazolium by measuring the absorbance at 490 nm. The experiments were performed in quadruplicate and the results were expressed as mean ± SD.

Analysis of cell migration and invasion potential of CAR-T cells

The cell migration and invasion potential of CAR-T cells was assessed in 24-well ChemoTx® plates with a 5-μm pore diameter (abCam, Cambridge, MA) (chemotaxis assay). In the lower chambers, 600 μL of unconditioned Dulbecco's Modified Eagle Medium with 10, 50 and

1000 ng/ml hU-IL-13Rα2Fc (R&D Systems), U251-or T98G tumor cell culture conditioned Dulbecco's Modified Eagle Medium with 1% fetal calf serum or control media were added. The upper chambers were loaded with 500,000 CAR-T cells/200 µl. After 6 and 20 hours at 37°C, residual cells were scraped off the polycarbonate filter, and the plate was centrifuged for 2 min at 400 X g. The filter was removed, and cells in the lower chamber were counted by trypan blue exclusion technique. Percentage migration was calculated as the number of cells in the lower chamber divided by the total number of cells plated per well. Each value is expressed as mean ± SD of four independent experiments.

Cytotoxic Activity:

To determine the cell killing activity of CAR-T cells, a robust homogeneous fluorescence-based non-isotopic cytotoxicity assay was performed. U251 and U87MG glioma cell lines, which are positive for IL-13Rα2 expression, were tested. In addition, IL-13Rα2 gene was silenced by SureSilencing™ shRNA plasmid technique using following siRNA sequence GCTACCATTTGGTTTCATCTT (SEQ ID NO: 17) for transfection of U251 and U87 MG cell lines as per manufacturer's protocol (cat# 336313 KH00597N, QIAGEN, Germantown, MD). The IL-13Rα2 positive and IL-13Rα2-silenced tumor cells were labeled by intracellular Calcein violet-acetoxymethyl ester (Lichtenfels *et al.* (1994) *J Immunol Methods* 172:227-239; Neri *et al.* (2001) *Clin Diagn Lab Immunol* 8:1131-1135). Release of Calcein Violet in the supernatants recovered at the end of 6 hour of co-culture of target:effector cells in the ratio of 1:10, 1:20, 1:30, 1:40 and 1:50 was measured quantitatively in a fluorescent plate reader. The data are shown as mean ± SD of four independent experiments performed in quadruplicate involving co-cultures of target and effector CAR-T cells.

IFN-γ Release:

IL13Rα2-CAR-T cells (100,000) were co-cultured in equal part to IL-13Rα2 positive, IL-13Rα2 negative, or IL-13Rα2 KO tumor cells for 20 hours in a 96 well round bottom plate. At the end of the incubation period, the cultures were centrifuged at 3,500 X g for 10 minutes and supernatants were harvested for quantitative determination of IFN-γ secretion by ELISA assay (BioLegend).

Statistical Analysis:

The data were compared using unpaired Student's t-test analyses. *P* values were calculated using GraphPad Prism® software (GraphPad Software, La Jolla, CA). *P* values of <0.05 were

considered a significant difference. Two-way analysis of variance was used to compare labeling conditions ($n = 4$) and the Wilcoxon test was used to obtain two-sided global P values for cell survival or proliferation ($n = 4$).

Example 2

Identification and Characterization of scFv-IL-13Rα2

The gene for scFv of IL-13Rα2 was identified using a scFv phase display library of synthetic v-gene segments and improved versions of VH and VL showing a hypervariable amino-acid structural composition from a human synthetic Fab Lox library. A novel conserved scFv fragment that provides optimum antigen binding based on amino acid residue analysis by four different schemes was selected. The analysis revealed structure-based scFv variable regions, defined loop structures that form the CDRs, and insertion points inside CDRs exhibiting a hypervariable amino acid composition. The clone derived protein showed >4-fold higher binding affinity to recombinant IL-13Rα2Fc protein as compared to the previously cloned scFv ECD-IL-13Rα2 (Kioi et al, *Mol Cancer Ther*, 7(6):1579-87, 2008).

The improved scFv-IL-13Rα2 is sequence-specific for higher binding affinity, and codon-optimized for better expression in human cells. Furthermore, since the improved scFv-IL-13Rα2 was generated from a library made from synthetic V-gene segments, its spectrum of binding to IL-13Rα2 is expected to be more specific as compared to scFvs derived from a single monoclonal antibody (*e.g.*, Balyasnikova, *et al, J Biol Chem.*, 287 (36):30215-30227, 2012).

The variable domain of IL-13Rα2 was analyzed by residue numbering schemes to define CDRs, frameworks, and amino acid residues from light and heavy chains, which may have influence on binding affinity and specificity. The analysis revealed 8 different regions with varying numbers of residues and lengths for the heavy chain (FIGS. 2 and 3A). The analyzed sequences exhibited variable lengths of gaps where insertions could only be included at precise positions. The distribution of residues in the heavy chain is shown in FIG. 3A. The data analysis further revealed that the amino acid residues 27-38 in ascending loop B (CDR1), 55-59 in ascending C loop, 62-65 in descending C loop C (CDR2), 105-110 in ascending loop F and 113-117 in descending loop F (CDR3) are structural constituents of heavy chain (FIG. 3B).

Analysis of the light chain showed 9 different regions, including a tail region of 275 amino acids (FIG. 5A). Similar to the heavy chain structural analysis for amino acid residues, light chain amino acids 24-29 in the ascending B loop and 36-39 in descending C loop belong to CDR1-light, 56-57 in the ascending C loop and 65-69 in the descending C loop to CDR2-light, and 105-108 in the ascending F loop and 114-117 descending G loop to CDR3-light, are conserved (FIG. 5B). The

distribution of amino acids in heavy chain and light chain after analyzing by Kabat numbering scheme is shown in FIGS. 6 and 7, respectively.

Kabat numbering of scFv residue sequence was also analyzed for a comparative alignment analysis with IMGT, Chothia, and Martin numbering schemes for antibody variable regions, loop structures that form the CDRs, position numbers of the insertion points within CDR-H(heavy) and CDR-L(light) chains, including variable domains from antibody heavy chain (FIG. 8) and light chain (FIG. 9) and amino acid sequence alignment of the germ-line V. These analyses revealed a structural position of amino acids that are involved in antigen binding and displaying hypervariable amino acid composition.

Example 3

Development of the scFv-IL-13Rα2 CAR Construct

The optimized CAR-T construct was placed into pCDH-MSCV-MCS-EF1-copGFP-T2A-Puro lentiviral vector (System Biosciences, MA). Sequencing data confirmed that the CAR construct is 1631 bp long, contains 60.35% GC, and consists of a CD8 signal peptide, the scFv-IL-13Rα2 transgene, a CD8 hinge, a CD28 transmembrane domain, a CD28 cytoplasmic domain, a 4-1BB, and a CD3ζ domain. FIG. 10 shows the corresponding amino acid sequence of the CAR-T construct with a 5' BamH1 restriction site as well as the location and sequence of each feature or domain. Restriction digest of CAR-T plasmid DNA revealed a 1640 bp band insert and a second band of cleaved vector (FIG. 11).

The recombinant lentiviral vector, along with three helper plasmids (pCD/NL-BHΔ1, pCEF-VSV-G and pCMV-Rev), were packaged to produce the final scFv-CAR-lentivirus in a HEK 293T producer cell line.

Example 4

Generation and Characteristics of IL-13Rα2-CAR-T Cells

The scFv-IL-13Rα2 CAR construct was transduced into Jurkat cells (an immortalized T cell line) to produce IL-13Rα2-CAR-Jurkat cells. Similarly, IL-13Rα2-CAR-T cells were generated from human peripheral blood mononuclear cells (PBMCs) that were activated, transduced with the vector, and expanded in culture.

Fluorescence-activated cell sorting (FACS) analysis confirmed expression of CD28 and CD3ζ in transduced CAR-Jurkat cells (FIGS. 12A and 12B). FACS analysis was performed after permeabilizing transduced Jurkat cells and immunostaining with anti-CD28 (CD28.2) mouse mAb PE conjugate (Cat#27826, Cell Signaling Technology, Danvers, MA) and CD3ζ monoclonal

antibody PE conjugate (Cat# 12-2479-82, ThermoFisher eBioscience, Carlsbad, CA). The data were expressed as normalized to mode values. Both CD28 cytoplasmic and CD3ζ signaling domains were detected.

Expression of the scFv-IL-13Rα2-CAR transgene was confirmed by an indirect immunofluorescence assay (IFA) (FIGS. 13A and 13B). The IFA assay was performed on transduced Jurkat and T cells. Cells expressing ≥ 2+ fluorescence intensity were counted at 200X magnification by viewing in Nikon® epifluorescence microscope. The percent of positive cells with >2+ immunostaining intensity was slightly higher in Jurkat cells than in CAR-T cells. A representative IFA image of transduced Jurkat cells is shown in FIG. 13B. Each value is mean ± SD of quadruple experiments determined in a blinded manner and expressed as % positive cells.

The viability of IL-13Rα2-CAR-Jurkat and CAR-T cells in cell culture was evaluated over the course of seven days by trypan blue exclusion technique. As shown in FIG. 14, both CAR-Jurkat and CAR-T cells continued to grow for 7 days and maintained cell viability. Similarly, both CAR-Jurkat and CAR-T maintained metabolic and proliferative activity for 7 days as determined by MTS assay (FIG. 15).

The cell phenotype and function of CAR-T cells was assessed by FACS analysis of cell surface and intracellular markers and compared with resting T cells. The CAR-T cells expressed CD25, CD44 and CD69 cell surface markers indicative of T cell activation and expression of all three markers was preserved during the expansion phase of CAR-T cell cultures as measured on day 7 (FIG. 16). Similarly, CAR-T cells expressed intracellular interferon-γ while resting T cells did not show any expression of IFN- γ (FIG. 16).

Example 5

CAR-T cells are cytotoxic to IL-13Rα2+ glioma tumor cells *in vitro*

Potency of the IL-13Rα2-CAR-T cells was investigated by measuring their cytotoxicity against two IL-13Rα2 positive malignant glioma tumor cell lines known to express high levels of IL-13Rα2 (U251 and U87MG). The IL-13Rα2-CAR-T cells were found to specifically kill IL-13Rα2 expressing tumor cells (IL-13Rα2+) in an effector cell number dependent manner (FIG. 17). The killing of tumor cells was highly specific to IL-13Rα2 expression on target tumor cells as gene silencing of *IL-13Rα2* on target tumor cells nearly eliminated all cytotoxic activity of CAR-T cells.

IL-13Rα2-CAR-T cell potency and specificity was also evaluated based on the release of IFN-γ upon exposure to IL-13Rα2 expressing tumor cells. As shown in FIG. 19, the CAR-T cells produced large and approximately equal amount of IFN-γ in the supernatant when co-cultured with three IL-13Rα2 positive glioma cell lines (U251, A172, and U87MG). In contrast, the CAR-T cells

only secreted a basal amount of IFN-γ when co-cultured with IL-13Rα2 negative or IL-13Rα2 KO tumor cell line, indicating a specific response to IL-13Rα2 positive tumor cells.

Finally, IL-13Rα2-CAR-T cell migration and invasion was investigated using a Boyden chamber assay. CAR-T cells were exposed to three different concentrations of recombinant IL-13Rα2Fc chimeric protein or conditioned medium obtained from IL-13Rα2 positive and IL-13Rα2 negative gene silenced human glioma cell lines. As shown in FIG. 20, CAR-T cell cultures from day 8 of expansion phase invaded and migrated to human IL-13Rα2Fc in a concentration dependent manner at 6 and 20 hr time points. Similarly, CAR-T cells invaded and migrated to conditioned medium from IL-13Rα2 positive glioma cells, but not to conditioned medium from IL-13Rα2 negative glioma cells.

Example 6

Testing *in vivo* Efficacy of IL-13Rα2-CAR-T Cells

This example describes methods that can be used to test efficacy of IL-13Rα2-CAR cells *in vivo*. While particular methods are provided, one of skill in the art will recognize that methods that deviate from these specific methods can also be used, including addition or omission of one or more steps.

A mouse solid cancer model can be used to evaluate efficacy of IL-13Rα2-CAR-T cells. In this model, immunodeficient mice are subcutaneously or orthotopically implanted with human tumor cells derived from brain cancer, pancreatic cancer, prostate cancer, or other tumors.

In one example, athymic nude mice are implanted with human brain tumor cells (U251 and U87MG glioma). In another example, NOD/Shi-*scid*/IL-2Rγ^{null} (NOG) mice are implanted with tumor cells. NOG mice lack mature T cells, B cells, and mononuclear cells due to NOD (nonobese diabetic) background and SCID mutation. In addition to the immunodeficiency, interleukin-2 receptor gamma deficiency in NOG mice allows engrafted human-T cells to subsequently differentiate.

In one example, humanized PBMC reconstituted NOG mice are used. NOG mice reconstituted with human PBMC develop mature human immune cells in peripheral blood and the spleen (Brady et al. (2014) *Clin Transl Immunology* 19: 3(12). These mice are implanted subcutaneously or orthotopically with tumor cells, such as U251 and U87 glioma tumor cells, or HS766, MiaPaca2, BXPC-3, Panc-1, HPAF-II or ASPC-1 pancreatic tumor cells. Mice are then administered IL-13-Rα2-CAR-T cells. A dose titration study is used to determine optimal cell dose for each tumor model. An additional group of NOG mice may be included that are injected with IL-13Rα2 gene knock out (KO) tumor cells to delineate the role of IL-13Rα2 as a tumor target.

The animals are followed for the regression of established tumors, metastasis and survival of control alongside CAR-T cell treated animals. Immune response of the animals is evaluated by measuring a number of parameters, such as $CD4^+$ and $CD8^+$ cells, regulatory cells (T regulatory cells (Tregs), myeloid derived suppressor cells (MDSC), antibody levels, and serum cytokine levels. The overall health of each animal is assessed at various time points by measuring body weight and through histological studies of vital organs at the time of sacrifice.

The efficacy of the IL-13Rα2-CAR-T cell therapy is also evaluated in combination with various tumor pre-treatments. The methods are similar, except tumor-bearing animals are pre-treated with a HDAC inhibitor (*e.g.*, Trichostatin A (TSA), sodium butyrate (NaB), SP600125, SR11302, or Suberoylanilide Hydroxamic Acid (SAHA)), adrenomedullin (AM), or IL-13-PE prior to CAR-T cell administration and compared to suitable control treatments (*e.g.*, animals not receiving CAR-T therapy, not receiving the combination therapy, and/or animals with IL-13Rα2 knocked-down tumor cells).

Example 7

Testing *in vivo* Efficacy of IL-13Rα2-CAR-T Cell in Combination with HDAC Inhibitor Treatment

This example describes methods that can be used to test efficacy of IL-13Rα2-CAR cells in combination with HDAC inhibitors *in vivo*. While particular methods are provided, one of skill in the art will recognize that methods that deviate from these specific methods can also be used, including addition or omission of one or more steps.

The effectiveness of IL-13Rα2-CAR-T cell therapy in combination with HDAC inhibitor can be evaluated in a cancer model using subcutaneous and orthotopic tumor implantation in immunodeficient mice. In this example, a pancreatic cancer cell line (such as Panc-1 or ASPC-1) is used to develop subcutaneous tumor implants in the flank of female athymic nude or NOG mice. About 4-6 days after tumor implantation, TSA or SAHA is administered. About 5 days after the start of TSA or SAHA administration, IL13Rα2-CAR-T cells or a control is also administered.

Mice body weight and tumor size is monitored and periodically measured. $CD4^+$, $CD8^+$, and T reg cell numbers, and cytokine profile in the blood are also periodically measured. Measurements continue until tumor size reaches 20 mm^2 in diameter. Animals are monitored during the course of the entire experiment for adverse toxic effects. An additional group of animals implanted with IL-13Rα2 knocked-down tumor cells and treated with the IL-13Rα2-CAR-T cell therapy may also be included as a control.

Example 8

Testing *in vivo* Efficacy of IL-13Rα2-CAR-T Cell Therapy in Combination with Adrenomedullin

This example describes methods that can be used to test efficacy of IL-13Rα2-CAR cells in combination with adrenomedullin treatment *in vivo*. While particular methods are provided, one of skill in the art will recognize that methods that deviate from these specific methods can also be used, including addition or omission of one or more steps.

In one example, PC-3 prostate cancer cells transfected with AM, or mock-transfected, or a non-transfected control are injected subcutaneously in immunodeficient mice, such as male athymic nude mice. When the tumor size reaches approximately 20 mm^2 (in approximately one week), mice are administered CAR-T cells and subsequently monitored for response to treatment.

In another example, mice are injected with PC-3 tumor cells, and when the tumor size reaches approximately 20 mm^2, AM peptide (or control) is periodically injected intratumorally. About forty-eight hours after the last AM treatment, the sensitivity of PC-3 tumors to CAR-T cells is examined. Animals injected with PC-3 cells may be included as additional controls.

The mice will be followed over several weeks to assess effectiveness of CAR-T therapy, for example, tumor shrinkage, general health of the animals, CD4+, CD8+, T regs, and blood cytokine levels.

Example 9

Testing *in vivo* Efficacy of IL-13Rα2-CAR-T Cell Therapy in Combination with IL-13-PE

This example describes methods that can be used to test efficacy of IL-13Rα2-CAR cells I combination with IL-13PE treatment *in vivo*. While particular methods are provided, one of skill in the art will recognize that methods that deviate from these specific methods can also be used, including addition or omission of one or more steps.

In this example, mice are treated with TGFβRI and PTEN in the oral epithelium and develop spontaneous squamous cell carcinoma of the head and neck. The mice are then treated with IL-13-PE. About one week later, the mice are treated with IL-13Rα2-CAR-T cells.

The mice are examined for antitumor activity and immunological changes in response to treatment. Some animals are sacrificed about three days after the last dose of CAR-T cell therapy to perform FACS analysis on splenocytes. MDSCs are analyzed by FACS analysis using the monocyte/macrophage markers CD11b+ and the granulocyte antigen Gr-1+.

In view of the many possible embodiments to which the principles of the disclosure may be applied, it should be recognized that illustrated embodiments are only examples and should not be taken as limiting the scope of the invention. Rather, the scope of the invention is defined by the following claims. We therefore claim as our invention all that comes within the scope and spirit of these claims.

We claim:

1. A single-chain fragment variable (scFv) that specifically binds interleukin-13 receptor α2 (IL-13Rα2), wherein the scFv comprises an amino acid sequence comprising variable heavy chain (VH) domain complementarity determining region 1 (CDR1), CDR2, and CDR3 amino acid sequences of amino acid positions 31-35, 50-66, and 99-107 of SEQ ID NO: 1, respectively, and variable light chain (VL) domain complementarity determining region 1 (CDR1), CDR2, and CDR3 amino acid sequences of amino acid positions 157-167, 183-189, and 222-229 of SEQ ID NO: 1, respectively.

2. The scFv of claim 1, wherein the amino acid sequence has at least 90% identity to SEQ ID NO: 1.

3. The scFv of claim 1 or claim 2, wherein the amino acid sequence comprises SEQ ID NO: 1.

4. The scFv of any one of claims 1 to 3, wherein the scFv binds to cells expressing IL-13Rα2.

5. A nucleic acid molecule encoding the scFv of any one of claims 1 to 4.

6. The nucleic acid molecule of claim 5, comprising a nucleic acid sequence having at least 90% identity to SEQ ID NO: 2.

7. The nucleic acid molecule of claim 6, comprising SEQ ID NO: 2.

8. A vector comprising the nucleic acid sequence of any one of claims 5 to 7.

9. A chimeric antigen receptor comprising:
(a) an antigen binding domain comprising the scFv of any one of claims 1 to 4;
(b) a hinge domain;
(c) a transmembrane domain; and
(d) an intracellular domain comprising one or more signaling domains.

10. The chimeric antigen receptor of claim 9, further comprising a signal peptide.

11. The chimeric antigen receptor of claim 10, wherein the signal peptide is a CD8α signal peptide.

12. The chimeric antigen receptor of any one of claims 9 to 11, wherein the one or more signaling domains comprise a CD28 domain, a 4-1BB domain, a CD3ζ domain, or any combination of two or more thereof.

13. The chimeric antigen receptor of claim 12, wherein the one or more signaling domains comprise a CD28 domain, a 4-1BB domain, and a CD3ζ domain.

14. The chimeric antigen receptor of any one of claims 9 to 13, wherein the hinge domain comprises a CD8α hinge domain.

15. The chimeric antigen receptor of any one of claims 9 to 14, wherein the transmembrane domain is a CD28 transmembrane domain.

16. The chimeric antigen receptor of any one of claims 9 to 15, comprising an amino acid sequence having at least 90% identity to SEQ ID NO: 3 or amino acids 22-538 of SEQ ID NO: 3.

17. The chimeric antigen receptor of any one of claims 9 to 16, comprising SEQ ID NO: 3 or amino acids 22-538 of SEQ ID NO: 3.

18. A nucleic acid molecule encoding the chimeric antigen receptor of any one of claims 9 to 17.

19. The nucleic acid molecule of claim 18, comprising a nucleic acid sequence having at least 90% identity to SEQ ID NO: 4.

20. The nucleic acid molecule of claim 19, comprising SEQ ID NO: 4.

21. A vector comprising the nucleic acid molecule of any one of claims 18 to 20.

22. The vector of claim 21, wherein the vector is a viral vector.

23. The vector of claim 22, wherein the viral vector is a lentiviral vector.

24. An immune cell expressing the chimeric antigen receptor of any one of claims 9 to 17.

25. An immune cell comprising the nucleic acid of any one of claims 18 to 20 or the vector of any one of claims 21 to 23.

26. The immune cell of claim 24 or claim 25, wherein the immune cell is obtained from a peripheral blood sample.

27. The immune cell of any one of claims 24 to 26, wherein the immune cell is a T cell, an NK cell, an NKT cell, or a macrophage.

28. A method of producing IL-13Rα2-CAR cells, comprising transforming or transfecting a population of immune cells with the vector of any one of claims 21 to 23.

29. The method of claim 28, wherein the population of immune cells is a population of T cells, NK cells, NKT cells, or macrophages.

30. The method of claim 29, wherein the T cells, NK cells, NKT cells or macrophages are obtained from a peripheral blood sample of a subject.

31. A method of treating a subject with cancer, comprising administering an effective amount of the immune cell of any one of claims 24 to 27 to the subject.

32. The method of claim 31, wherein the immune cell is autologous to the subject with cancer.

33. The method of claim 31 or claim 32, wherein the subject has a cancer that expresses IL-13Rα2.

34. The method of any one of claims 31 to 33, wherein the subject has a solid tumor.

35. The method of any one of claims 31 to 34, wherein the subject has pancreatic cancer, glioblastoma, head and neck squamous cell carcinoma, ovarian cancer, renal cell carcinoma, uterine

cancer, prostate cancer, breast cancer, melanoma, non-small cell lung cancer (NSCLC), Kaposi sarcoma, or adrenal carcinoma.

36. The method of any one of claims 31 to 35, further comprising treating the subject with one or more of surgery, radiation, chemotherapy, or an additional immunotherapy.

37. The method of any one of claims 31 to 35, further comprising administering to the subject a histone deacetylase (HDAC) inhibitor, a cell cycle or checkpoint inhibitor, adrenomedullin, an IL-13-PE immunotoxin, or any combination of two or more thereof.

FIG. 1

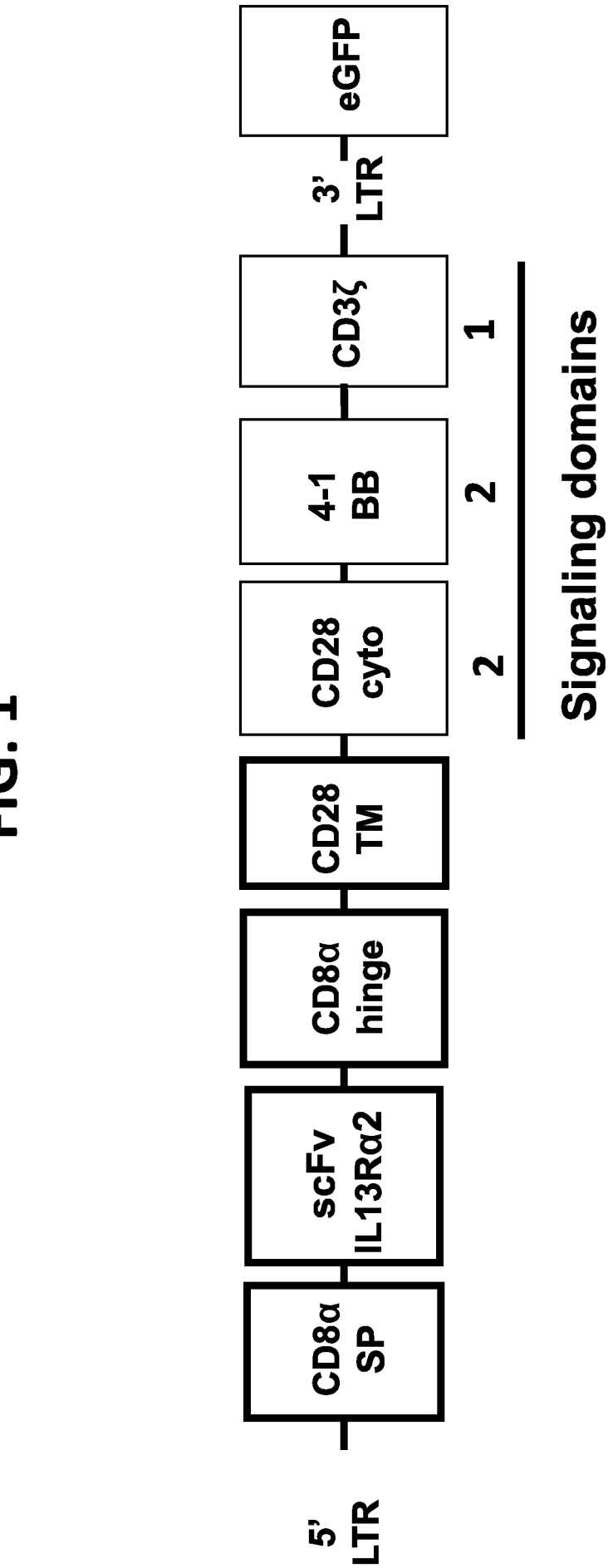

5' LTR
CD8α SP
scFv IL13Rα2
CD8α hinge
CD28 TM
CD28 cyto
4-1 BB
CD3ζ
3' LTR
eGFP
2
2
1
Signaling domains

FIG. 2

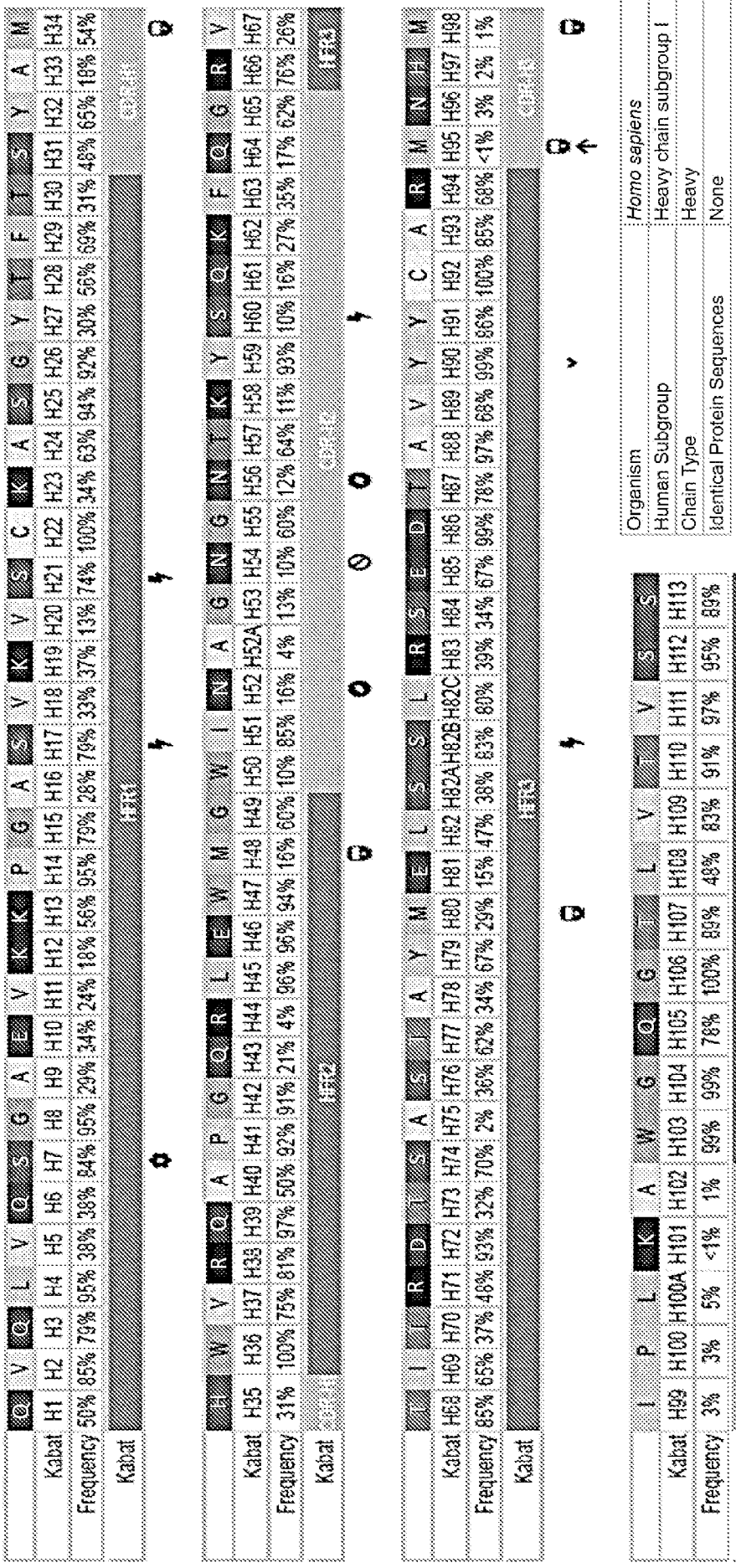

Organism
Homo sapiens
Human Subgroup
Heavy chain subgroup I
Chain Type
Heavy
Identical Protein Sequences
None

FIG. 3A

Regions Definition: Kabat

SEQ ID NO (residues)	Region	Sequence Fragment	Residues	Length
SEQ ID NO: 3 (1-21)	Leader	MALPVTALLLPLALLLHAARP	1 - 21	21
SEQ ID NO: 3 (22-51)	HFR1	QVQLVQSGAEVKKPGASVKVSCKASGYTFT	22 - 51	30
SEQ ID NO: 3 (52-56)	CDR-H1	SYAMH	52 - 56	5
SEQ ID NO: 3 (57-70)	HFR2	WVRQAPGQRLEWMG	57 - 70	14
SEQ ID NO: 3 (71-87)	CDR-H2	WINAGNGNTKYSQKFQG	71 - 87	17
SEQ ID NO: 3 (88-119)	HFR3	RVTITRDTSASTAYMELSSLRSEDTAVYYCAR	88 - 119	32
SEQ ID NO: 3 (120-128)	CDR-H3	MNHMIPLKA	120 - 128	9
SEQ ID NO: 3 (129-139)	HFR4	WGQGTLVTVSS	129 - 139	11
				139

FIG. 3B

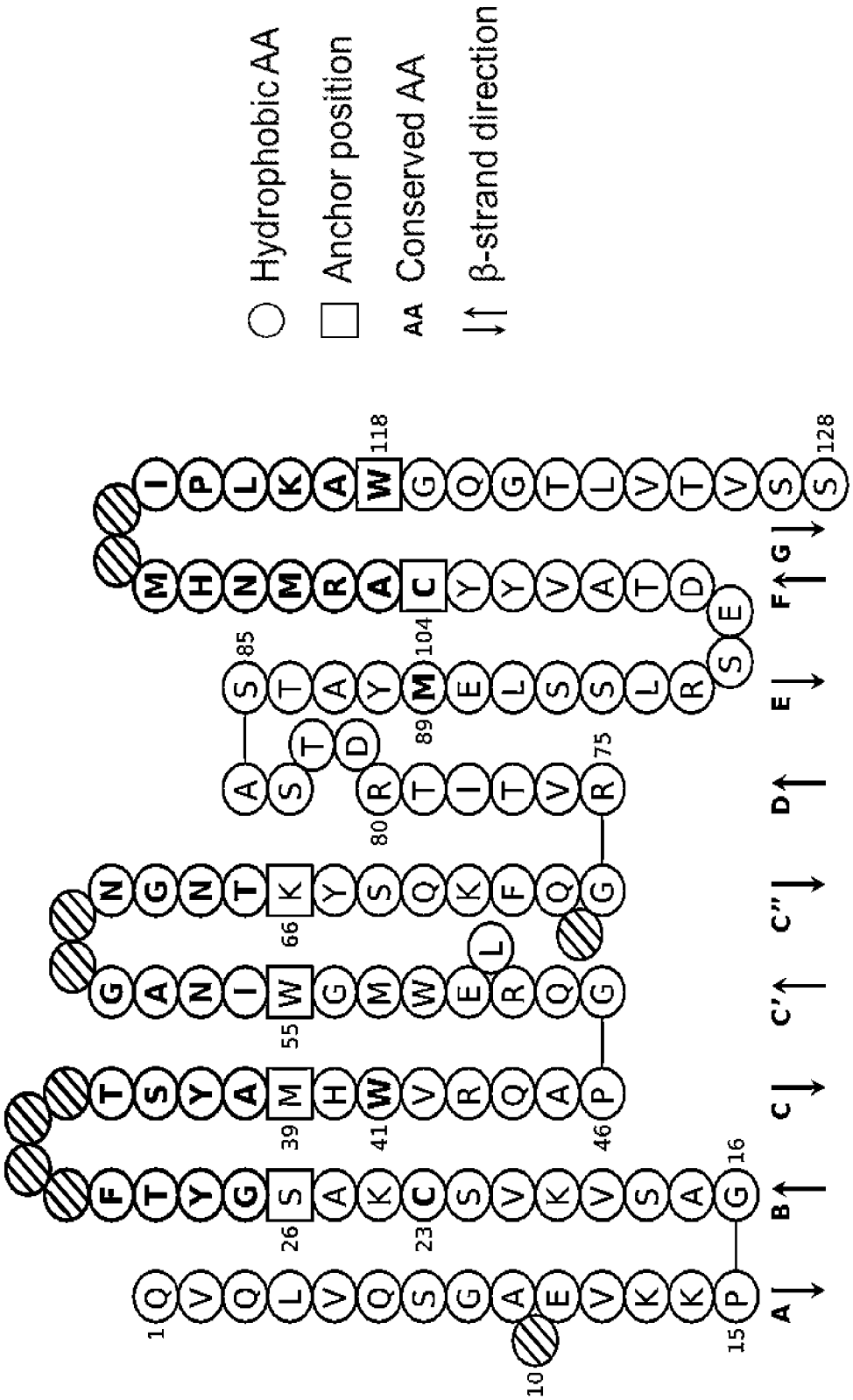

Hydrophobic AA
Anchor position
AA Conserved AA
β-strand direction
A
B
C
C'
C''
D
E
F
G

FIG. 4

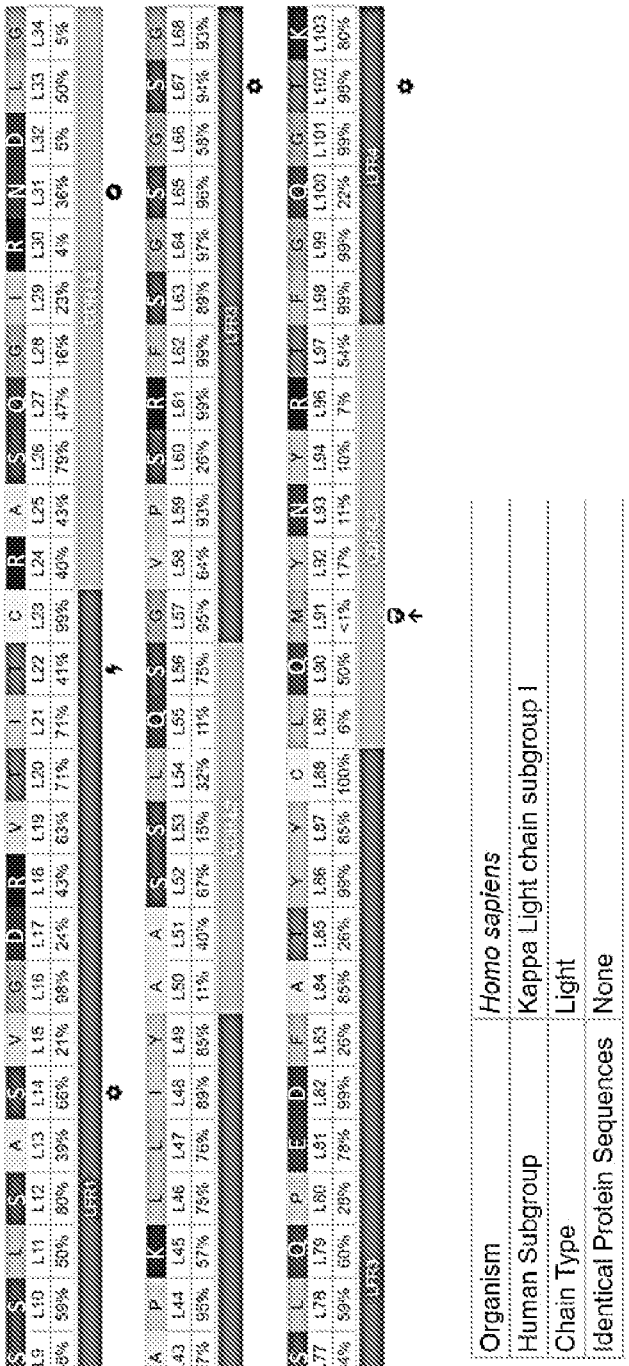

Organism
Homo sapiens
Human Subgroup
Kappa Light chain subgroup 1
Chain Type
Light
Identical Protein Sequences
None

FIG. 5A

Regions Definition: Kabat

SEQ ID NO (residues)	Region	Sequence Fragment	Residues	Length
SEQ ID NO: 3 (153-154)	Leader	AL	1 - 2	2
SEQ ID NO: 3 (155-177)	LFR1	AIQMTQSPSSLSASVGDRVTITC	3 - 25	23
SEQ ID NO: 3 (178-188)	CDR-L1	RASQGIRNDLG	26 - 36	11
SEQ ID NO: 3 (189-203)	LFR2	WYQQKPGKAPKLLIY	37 - 51	15
SEQ ID NO: 3 (204-210)	CDR-L2	AASSLQS	52 - 58	7
SEQ ID NO: 3 (211-242)	LFR3	GVPSRFSGSGSGTDFTLTISSLQPEDFATYYC	59 - 90	32
SEQ ID NO: 3 (243-250)	CDR-L3	LQMYNYRT	91 - 98	8
SEQ ID NO: 3 (251-263)	LFR4	FGQGTKLEIKRAA	99 - 111	13
SEQ ID NO: 3 (264-538)	Tail	KPTTTPAPRPPTPAPTIASQPLSLRPEACRPAAGGAVHTRGLDFACDFWVLVVVGGVLAC YSLLVTVAFIIFWVRSKRSRGGHSDYMNMTPRRPGPTRKHYQPYAPPRDFAAYRSVDKRG RKKLLYIFKQPFMRPVQTTQEEDGCSCRFPEEEEGGCELGGGRVKFSRSADAPAYQQGQN QLYNELNLGRREEYDVLDKRRGRDPEMGGKPQRRKNPQEGLYNELQKDKMAEAYSEIGMK GERRRGKGHDGLYQGLSTATKDTYDALHMQALPPR	111 - 386	275
				386

FIG. 5B

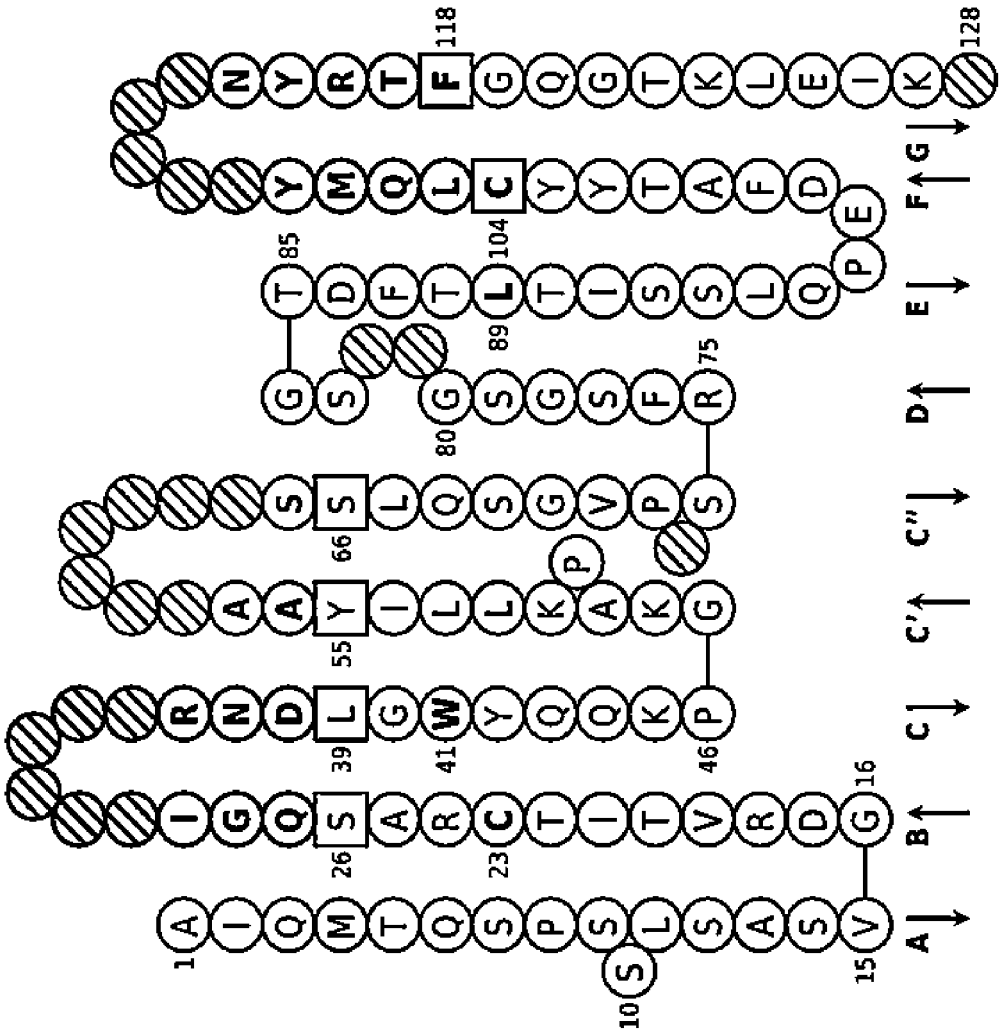
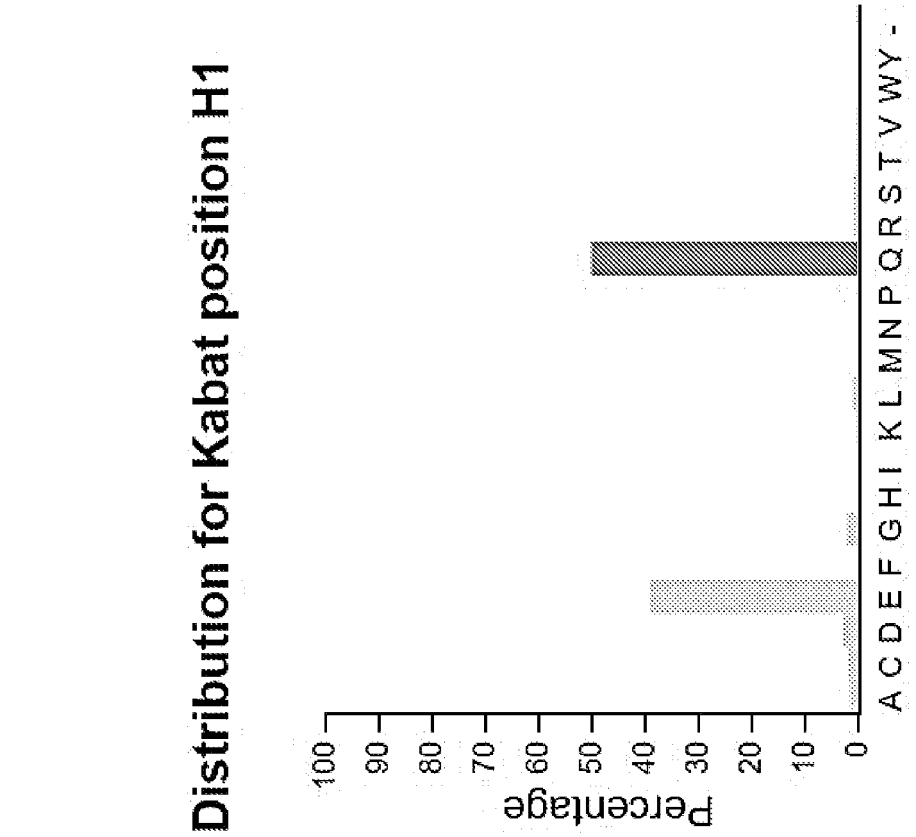

Hydrophobic AA
Anchor position
AA Conserved AA
β-strand direction
A
B
C
C'
C''
D
E
F
G

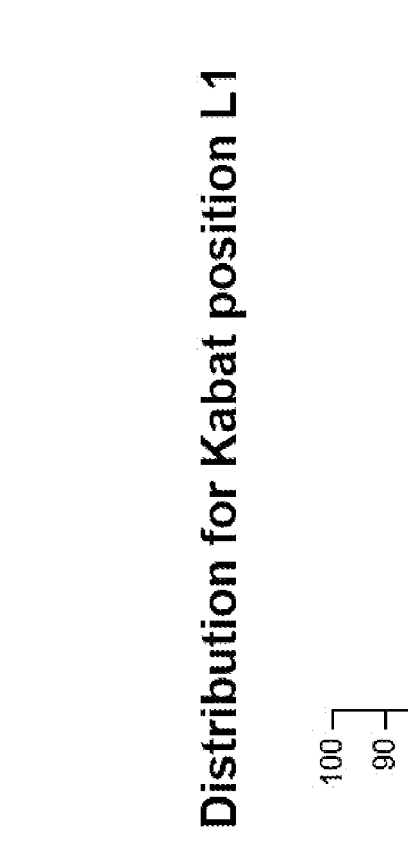
FIG. 6



AA	Sequences	Frequency
A	655	1%
C	877	2%
D	1416	3%
E	20608	39%
F	4	<1%
G	977	2%
H	99	<1%
I	45	<1%
K	136	<1%
L	433	<1%
M	21	<1%
N	46	<1%
P	121	<1%
Q	25440	48%
R	365	<1%
S	359	<1%
T	146	<1%
V	149	<1%
W	4	<1%
Y	14	<1%
-	124	<1%
Total	53047	100%

CSV

FIG. 7



AA	Sequences	Frequency
A	1499	5%
C	6	<1%
D	10141	36%
E	4689	16%
F	31	<1%
G	203	<1%
H	136	<1%
I	39	<1%
K	72	<1%
L	55	<1%
M	41	<1%
N	408	1%
P	47	<1%
Q	8768	31%
R	43	<1%
S	2032	7%
T	50	<1%
V	41	<1%
W	4	<1%
Y	141	<1%
-	20	<1%
Total	28466	100%

CSV

FIG. 8

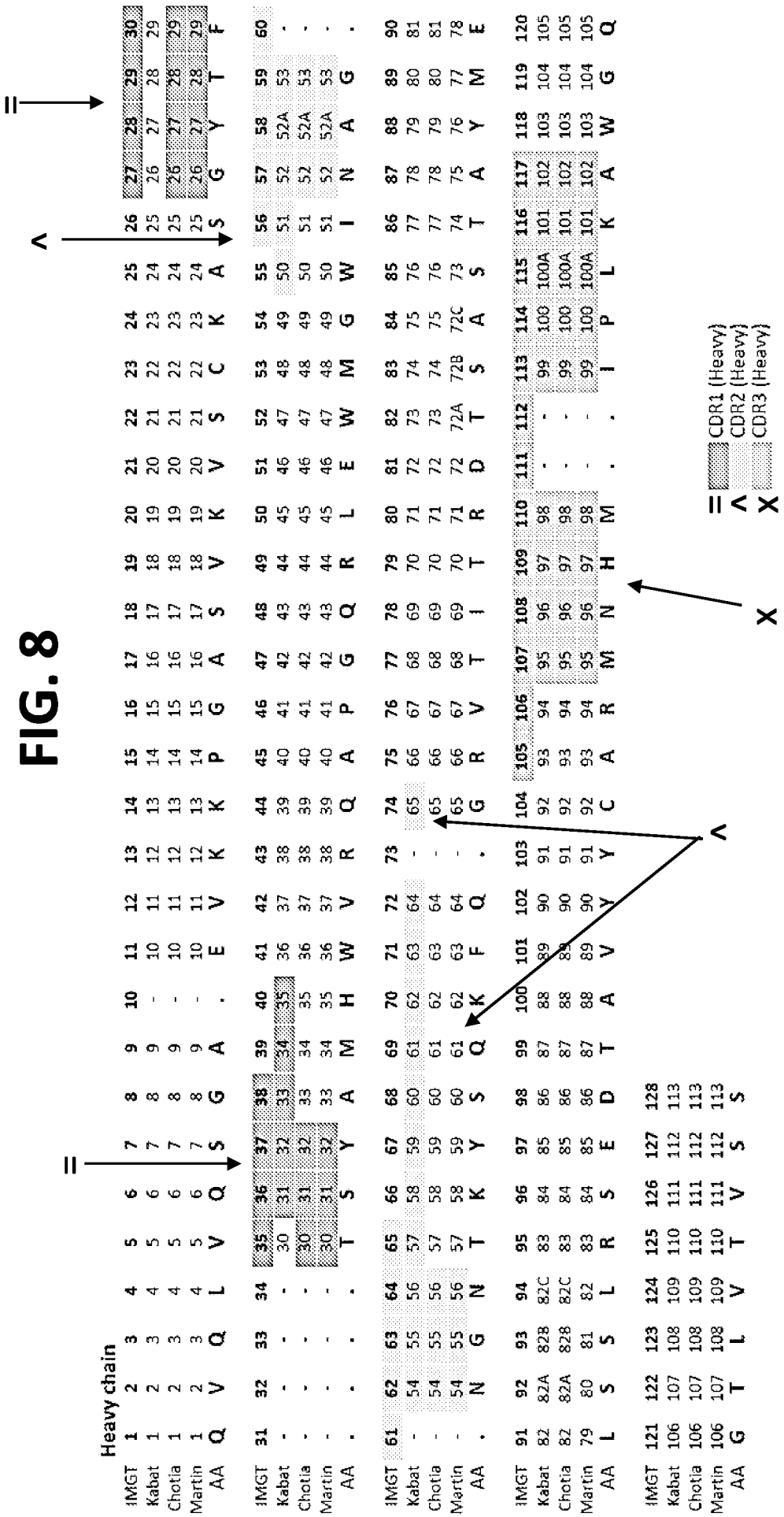


FIG. 9

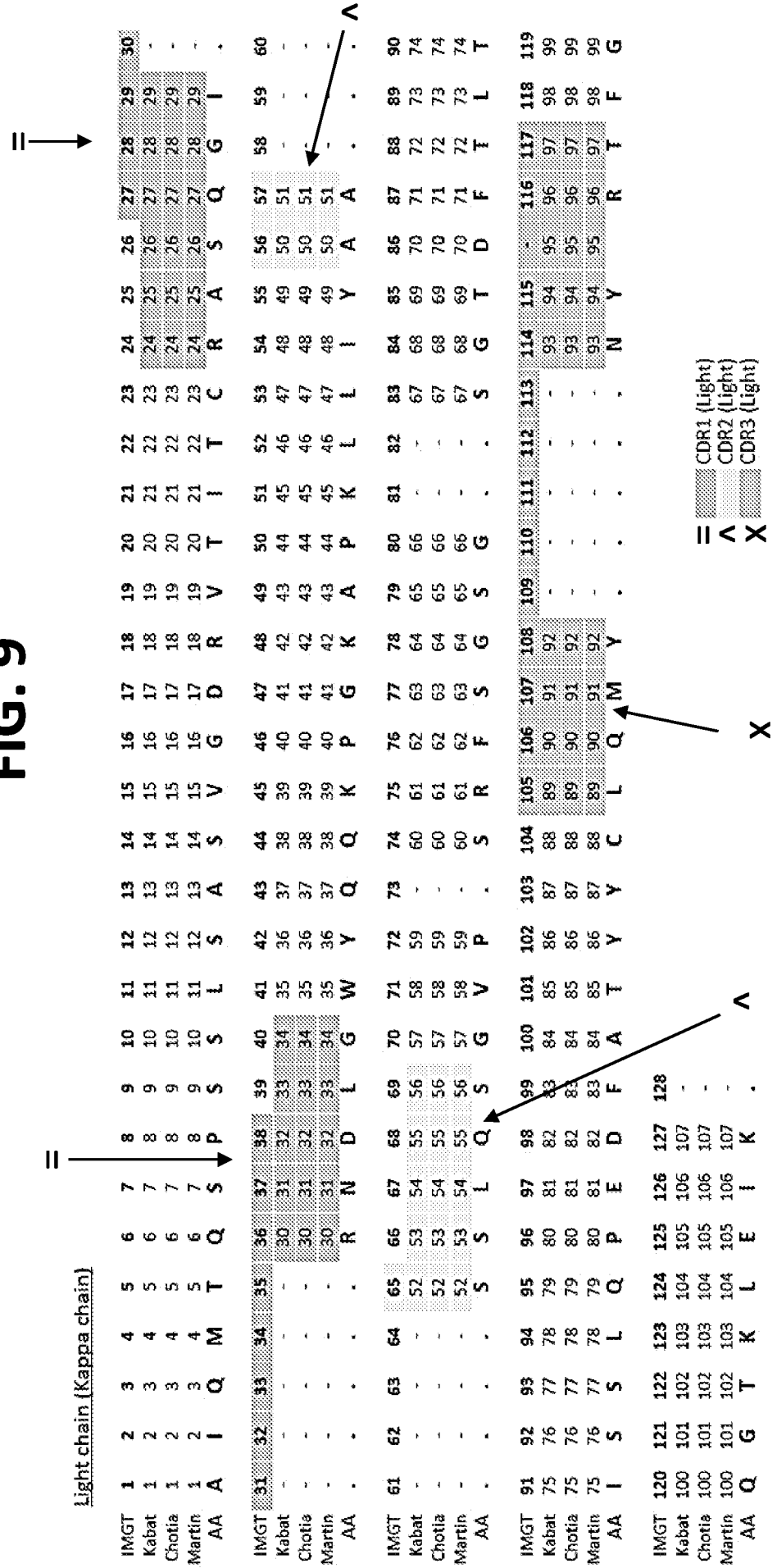
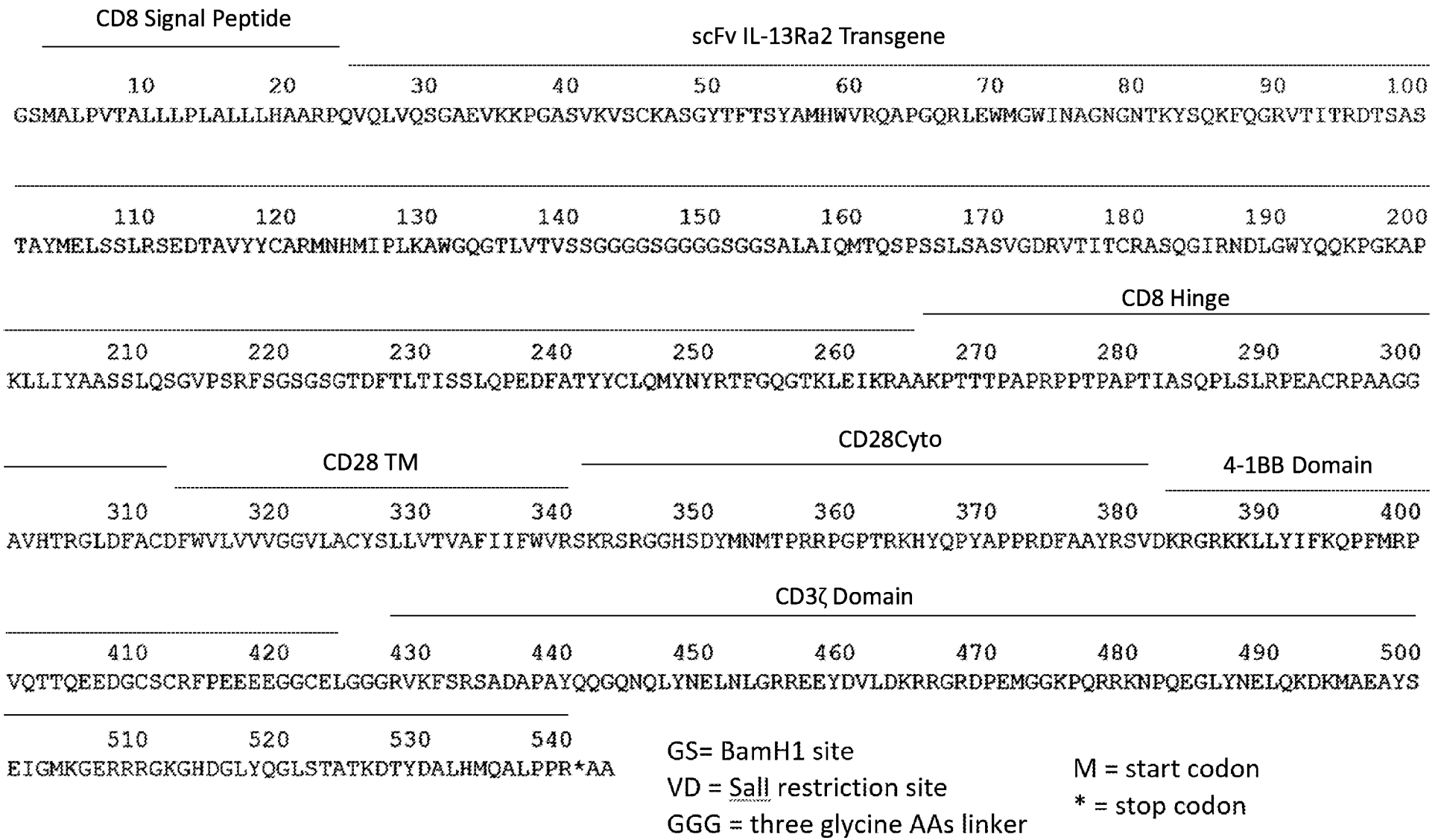


FIG. 10



GS= BamH1 site
VD = SalI restriction site
GGG = three glycine AAs linker

M = start codon
* = stop codon

FIG. 11

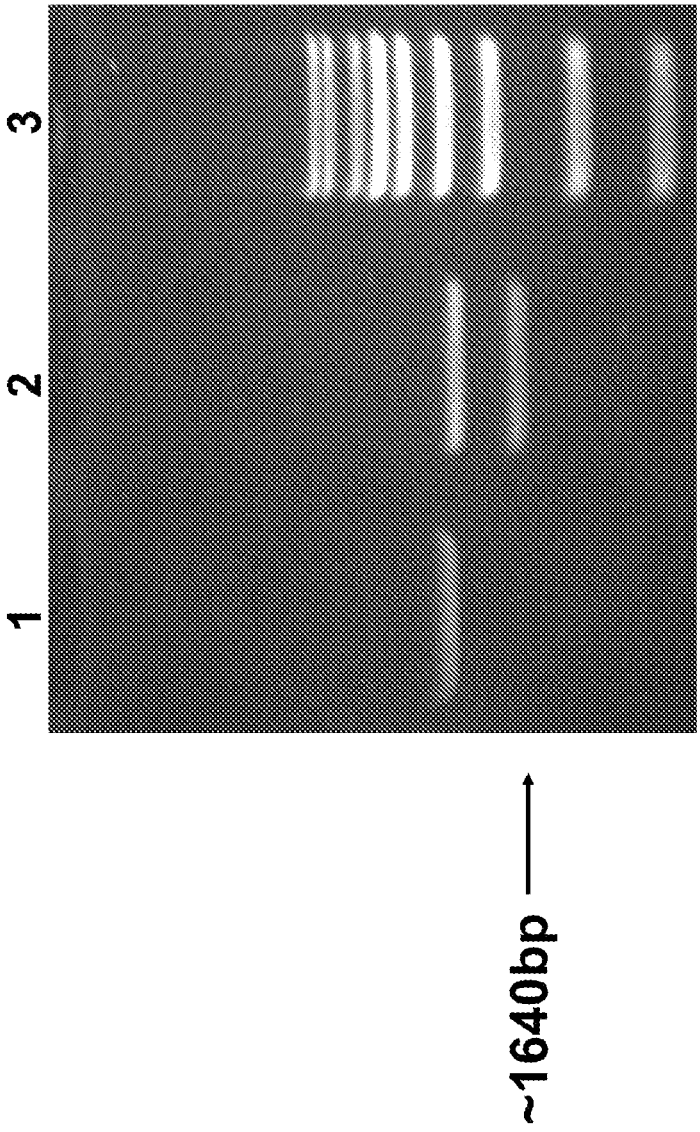


FIG. 12A

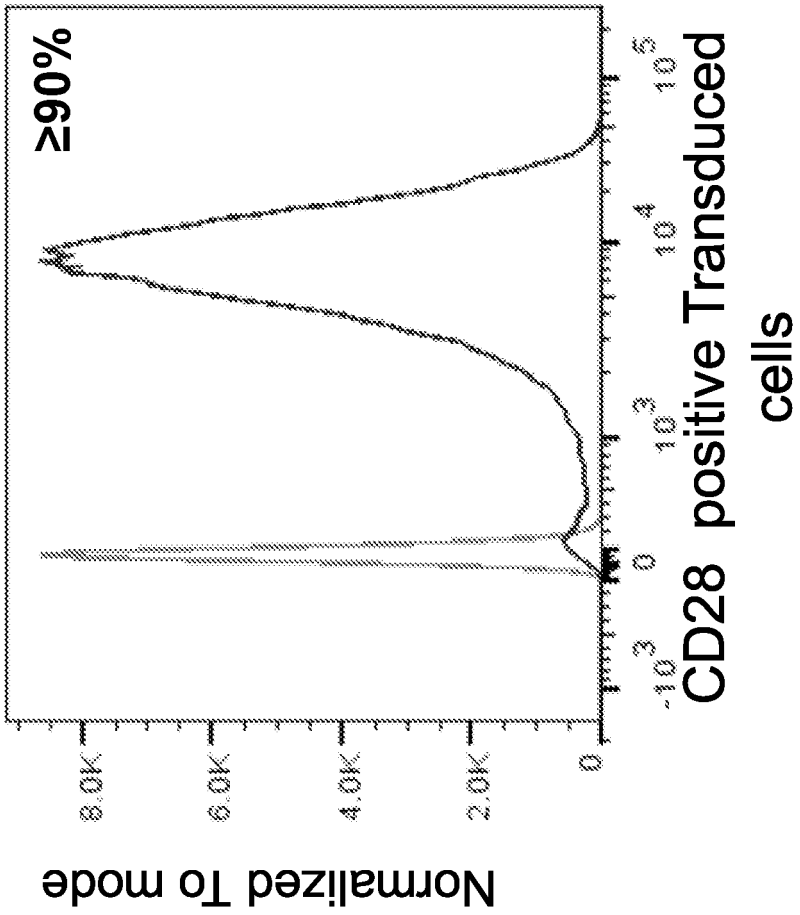


FIG. 12B

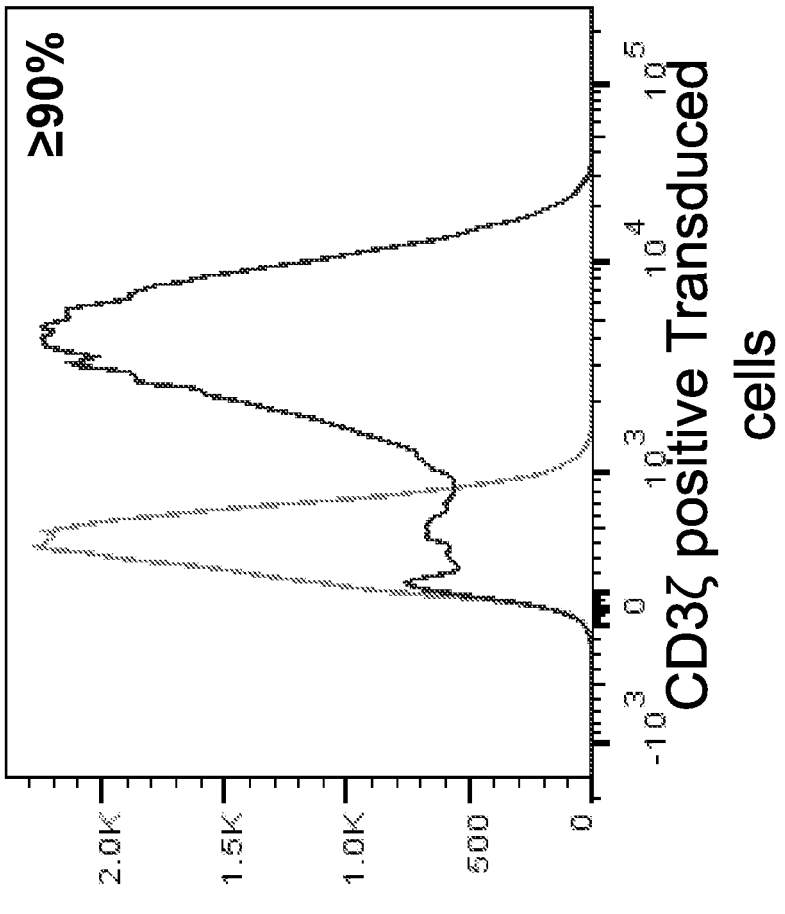


FIG. 13A

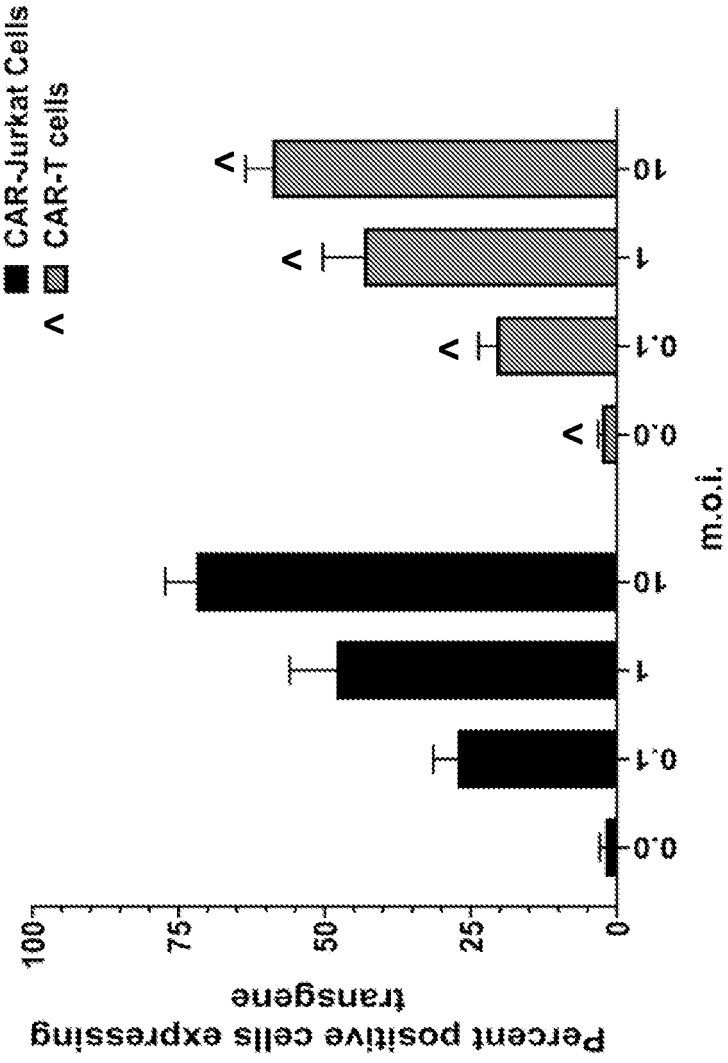


FIG. 13B

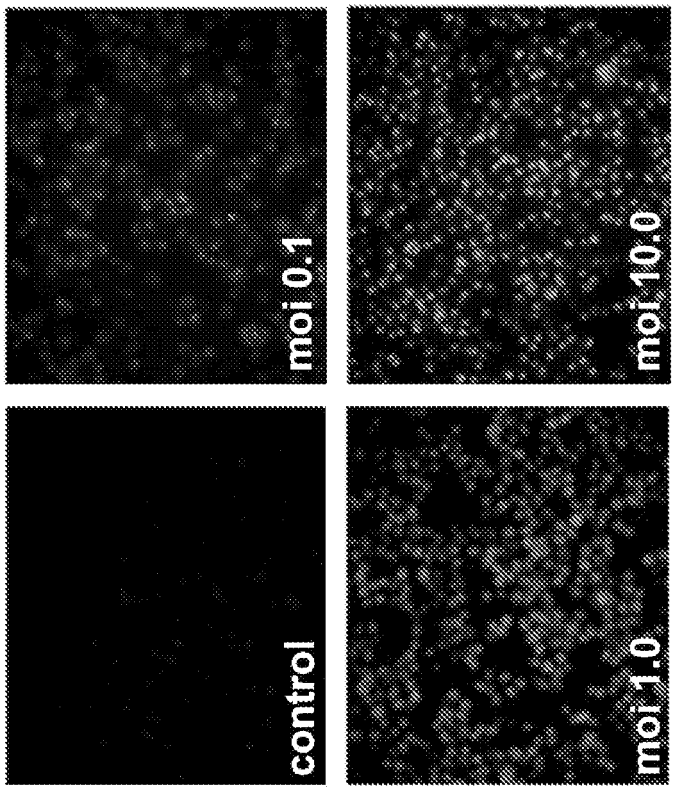


FIG. 14

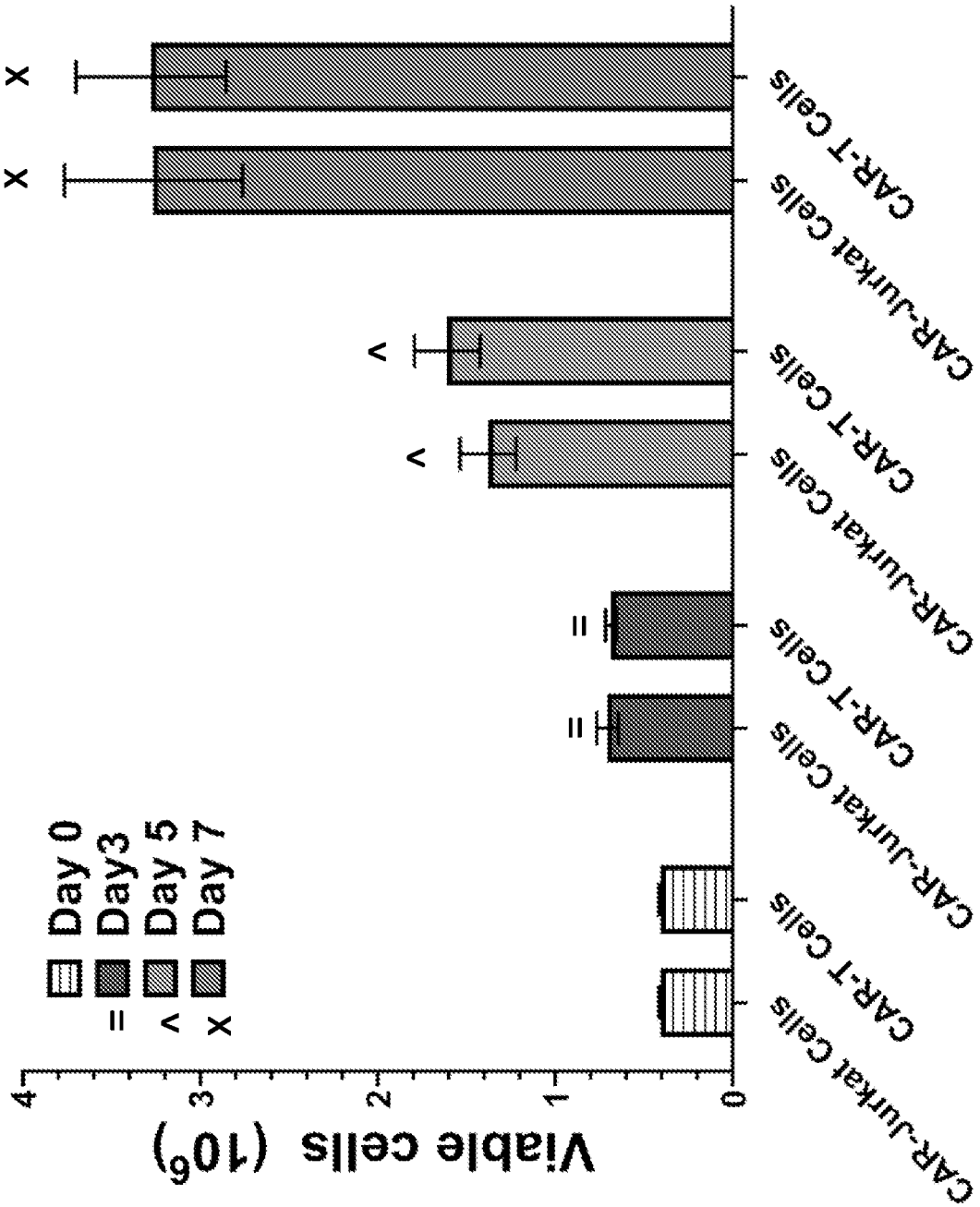

Viable cells (10^6)
0
1
2
3
4
Day 0
Day3
Day 5
Day 7
=
<
>
x
CAR-Jurkat cells
CAR-T cells
CAR-Jurkat cells
CAR-T cells
CAR-Jurkat cells
CAR-T cells
CAR-Jurkat cells
CAR-T cells

FIG. 15

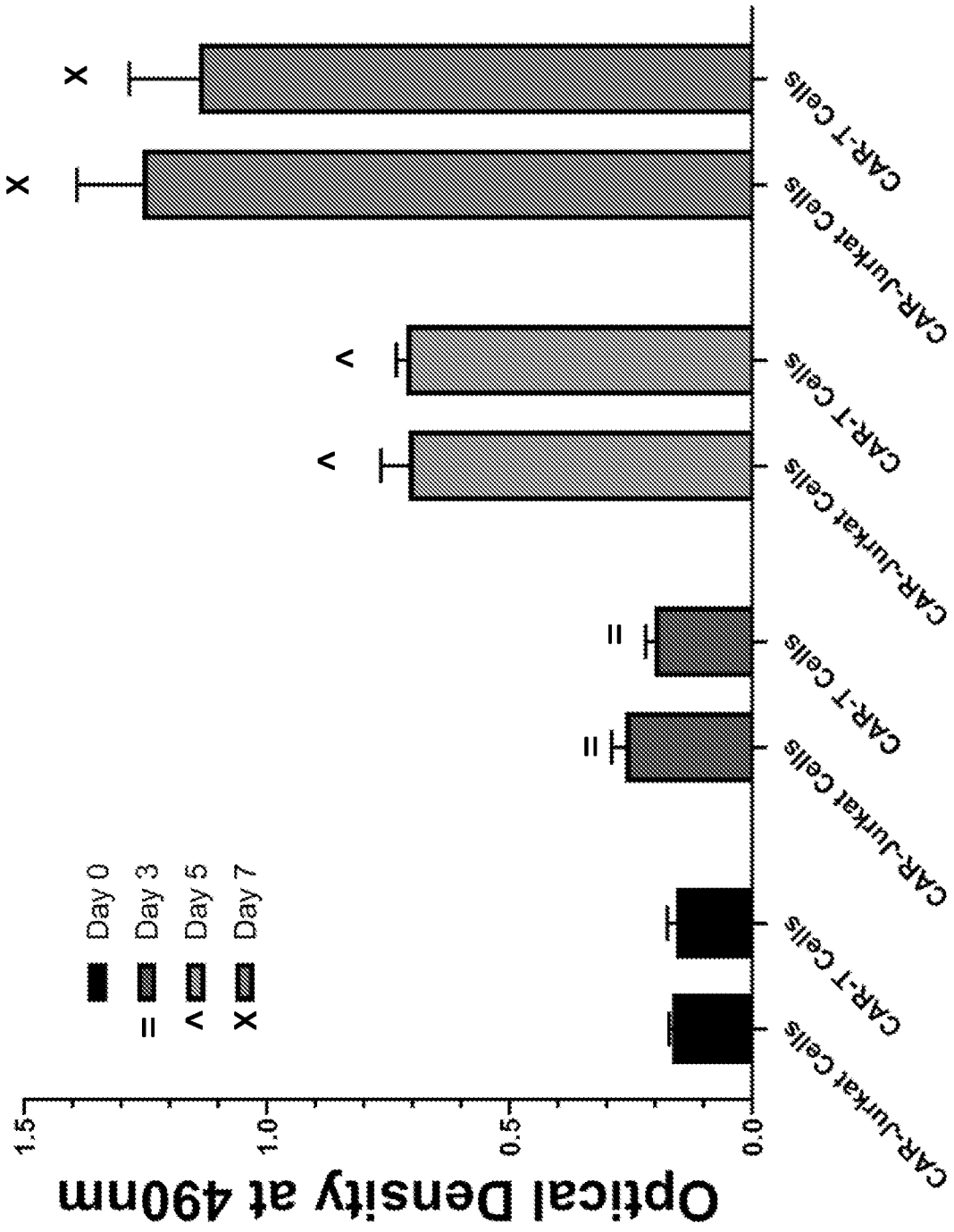

Optical Density at 490nm
0.0
0.5
1.0
1.5
Day 0
Day 3
Day 5
Day 7
=
<
X
CAR-Jurkat cells
CAR-T cells
CAR-Jurkat cells
CAR-T cells
CAR-Jurkat cells
CAR-T cells
CAR-Jurkat cells
CAR-T cells

FIG. 16

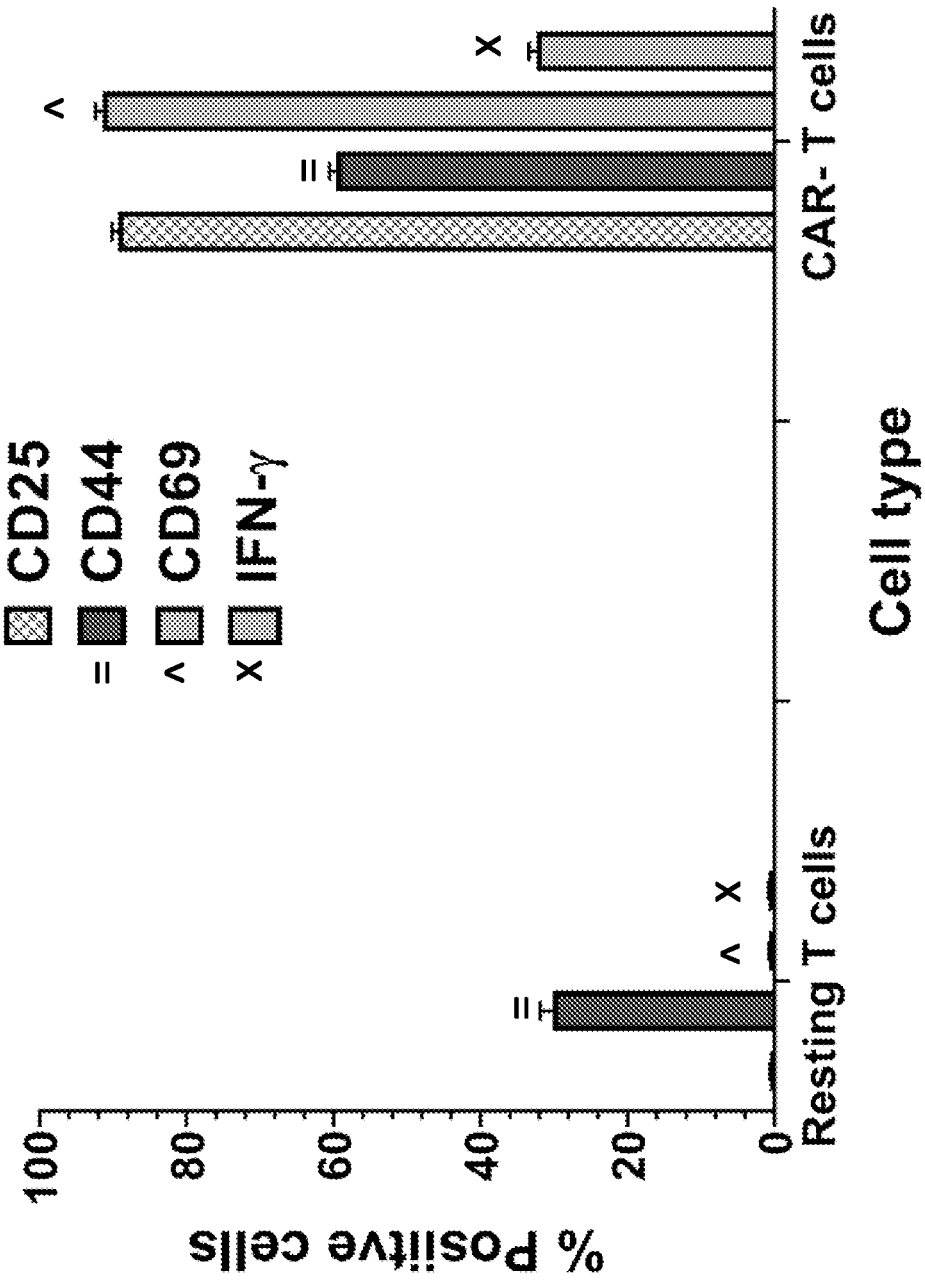

CD25
CD44
CD69
IFN-γ
% Positive cells
100
80
60
40
20
0
Resting T cells
CAR- T cells
Cell type

FIG. 17

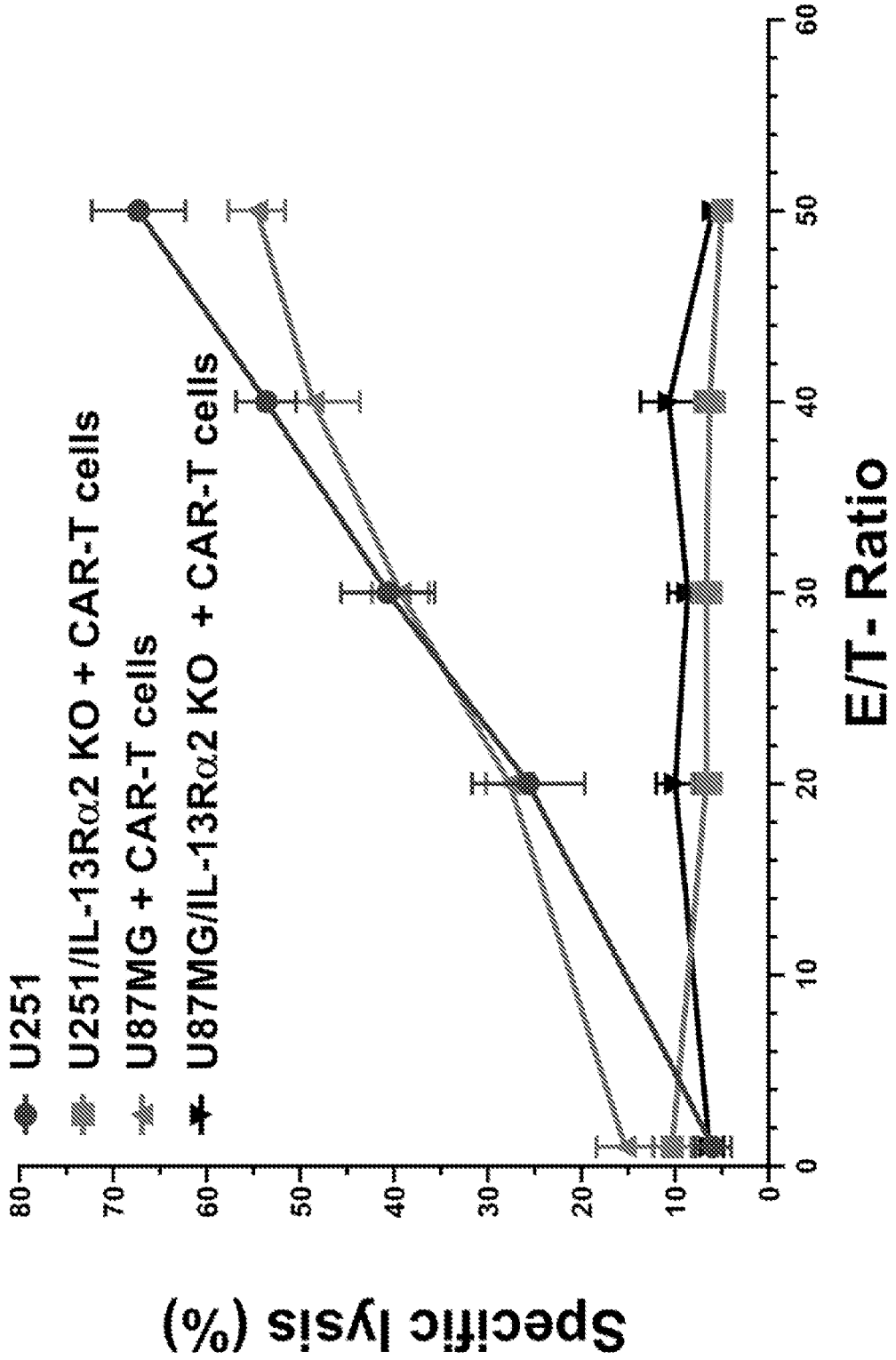

U251
U251/IL-13Rα2 KO + CAR-T cells
U87MG + CAR-T cells
U87MG/IL-13Rα2 KO + CAR-T cells
Specific lysis (%)
E/T- Ratio
0
10
20
30
40
50
60
70
80

FIG. 18

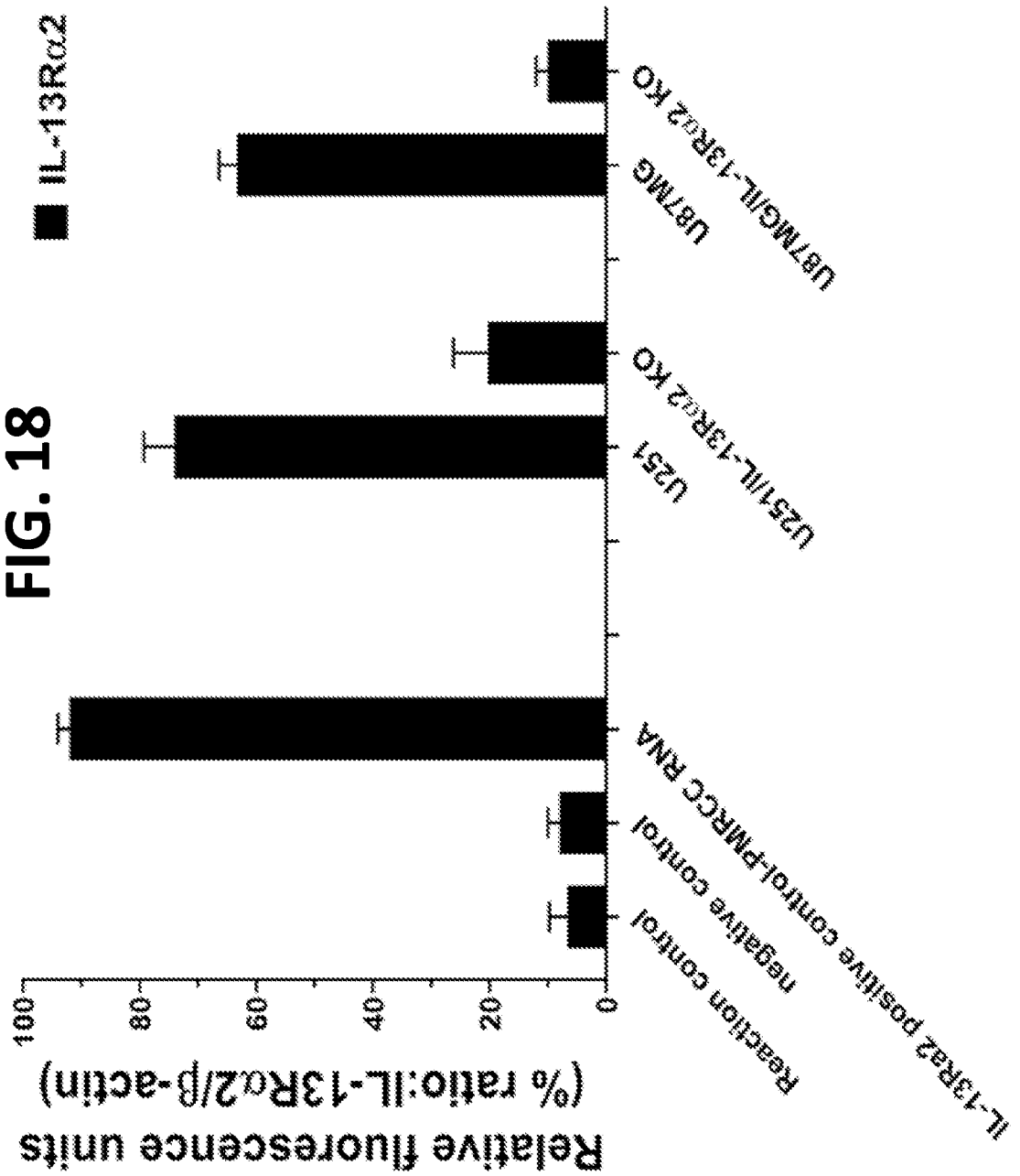

IL-13Rα2
Relative fluorescence units
(% ratio:IL-13Rα2/β-actin)
100
80
60
40
20
0
Reaction control
negative control
IL-13Rα2 positive control-PMRCC RNA
U251
U251/IL-13Rα2 KO
U87MG
U87MG/IL-13Rα2 KO
Cell type

FIG. 19

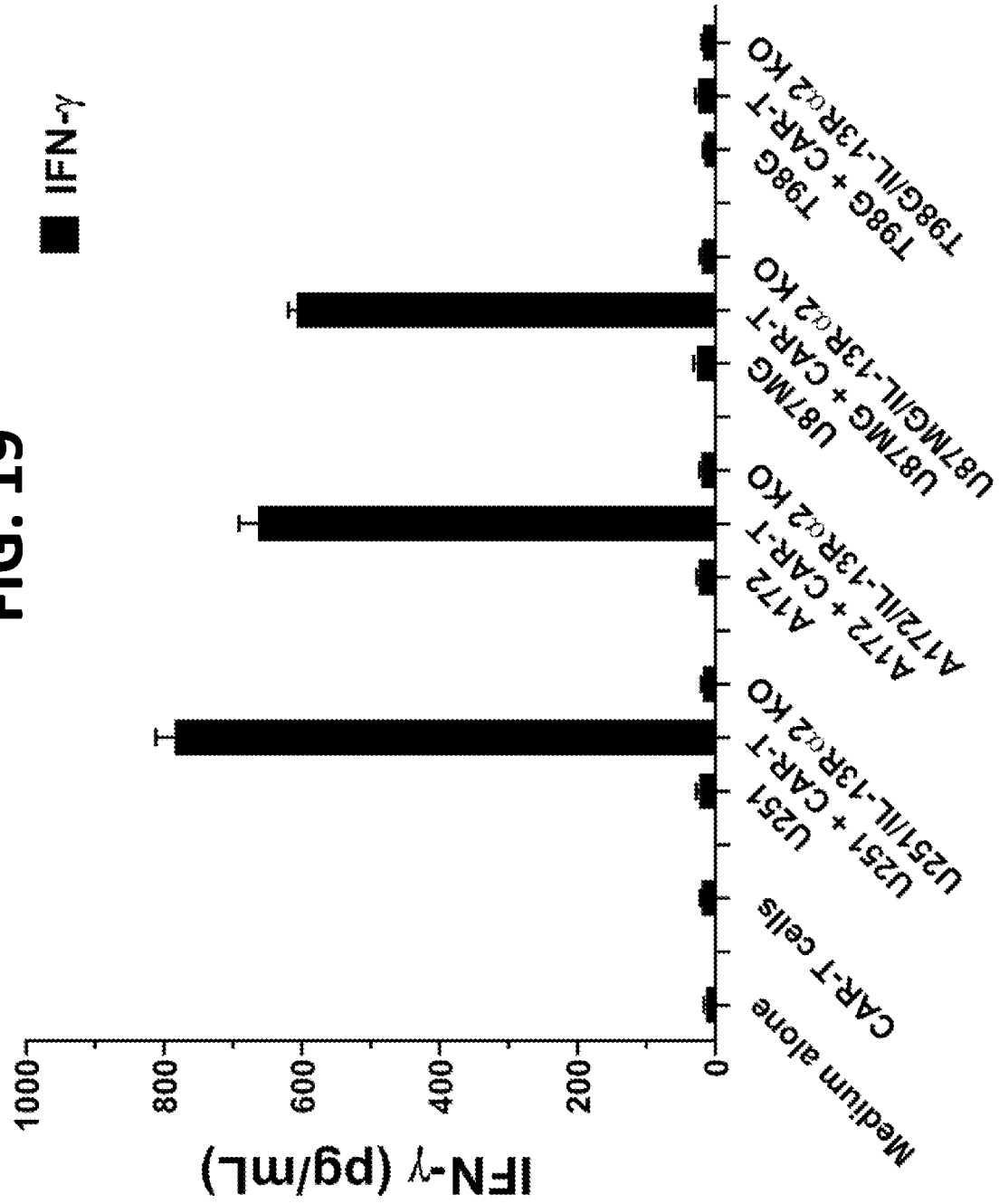
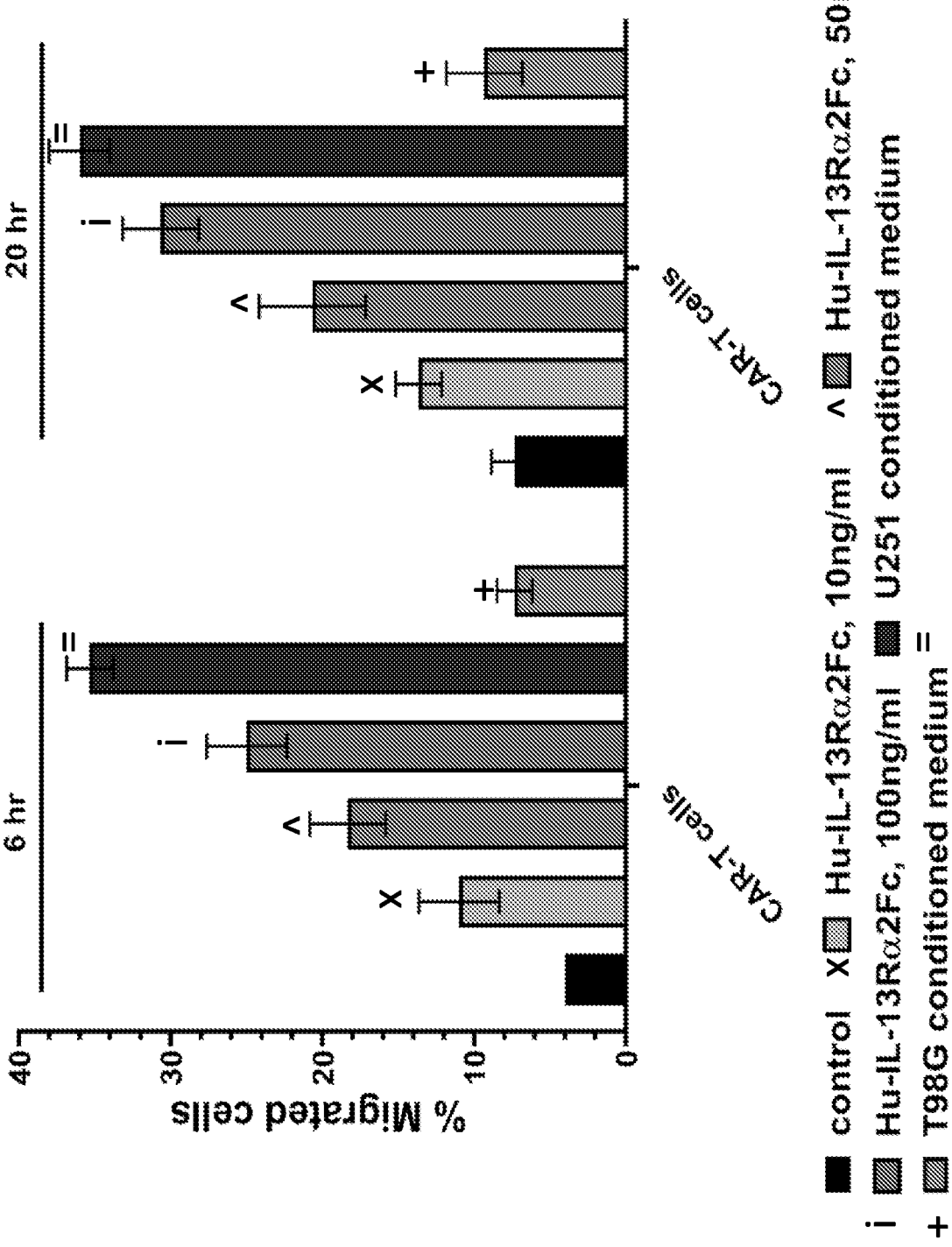

IFN-γ
IFN-γ (pg/mL)
1000
800
600
400
200
0
Medium alone
CAR-T cells
U251
U251 + CAR-T
U251/IL-13Rα2 KO
A172
A172 + CAR-T
A172/IL-13Rα2 KO
U87MG
U87MG + CAR-T
U87MG/IL-13Rα2 KO
T98G
T98G + CAR-T
T98G/IL-13Rα2 KO

FIG. 20


% Migrated cells
40
30
20
10
0
6 hr
20 hr
CAR-T cells
CAR-T cells
control
X
Hu-IL-13Rα2Fc, 10ng/ml
^
Hu-IL-13Rα2Fc, 50ng/ml
i
Hu-IL-13Rα2Fc, 100ng/ml
U251 conditioned medium
+
T98G conditioned medium
=

INTERNATIONAL SEARCH REPORT

International application No
PCT/US2022/023112

A. CLASSIFICATION OF SUBJECT MATTER
INV. C07K16/28 A61K39/00 C07K14/725
ADD.

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)
C07K A61K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)
EPO-Internal

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	WO 2016/123142 A1 (UNIV CHICAGO [US]) 4 August 2016 (2016-08-04)	1-36
Y	paragraphs [0276], [0275], [0244] example 11	37
X	WO 2020/210665 A1 (ST JUDE CHILDRENS RES HOSPITAL INC [US]) 15 October 2020 (2020-10-15)	1-36
Y	example 7 claim 47	37
X	WO 2019/178078 A1 (SEATTLE CHILDRENS HOSPITAL DBA SEATTLE CHILDRENS RES INST [US]) 19 September 2019 (2019-09-19)	1-36
Y	examples 2-3 figure 2	37

-/--

[X] Further documents are listed in the continuation of Box C.

[X] See patent family annex.

* Special categories of cited documents :

"A" document defining the general state of the art which is not considered to be of particular relevance

"E" earlier application or patent but published on or after the international filing date

"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)

"O" document referring to an oral disclosure, use, exhibition or other means

"P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance;; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

"Y" document of particular relevance;; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art

"&" document member of the same patent family

Date of the actual completion of the international search	Date of mailing of the international search report
20 June 2022	28/06/2022

Name and mailing address of the ISA/	Authorized officer
European Patent Office, P.B. 5818 Patentlaan 2 NL - 2280 HV Rijswijk Tel. (+31-70) 340-2040, Fax: (+31-70) 340-3016	Bumb, Peter

INTERNATIONAL SEARCH REPORT

International application No
PCT/US2022/023112

C(Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	US 2019/359723 A1 (WANG PENG [CN] ET AL) 28 November 2019 (2019-11-28)	1-36
Y	example 12 figure 13	37
Y	DE-GANG SONG ET AL: "Chimeric NKG2D CAR-Expressing T Cell-Mediated Attack of Human Ovarian Cancer Is Enhanced by Histone Deacetylase Inhibition", HUMAN GENE THERAPY, vol. 24, no. 3, 1 March 2013 (2013-03-01), pages 295-305, XP055370704, GB ISSN: 1043-0342, DOI: 10.1089/hum.2012.143 the whole document	37

INTERNATIONAL SEARCH REPORT

International application No.

PCT/US2022/023112

Box No. I Nucleotide and/or amino acid sequence(s) (Continuation of item 1.c of the first sheet)

1. With regard to any nucleotide and/or amino acid sequence disclosed in the international application, the international search was carried out on the basis of a sequence listing:

   a. [X] forming part of the international application as filed:

      [X] in the form of an Annex C/ST.25 text file.

      [ ] on paper or in the form of an image file.

   b. [ ] furnished together with the international application under PCT Rule 13ter.1(a) for the purposes of international search only in the form of an Annex C/ST.25 text file.

   c. [ ] furnished subsequent to the international filing date for the purposes of international search only:

      [ ] in the form of an Annex C/ST.25 text file (Rule 13ter.1(a)).

      [ ] on paper or in the form of an image file (Rule 13ter.1(b) and Administrative Instructions, Section 713).

2. [ ] In addition, in the case that more than one version or copy of a sequence listing has been filed or furnished, the required statements that the information in the subsequent or additional copies is identical to that forming part of the application as filed or does not go beyond the application as filed, as appropriate, were furnished.

3. Additional comments:

INTERNATIONAL SEARCH REPORT

Information on patent family members

International application No
PCT/US2022/023112

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
WO 2016123142 A1	04-08-2016	CN 107683289 A	09-02-2018
		EP 3250609 A1	06-12-2017
		HK 1245287 A1	24-08-2018
		JP 2018506301 A	08-03-2018
		US 2018134796 A1	17-05-2018
		US 2019300616 A1	03-10-2019
		US 2021221895 A1	22-07-2021
		WO 2016123142 A1	04-08-2016
WO 2020210665 A1	15-10-2020	NONE	
WO 2019178078 A1	19-09-2019	AU 2019234573 A1	08-10-2020
		BR 112020018670 A2	05-01-2021
		CA 3093810 A1	19-09-2019
		CN 112236151 A	15-01-2021
		EA 202091982 A1	10-06-2021
		EP 3765041 A1	20-01-2021
		JP 2021518108 A	02-08-2021
		KR 20200131279 A	23-11-2020
		SG 11202008795S A	29-10-2020
		US 2021000875 A1	07-01-2021
		WO 2019178078 A1	19-09-2019
US 2019359723 A1	28-11-2019	AU 2018221110 A1	26-09-2019
		BR 112019017008 A2	14-04-2020
		CA 3053592 A1	23-08-2018
		CL 2019002323 A1	06-12-2019
		CN 108456250 A	28-08-2018
		EP 3594241 A1	15-01-2020
		JP 7064663 B2	11-05-2022
		JP 2020508657 A	26-03-2020
		KR 20190127740 A	13-11-2019
		RU 2019128921 A	17-03-2021
		SG 11201907528T A	27-09-2019
		TW 201835106 A	01-10-2018
		US 2019359723 A1	28-11-2019